



ISSN 2228-7701

Online Journal of Animal and Feed Research



BOOKLET

Online Journal of Animal and Feed Research

An international peer-reviewed journal which publishes in electronic format (online)

Online J. Anim. Feed Res., 15 (1): 1-59; January, 2025

Editors-in-Chief

Habib Aghdam Shahryar, PhD, Professor of Animal Nutrition; Department of Animal Science, Islamic Azad University, Shabestar, **IRAN** ([Google Scholar](#), [SCOPUS](#), Email: ha_shahryar@yahoo.com)

Saeid Chekani Azar, PhD, Faculty of Veterinary Medicine, Animal Physiology, Atatürk University, **TURKEY** ([Google Scholar](#), [SCOPUS](#), [WoS Metrics](#), Email: saeid.azar@atauni.edu.tr)

Managing Editor

Alireza Lotfi, PhD, Animal Physiology, Islamic Azad University, **IRAN** ([Google Scholar](#), [SCOPUS](#), [ResearchGate](#), Email: arlotfi@gmail.com)

Section Editors

Arda Yildirim, PhD, Professor Dr., Department of Animal Science, Faculty of Agriculture, Gaziosmanpasa University, 60240 Tokat, **TURKEY** (Email: arda.yildirim@gop.edu.tr); Animal Science, Nutrition-non Ruminants, Breeding, Nutritive Value

Çağrı Kandemir, PhD, Assistant Professor, Institute of Science, Department of Animal Science, Ege University, Izmir, **TURKEY** ([Website](#); Email: cagri.kandemir@ege.edu.tr); Animal Science, Nutrition – Ruminants, Animal Reproduction

Ehsan Gharib Mombeni, DVM, PhD in Bacteriology; Department of Pathobiology, Shahid Chamran University of Ahvaz, **IRAN** (Emails: e.mombeni@hotmail.com; e-gharibmombeni@stu.scu.ac.ir)

İbrahim Çakmak, Prof. Dr., Director of Beekeeping Development Application and Research Center, Animal Science Department, Faculty of Agriculture, Bursa Uludag University, Gorukle Campus, Nilüfer, Bursa, **TURKEY** (E-mail: icakmak@uludag.edu.tr); Apiculture, Honey bee biology, genetics, behavior, health and products, pollination, beekeeping materials

John Cassius Moreki, PhD, Department of Animal Science and Production, College of Agriculture, **BOTSWANA** (Email: jcmoreki@gmail.com); Nutrition - Non-Ruminants, Breeders, Livestock management

Mohamed Shakal, Professor & Head of Poultry Diseases Department, Faculty of Veterinary Medicine, Cairo University, **EGYPT**; Director of the Endemic and Emerging Poultry Diseases Research Center, Cairo University, Shek Zaed Branch, EGYPT; Chairman of The Egyptian Poultry Forum Scientific Society. Representative for Egypt & Mena Region. (Email: shakal2000@gmail.com)

Muhammad Saeed, PhD, Northwest A&F University, Yangling, 712100, **CHINA** (Email: muhammad.saeed@nwsuaf.edu.cn), Nutrition – Ruminants

Language Editors

Mehrdad Ehsani-Zad, MA in TEFL, Takestan, Islamic Azad University, **IRAN** (Email: mehrdad_single2004@yahoo.com)

Samuel Stephen Oldershaw, Master of TESOL, The Humberston School & The Grimsby Institute, North East Lincolnshire, **UK** (Email: s.s.oldershaw@hotmail.com)

Statistical Editor

Ömer Eltas, PhD, Assist. Prof., Atatürk University, Faculty of Veterinary Medicine, Department of Biometry, Erzurum, **TURKEY** (Email; [omer.eltas@atauni.edu.tr](#)); Health Sciences, Veterinary Sciences, Zootechnical and Animal Feed, Biometry

Technical Editor

Alireza Lotfi, PhD, Animal Physiology, Islamic Azad University, **IRAN**

Editorial Team

Abdelfattah Y.M. Nour, DVM, PhD, Professor of Veterinary Physiology, Purdue University, **USA** (Email: nour@purdue.edu)

Adnan Yousaf, DVM, MPhil of Poultry Science (Gold Medalist), Ph.D. of Avian Embryology; Sindh Agricultural University Tandojam, **PAKISTAN** (E-mails: dr.adnan011@gmail.com; dr.adnan@salmanpoultry.com)

Ahmad Yildiz, PhD, Professor, Animal Science and Production Department, Faculty of Veterinary Medicine, Atatürk University, **TURKEY** (Email: ahmtstar@gmail.com); Nutrition – Ruminants

Ali Halajian, PhD, DVM, Professor of Parasitology, Department of Biodiversity, Faculty of Science and Agriculture, University of Limpopo, **SOUTH AFRICA** (Email: ali_hal572002@yahoo.com)

Ali Nobakht, PhD, Assistant Professor, Animal Science Department, Islamic Azad University, Maragheh, **IRAN** (Email: anobakht20@yahoo.com); Nutrition - Non-Ruminants

Alireza Radkhah, PhD, Department of Fisheries, Faculty of Natural Resources, University of Tehran, Karaj, **IRAN** (Email: alirezaradkhah@ut.ac.ir); Aquatic Biology, Aquaculture and Fisheries Biotechnology

Bahareh Hafezi, DVM, PhD Candidate for Veterinary Surgery, Ferdowsi University Veterinary, Mashhad, **IRAN** (Email: hafezibahareh@yahoo.com); Nutrition - Non-Ruminants: Small Animal and Poultry Internal Surgery

Ekrem Laçin, PhD, Professor of Animal Science, Faculty of Veterinary Medicine, Atatürk University, **TURKEY** (Email: ekremlacin@hotmail.com); Nutrition - Non-Ruminants

Erol Aydin, PhD, Professor Dr., Department of Animal Health Economics and Management, Faculty of Veterinary Medicine, Kafkas University, TR-36100 Kars, **TURKEY** ([Website](#), [Google Scholar](#), [SCOPUS](#), Email: dr-erolaydin@hotmail.com; ORCID: <https://orcid.org/0000-0001-8427-5658>);

Fazul Nabi Shar, PhD, Lecturer, Faculty of Veterinary & Animal Sciences, Lasbela University of Agriculture Water & Marine Sciences, Uthal Balochistan, **PAKISTAN** (Email: fazulnabishar@yahoo.com); Clinical Veterinary Medicine

Ferdous Mohd. Altaf Hossain, DVM, Sylhet Agricultural University, **BANGLADESH** (Email: ferdaus.dps@sau.ac.bd); Microbiology, Immunology, Poultry Science, and Public Health

Godadaw Misganaw, PHD; Department of Animal Science, College of Veterinary and Animal Sciences, University of Gondar, P.O.Box 196, Gondar, **ETHIOPIA** ([SCOPUS](#); Email: godadaw@gmail.com; ORCID: <https://orcid.org/0000-0001-5624-7983>); Nutrition - Ruminants

Hazim Jabbar Al-Daraji, PhD, Professor, University of Baghdad, College of Agriculture, Abu-Ghraib, Baghdad, **IRAQ** (Email: prof.hazimaldaraji@yahoo.com); Avian Reproduction and Physiology

Mohammed Yousuf Kurtu, Associate Professor, Animal Sciences Department, Haramaya University, Dire-Dawa, **ETHIOPIA** (Email: mkurtu2002@yahoo.com); Animal Science, Nutrition

Mohamed M. El-Deeb, PhD, Animal Nutrition Research Department, Animal Production Research Institute, Agriculture Research Center, Ministry of Agriculture, Dokki, Giza, **EGYPT**; Email: deep121eg@yahoo.com; ORCID

Moshood Adewale Belewu, PhD, Professor, Department of Animal Science, University of Ilorin, **NIGERIA** ([SCOPUS](#), [Google Scholar](#); Emails: mabel@unilorin.edu.ng; moshood.belewu@uniabuja.edu.ng); Nutrition - Ruminants; Dairy Science

Murat Genç, PhD, Associate Professor, Department of Animal Science and Production, Atatürk University, **TURKEY** ([Website](#); ocoban@atauni.edu.tr); Veterinary Sciences, Zootechnical and Animal Feed, Breeding - Ruminants

Nilüfer Sabuncuoğlu Çoban, PhD, Professor, Department of Animal Science and Production, Faculty of Veterinary Medicine, Atatürk University, **TURKEY** ([Website](#); Email: ncoban@atauni.edu.tr); Animal Hygiene and Welfare, Physiology

Ömer Çoban, PhD, Professor, Department of Animal Science and Production, Atatürk University, **TURKEY** ([Website](#); ocoban@atauni.edu.tr); Nutrition - Ruminants

Paola Roncada, PhD, Associate Professor, Veterinary Pharmacology and Toxicology, University of Bologna, **ITALY** (Email: paola.roncada@unibo.it); Pharmacokinetics

Raga Mohamed Elzaki Ali, PhD, Assistant Professor, Department of Rural Economics and Development, University of Gezira, **SUDAN** (Email: ragaalzaki@yahoo.co.uk); Animal-feed interactions, Nutritive value

Rashid Habiballa Osman, PhD, Assistant Prof., in Department of Poultry Production, Faculty of Animal Production, West Kordofan University, **SUDAN** (E-mail: rashid@wku.edu.sd); Nutrition - Non-Ruminants

Raziye Raeesi, PhD student in Fisheries Engineering, Gorgan University of Agricultural Sciences and Natural Resources, Gorgan, **IRAN** (Email: r.raeesi2012@yahoo.com); Aquaculture, Fish nutrition

Sesotya Raka Pambuka, MSc, Sinta Prima Feedmill, Poultry and Aqua Feed Formulation, Sulaiman Rd 27A, West Jakarta, **INDONESIA**

Shigdaf Mekuriaw, Andassa Livestock research center, **ETHIOPIA** (Email: shiqdafmekuriaw@yahoo.com); Animal production and Nutrition

Terry Ansah, PhD, University for Development Studies-Ghana and Harper Adams University College, **UK** (Email: ansahterry@yahoo.com); Nutrition - Ruminants

Tohid Vahdatpour, PhD, Assistant Professor, Department of Physiology, Islamic Azad University, Shabestar, **IRAN** ([Scopus](#); [Google Scholar](#); Emails: vahdatpour@iaushab.ac.ir; tvahdatpour@gmail.com); Physiology and Functional Biology of Systems

Vassilis Papatsiros, PhD, Department of Porcine Medicine, University of Thessaly, Trikalon str 224, GR 43100, **GREECE** (Email: vpapatsiros@yahoo.com); Dietary input, Animal and Feed interactions

Wafaa Abd El-Ghany Abd El-Ghany, PhD, Associate Professor, Poultry and Rabbit Diseases Department, Cairo University, Giza, **EGYPT** (Email: wafaa.ghany@yahoo.com); Poultry and Rabbit Diseases

Wesley Lyevertton Correia Ribeiro, MSc, DVM, College of Veterinary, Medicine, State University of Ceará, Av. Paranjana, 1700, Fortaleza, **BRAZIL** (Email: wesleylyevertton@yahoo.com.br); Animal Health and Welfare, Veterinary Parasitology

Yavuz Gurbuz, Professor, University of Kahramanmaraş Sutcu Imam, Department of Animal Nutrition, Campus of Avsar, Kahramanmaraş, **TURKEY** (Email: yavuzgurbuz33@gmail.com); Animal Nutrition, Feed Technology and Evaluation

Yonas Gizaw Habtemichae, DVM, MVSc; Jigjiga University, College of Veterinary Medicine, P.O.Box.1020 Jigjiga, **ETHIOPIA** (Email: yonasg5@gmail.com; ORCID: 0000-0003-4208-5682)

Advisory Board

Alireza Ahmadzadeh, PhD, Assistant Professor, Department of Animal Science, Islamic Azad University, Shabestar, **IRAN** (Emails: a.r.ahmadzadeh@gmail.com; ahmadzadeh@iaushab.ac.ir); Biometry - Plant Breeding (Biotechnology)

Daryoush Babazadeh; DVM, DVSc, PhD of Avian/Poultry Diseases, School of Veterinary Medicine, Shiraz University, Shiraz, **IRAN** (Scopus; ORCID ID; Publons; Full Member of WAME; Member of IAVE; Email: daryoush.babazadeh@shirazu.ac.ir)

Fikret Çelebi, PhD, Professor of Physiology, Faculty of Veterinary Medicine, Atatürk University, Erzurum, **TURKEY** (Email: fncelebi@atauni.edu.tr); Physiology and Functional Biology of Systems

Mohamed Shakal, Professor, Poultry Diseases Department, Faculty of Veterinary Medicine, Cairo University, **EGYPT**; Director of the Endemic and Emerging Poultry Diseases Research Center, Cairo University, Shek Zaed Branch, **EGYPT**; Chairman of The Egyptian Poultry Forum Scientific Society. REPRESENTATIVE FOR EGYPT & MENA REGION. Email: shakal2000@gmail.com

Naser Maheri Sis, PhD, Assistant Professor, Dept. Anim. Sci., Islamic Azad University, Shabestar, **IRAN** (Website; Emails: maherisis@iaushab.ac.ir; nama1349@gmail.com); Nutrition - Ruminants, Nutritive Value, Utilization of Feeds

Join OJAFR Team

Online Journal of Animal and Feed Research is always striving to add diversity to our editorial board and staff. Applicants who have previous experience relevant to the position they are applying for may be considered for more senior positions within OJAFR. All applicants should begin as section reviewers before progressing on to more senior roles. Editorial board members do not receive any remuneration unless in overtime working conditions. These positions are voluntary. If you are currently graduated from MSc, or PhD at university and interested in working for OJAFR, please fill out the application form below. Once your filled application form is submitted, the editorial board of the journal will review your request and inform you within a week of their decision for membership in the editorial board. The list of the editorial board will be updated yearly and the new members will be listed each year. If you are a PhD, assistant, associate professor, distinguished professor, or an active researcher, please send us a copy of your resume (CV) and your ORCID ID. You should briefly express any leadership positions, editorial or publishing activities, and other experiences you have had that are relevant to applied research, conducted studies, and published articles. Also, the volunteer editor/reviewer should declare if he/she has any conflict of interest for joining the journal editorial board in requesting time, and also during his/her activity as an editor or reviewer.

If you would like to represent the OJAFR at your university, join our volunteer staff today! OJAFR representatives can include any assistant students, teachers, instructors, researchers, and professors at university or international institutes. You can also register as a member of the journal for subsequent contacts by email and or invitation for joining educational webinars.

Editors affiliated with the *Online Journal of Animal and Feed Research* who are also serving on the editorial boards of other journals sharing similar goals and scope are expected to adhere to the policies of the *Online Journal of Animal and Feed Research* while they are involved in editorial responsibilities at the *Online Journal of Animal and Feed Research*. For such editors, it is important to declare any potential conflict of interest transparently. If at any stage of the journal's peer review process, it becomes apparent that a submitted article is under consideration in a journal where our editor also serves, the *Online Journal of Animal and Feed Research* immediately reassigns the article to another editor. Similarly, if such a situation involves the editor-in-chief, and the editor-in-chief collaborates with another journal, the responsibility for handling that article is delegated to the second editor-in-chief/associate editor-in-chief/handling editor/managing editor. In case of a conflict of interest between the editor-in-chief and any of the mentioned roles, the article will be handled by one of the editorial board members.

Download [OJAFR Application Form](#)

Volume 15 (1); January 30, 2025

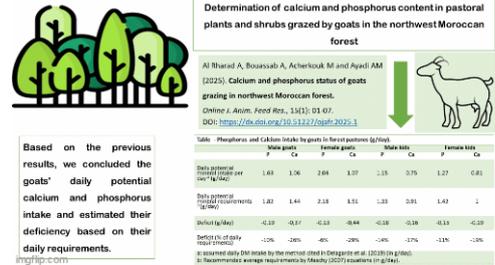
Research Paper

Calcium and phosphorus status of goats grazing in northwest Moroccan forest

Al Rharad A, Bouassab A, Acherkouk M and Ayadi M.
Online J. Anim. Feed Res., 15(1): 01-07, 2025; pii: S222877012500001-15
 DOI: <https://dx.doi.org/10.51227/ojafr.2025.1>

Abstract

In the present study, leaves and twigs from 17 shrubs and trees consumed by the west-north Moroccan indigenous goats were collected and evaluated for their calcium and phosphorus (Ca and P) content. The potential mineral needs of adults and young goats of both sexes (male and female) from three localities were estimated to assess their mineral deficiency. This assessment was based on their weight and the diet composition determined through direct observation and the bites method. The browse species had a higher Ca content than P (1.79 vs 1.57 g/kg DM). The adult female goats had the highest P intake (2.04 g/day) with the highest deficit compared to the male adult (-29 vs -26) % of their daily requirements. Young kids (males and females) had the lowest Ca intake (0.81 and 0.75 g/day, respectively) and recorded the lowest deficit (-17 vs -19) %, respectively. Goats also showed a higher Ca deficit than P. In conclusion, the present results offer valuable information about the main mineral intake of the goats in the forest pasture of this region. Supplementing these two minerals is essential for enhancing goat performance in the traditional semi-extensive goat farming system that relies on forest pastures in the western-northern region of Morocco.



Keywords: Diet composition, Goat, Indigenous breeds, Mineral requirement, Pastoral plants.

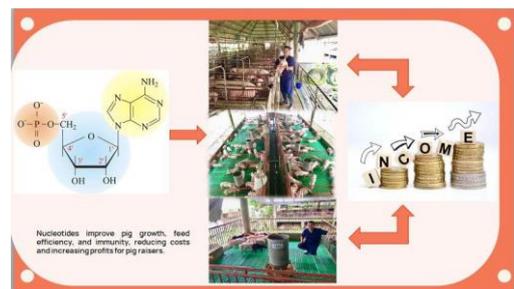
[Full text-PDF]

Growth performance and profitability of weaning pigs (*Sus scrofa domestica* L.) fed pre-starter diet supplemented with nucleotide

Verdijo A and Mondejar H.
Online J. Anim. Feed Res., 15(1): 08-14, 2025; pii: S222877012500002-15
 DOI: <https://dx.doi.org/10.51227/ojafr.2025.2>

Abstract

Nucleotides can improve intestinal health by modulating the local immune response and intestinal mucosa development in weaned piglets. This study was conducted to evaluate the growth performance of post weaned piglets and evaluated the economic analysis of nucleotides supplementation for 30 days. A total of 120 mixed breed piglets were selected at the weaning stage and were used in the experiment as control group vs. treatment 2 supplemented with nucleotide. Each treatment consisted of 60 heads with three replications with 20 heads per replication arranged in Complete Randomized Design. Results were analyzed through Pairwise T and LSD tests. In terms of growth performance, results showed that supplementation of nucleotide had significantly increased the average daily gain, feed conversion ratio and weight gain by 0.50, 1.20 and 15.02 kg, respectively. However, there was no significant difference in terms of the average daily feed intake. With regards to the economic analysis, total production input had no effect with or without nucleotide supplementation but surprisingly, it had a gross margin of Php 95,900 (Philippine peso money) which was 5% more as with those that have supplementation. As to the net income, supplementation of nucleotides increased about 40.7% in comparison to control. Furthermore, a peso of investment could have a return of about 18 cents (1050 Php) more returns with supplementation, which apparently had 0.11 cents leverage compared to control group (0.07 cents). In conclusion, nucleotide supplementation not only improved the growth performance of post-weaned piglets but also enhanced profitability, offering a significant return on investment for swine producers. This makes nucleotide supplementation a promising strategy for improving both animal health and economic outcomes in swine production.



Keywords: Benefit Cost Ratio, Daily weight gain, Feed conversion ratio, Net Income, Nucleotides.

[Full text-PDF]

Research Paper

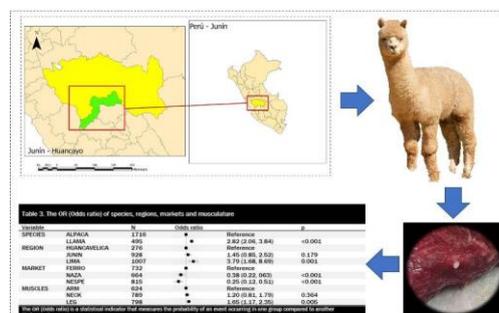
Sarcocystosis in alpacas and llamas: regional, market, and muscle-specific prevalence patterns

Garcia-Olarte E, Ninahuanca J, Suarez-Reynoso W, Mauricio-Ramos Y, Guillen MF, Payano IU, Tacza AA, and Condor WG.

Online J. Anim. Feed Res., 15(1): 15-20, 2025; pii: S222877012500003-15
DOI: <https://dx.doi.org/10.51227/ojaf.2025.3>

Abstract

The objective of this study was to determine the effect of species (alpacas and llamas), markets in the city of Huancayo (Ferrocarril Commercial Center, Nueva Esperanza, and Nazareth), and muscle groups on the prevalence of *Sarcocystis* sp. Between January and October 2023, a total of 2,211 carcasses were inspected, comprising 1,716 alpacas and 495 llamas. The results indicated a prevalence of 21% (104/495 carcasses) in llamas and 8% (138/1,716 carcasses) in alpacas. By region of origin, the prevalence in alpacas was reported as follows: Huancavelica (7.7%) with 14/181 carcasses, Junín (6.7%) with 55/820 carcasses, and Lima (9.7%) with 69/715 carcasses. For llamas, the Lima region exhibited the highest prevalence of sarcocystosis (33.9%) with 72/212 carcasses, followed by Huancavelica (14.7%) with 14/102 carcasses, and Junín (9.8%) with 18/181 carcasses. Regarding the markets, the Ferrocarri market presented the highest risk of contamination, serving as the reference group for comparison. In contrast, the Nazareth and Nueva Esperanza markets showed significantly lower odds of *Sarcocystis* sp. presence, with Odds Ratios (ORs) of 0.38 and 0.25, respectively. For muscle groups, the anatomical distribution of *Sarcocystis* sp. cysts revealed a preferential localization in the leg (OR = 1.65) and neck (OR = 1.20) compared to the shoulder. This investigation provides significant data on the prevalence of *Sarcocystis* sp. in alpacas and llamas, highlighting a higher prevalence in llamas despite their smaller sample size. These findings emphasize the need for targeted interventions to address this parasitic infection in camelid production systems.



Keywords: Animal products, Camelids, Carcass quality, Mantaro valley, Parasite.

[Full text-PDF]

Research Paper

Influence of feather genotype, storage duration and temperature on the external and internal qualities of chicken table eggs

Kanasuah DN, Adomako K, Hagan BA and Olympio OS.

Online J. Anim. Feed Res., 15(1): 21-32, 2025; pii: S222877012500004-15
DOI: <https://dx.doi.org/10.51227/ojaf.2025.4>

Abstract

A study was carried out to determine the influence of the feather genotype, storage duration, temperature and method on the internal and external qualities of chicken table eggs. A total of 864 table eggs collected from naked neck (Nanaff), frizzle (nanaff) and normal feathered (nanaff) birds were used in the study. A Completely Randomized Design of four factors namely, feather genotypes, storage temperatures (5°C and 26°C), storage duration (0, 7, 14, 21 and 28 days) and storage methods (with or without vegetable oil application) was used. The GLM procedure of GenStat (17th Edition) was used to determine the effects of the four factors and their interactions on external qualities (egg weight, length, and width, shell weight and thickness) and internal qualities (albumen height and weight, yolk height, weight, diameter and colour and Haugh unit) of table eggs. The effect of chicken genotype on proximate composition and nutritional values of table eggs were also determined. Feather genotype had significant ($P<0.05$) effect on yolk colour and weight whilst storage duration, temperature and method had significant ($P<0.05$) effects on all the internal qualities of eggs studied except effect of storage duration on yolk colour. The 2-way and 3-way interactions of the factors studied were important sources of variation for many of the internal qualities of eggs studied. With the exception of storage temperature, the other factors studied had significant ($P<0.05$) effects on many of the external qualities of eggs. The interactions of the factors were not significant ($P>0.05$) sources of variation for most of the external qualities of eggs. Mutant feather genes (Na and F) positively influence egg qualities which could be utilised to segment the commercial chicken egg market.



Keywords: Feather, Frizzle, Naked neck, Nutritional value, Yolk colour,.

[Full text-PDF]

Primal cuts of carcass and meat characteristics of Kacang goat fed total mixed ration containing different sources of ruminally undegraded protein

Adiwinarti R, Kustantinah, Rusman, Rianto E, Purnomoadi A, Arifin M, Sutaryo, and Restitrisnani V.

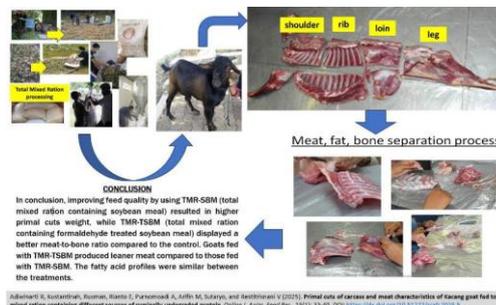
Online J. Anim. Feed Res., 15(1): 33-40, 2025; pii: S222877012500005-15
DOI: <https://dx.doi.org/10.51227/ojaf.2025.5>

Abstract

This study was designed to evaluate the effect of feed quality improvement using gliricidia and different sources of protein in total mixed ration (TMR) on the primal cuts, loin eye area, and fatty acids profile of goat meat. This study used twenty yearling Kacang goats weighing 17.42 ± 1.63 kg. The goats were randomly allocated into 4 different treatments in a completely randomized design. The treatments involved the use of natural grass from rangeland (NGFR; control) as well as improving the quality of feed through TMR containing various ruminally undegraded protein sources, i.e. TMR contains fish meal (TMR-FM), TMR contains soybean meal (TMR-SBM) and TMR contains formaldehyde treated soybean meal (TMR-TSBM). The parameters observed were primal cuts yield, loin eye area, meat, fat, bone of primal cuts, and fatty acids profile. Data were analyzed using a one-way analysis of variance. The results showed that the goats fed TMR-FM and TMR-TSBM produced significantly higher meat percentage than control goats. The meat yield of TMR-SBM and TMR-TSBM goats were significantly higher than those of control goats. Goats fed TMR-SBM produced the highest primal cuts yield and shoulder weight, while the weight of rib, loin, and leg of TMR-SBM goats were similar to those of TMR-TSBM goats. Loin eye area was similar between the treatments. Saturated fatty acids content in TMR groups was similar to those in control. It can be concluded that improved feed quality using TMR-SBM produced significantly higher primal cuts weight, while TMR-TSBM had better meat-to-bone ratio than control. TMR-TSBM goats produced significantly leaner meat than TMR-SBM goats. Fatty acid profiles were similar between treatments.

Keywords: Fatty acids, Fish meal, Goat meat, Meat quality, Total mixed ration.

[Full text-PDF]



Short Communication

Hematological and biochemical parameters of captive fallow deer (*Dama dama*) in a zoo environment

Hažimusić N, Škapur V, Hadžijunuzović-Alagić D, and Livnjak A.

Online J. Anim. Feed Res., 15(1): 41-46, 2025; pii: S222877012500006-15
DOI: <https://dx.doi.org/10.51227/ojaf.2025.6>

Abstract

Accurate health assessment of wild, semi-captive, or domesticated animals is essential for their well-being. Despite this necessity, limited studies have been conducted on deer species, and there is a paucity of information on the hemato-biochemical parameters of different deer species globally. Present study aimed to fill this gap by determining the hematological and serum biochemical parameters of fallow deer (*Dama dama*) maintained in semi-captivity within zoo environments for the first time in Bosnia and Herzegovina. Present research involved six healthy male fallow deer, aged 2 to 5 years. The deer were immobilized using xylazine hydrochloride and ketamine hydrochloride, and blood samples were collected from the external jugular vein. The hematological parameters measured included RBC, PCV, HGB, MCV, MCH, MCHC, RDW, RETIC, WBC, WBC differential, PLT, MPV, PDW, and PCT. Biochemical parameters included glucose, urea, creatinine, albumin, triglycerides, cholesterol, and enzymes (AST, ALT, ALKP, and GGT) activities. The results showed the higher glucose and urea concentrations and the same values for creatinine, triglycerides, and enzyme activities when compared to some previous reports. These findings highlighted the importance of considering handling methods and environmental conditions when interpreting biochemical parameters, contributing to improved health assessments and management practices for deer in captivity.

Keywords: Biochemical and hematological parameters, Captive wildlife, Domesticated animals, Fallow deer.

[Full text-PDF]



Effect of graded levels of dietary tomato waste on performance and carcass characteristics of Japanese quail reared under intensive system

Bhawa S, Moreki JC and Manyeula F.

Online J. Anim. Feed Res., 15(1): 47-59, 2025; pii: S222877012500007-15
DOI: <https://dx.doi.org/10.51227/ojafnr.2025.7>

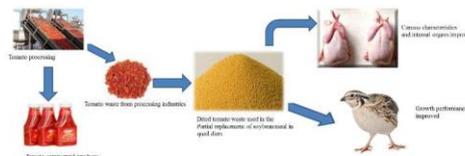
Abstract

This study was carried out to evaluate the effects of partial replacement of soybean meal (SM) with tomato waste (TW) in Japanese quail diets on the resulting yield, internal organs, and carcass characteristics. Eighty unsexed 1-day-old chicks were housed in battery cages with cardboard boxes used as solid floors and randomly assigned to 1 of 4 dietary groups, 46.2% SM, 44.2% SM + 2% TW, 42.2% SM + 4% TW, or 40.2% SM + 6% TW, over a 6 weeks growth period. Yields and carcass characteristics were then determined. Data were analysed using the General Linear Model (GLM) procedures followed by a response procedure for surface regression analysis (Proc RSREG; SAS 9.4) to describe the parameters' responses to graded levels of dietary tomato waste. Repeated measures analyses showed significant week × diet interaction effects on feed intake (FI, $P = 0.03$), body weight gain (WG, $P = 0.0006$), feed conversion ratio (FCR, $P = 0.002$), protein efficient ratio ($P = 0.0001$), and growth efficiency ($P = 0.0001$). By supplementing the diets of quails with a 2% inclusion level, a diet significantly affected quails' FI on weeks 1, 2, 3, and 6. A diet containing 2% TW significantly affected live weight (LW), hot carcass weight (HCW), and cold-dressed weight (CDW). It is concluded that the diet supplementation with 44.2% SM + 2% TW seemed ideal for optimum performance in Japanese quails based on the insignificant change in feed intake and growth efficiency results compared to 46.2% SM for weeks 1 and 2. Further research is needed on the application method that could be used to enhance the utilization of tomato waste in Japanese quails.

Keywords: Carcass characteristics, Dietary replacements, Growth performance, Japanese quails, Tomato waste.

[Full text-[PDF](#)]

PARTIAL REPLACEMENT OF SOYBEAN MEAL WITH TOMATO WASTE IN JAPANESE QUAIL DIETS: GROWTH PERFORMANCE, INTERNAL ORGANS, AND CARCASS CHARACTERISTICS



Bhawa S, Moreki JC and Manyeula F (2025). Effect of graded levels of dietary tomato waste on performance and carcass characteristics of Japanese quail reared under intensive system. Online J. Anim. Feed Res., 15(1): 47-59, 2025. DOI: <https://dx.doi.org/10.51227/ojafnr.2025.7>

Online Journal of Animal and Feed Research



ISSN 2228-7701

ISSN: 2228-7701

Frequency: Bimonthly

Current Issue: 2025, Vol: 15, No: 1 (January)

DOI Prefix: [10.51227](https://doi.org/10.51227)

Publisher: [SCIENCLINE](https://www.science-line.com)

Online Journal of Animal and Feed Research is an international peer-reviewed journal, publishes the full text of original scientific researches, reviews, and case reports in all fields of animal and feed sciences, bimonthly and freely on the internet [...view full aims and scope](#)

www.ojafr.ir and www.ojafr.com

» OJAfr indexed/covered by [Scopus](#), [AGRIS](#), [EBSCO](#), [Ulrich's™](#), [HINARI](#), [NSD](#), [AKSTEM](#), [BASE](#), [ZDB](#), [ICV](#), [EZB](#) [...details](#)

Journal metrics: [h5-index=9](#); [h5-median=12](#)

» Full texts and XML articles are available in [Crossref](#) and [AGRIS](#).

» Digital Archiving: [Journal Repository \(eprints\)](#)

» This journal is in full compliance with [BOAI](#) and [ICMJE's Recommendations](#).

» High visibility of articles over the internet.

» Publication Ethics and Policies [...details](#)

» High visibility of articles over the internet through [Gold Open Access](#).

» Publisher Item Identifier [...details](#)

» This journal encourage the academic institutions in low-income countries to publish high quality scientific results, free of charges... [Peer Review Process](#)



ICMJE INTERNATIONAL COMMITTEE of MEDICAL JOURNAL EDITORS



Scienceline Publication, Ltd.

Ömer Nasuhi Bilmen Road, Dönmez Apart., G Block, No:1/6, Yakutiye, Erzurum/25100, TURKEY

Phone: +90 538 770 8824 (TURKEY)

Homepage: www.science-line.com

Emails: administrator@science-line.com; saeid.azar@atauni.edu.tr

[ABOUT US](#)

| [CONTACT US](#)

| [PRIVACY POLICY](#)

CALCIUM AND PHOSPHORUS STATUS OF GOATS GRAZING IN NORTHWEST MOROCCAN FOREST

Asma AL RHARAD^{1,2} , Abderrahman BOUASSAB² , Mohamed ACHERKOUK¹  and Mohammed AYADI¹ 

¹Regional Centre of Agricultural Research of Tangier, National Institute of Agricultural Research, Avenue Ennasr, BP 415 Rabat Principale, 10090 Rabat, Morocco

²Laboratory of Physical-Chemistry of Materials, Natural Substances and Environment, Department of Chemistry, Faculty of Science and Technology, Abdelmalek Essâadi University, Tangier, 90010, Morocco

✉Email: asma.alrharad@etu.uae.ac.ma

↳Supporting Information

ABSTRACT: In the present study, leaves and twigs from 17 shrubs and trees consumed by the west-north Moroccan indigenous goats were collected and evaluated for their calcium and phosphorus (Ca and P) content. The potential mineral needs of adults and young goats of both sexes (male and female) from three localities were estimated to assess their mineral deficiency. This assessment was based on their weight and the diet composition determined through direct observation and the bites method. The browse species had a higher Ca content than P (1.79 vs 1.57 g/kg DM). The adult female goats had the highest P intake (2.04 g/day) with the highest deficit compared to the male adult (-29 vs -26) % of their daily requirements. Young kids (males and females) had the lowest Ca intake (0.81 and 0.75 g/day, respectively) and recorded the lowest deficit (-17 vs -19) %, respectively. Goats also showed a higher Ca deficit than P. In conclusion, the present results offer valuable information about the main mineral intake of the goats in the forest pasture of this region. Supplementing these two minerals is essential for enhancing goat performance in the traditional semi-extensive goat farming system that relies on forest pastures in the western-northern region of Morocco.

Keywords: Diet composition, Goat, Indigenous breeds, Mineral requirement, Pastoral plants.

INTRODUCTION

Calcium (Ca) and phosphorus (P) play pivotal roles in the productivity and health of small ruminants, particularly goats, as they are essential macrominerals for bone development, metabolic processes, and overall performance. Adequate levels of these minerals are critical for growth, reproduction, lactation, and maintaining physiological functions, including energy metabolism and enzymatic activity. Deficiencies in Ca and P can lead to reduced productivity, poor skeletal development, and metabolic disorders, highlighting the importance of understanding their availability and contribution to grazing systems (Drogoul et al., 2004).

In Morocco's northern region, recognized as the most forested area in the country with a woodland rate of 26% (MAPMDREF, 2018), forest pastures serve as a vital year-round feed source for grazing goats. Trees and shrubs, the primary feed sources in this region, provide energy, protein, and potentially essential minerals to support the nutritional needs of goats. These forest ecosystems underpin traditional smallholder livestock production systems, where farmers rely heavily on grazing goats (Chebli et al., 2021; Ayadi et al., 2022; Chebli et al., 2022a, 2022b).

While numerous studies have explored goat feeding behavior in Mediterranean regions (Glasser et al., 2012; Manousidis et al., 2016, and 2018), research on forage mineral content, mainly Ca and P, remains limited. Most evaluations focus on the chemical composition of forage, such as protein and energy, with insufficient attention to the quantitative assessment of mineral contributions to animal productivity (Chebli et al., 2022c; Jimenez et al., 2024). Understanding the mineral content of forage and its contribution to goat requirements is essential for improving feeding and grazing strategies and enhancing the sustainability of livestock production systems.

This study aims to estimate the Ca and P contents in pastoral shrubs and trees and evaluate their potential intake by four goat categories grazing in forest pastures in the western-northern region of Morocco. This information is crucial for addressing potential mineral deficiencies and optimizing the productivity and sustainability of goat farming in the region.

MATERIALS AND METHODS

Study area

This study was conducted in the Chefchaouen region, specifically in the Talassemrane high mountains. This region is characterized by local goat breeding. The herds are managed in an extensive system on natural forest pastures, from which the animals obtain over 90% of their nutritional needs, which the breeders consider "free" feed. The climate of the

RESEARCH ARTICLE
 PII: S222877012500001-15
 Received: October 04, 2024
 Revised: January 10, 2025
 Accepted: January 11, 2025

study area is Mediterranean, with dry summers. The mean annual temperature is 18.6°C, with an average rainfall of around 640 mm yearly (Acherkouk et al., 2022).

Animals and feeding behaviors

The study was conducted in the autumn and winter of 2022. It took place in three localities to represent the Chefchaouen region: Bouhalla (35°6'0" N and 5°6'36" W), Chrafat (35°4'60" N and 5°6'8" W), and Kalaâ (35°3'50" N and 4°31'46" W), with one breeder by locality. A group of 21 indigenous goats of each category (male adult, female adult, male kids, female kids) was chosen from each breeder. Each group within each category had approximately the same age and similar body weight (Table 1). The goats grazed on natural pasture for about seven hours daily when conditions permitted.

Diet selection

For each goat category, we were interested in two key parameters: (i) their daily dry matter intake (DDMI) on forest pasture and (ii) their weight. DDMI (Table 2) was deduced by the difference in the goat's weight before being released for grazing and upon their return in the evening, taking into account excretion-related losses (Delagarde et al., 2019). The importance of grazed pastoral species in the four categories of animal diets (Table 3) was assessed by direct observation of the animals during their grazing in the cold season during the study period and by the bite method described by Meuret et al. (1985).

Table 1 - The age and body weight of the chosen goats

Animal categories	Heads/category number	Age (months)	Mean body weight (kg)
Male adult	7	24-30	36±9.07
Female adult	7	24	33±9.81
Male kids	7	9	17±1.02
Female Kids	7	9- 12	18±6.56

Table 2 - The goat's daily dry matter intake in the three localities

Animal category	Daily dry matter Intake (g)		
	Breeder 1	Breeder 2	Breeder 3
Male adult	1500	1500	1800
Female adult	1500	2000	2500
Male kids	1200	1000	1200
Female Kids	1000	1000	1200

Table 3 - Qualitative and quantitative diet composition of the goats

Breeder 1		Breeder 2		Breeder 3	
Species	Percentage (%)	Species	Percentage (%)	Species	Percentage (%)
<i>Quercus ilex</i>	67	<i>Quercus ilex</i>	50	<i>Quercus ilex</i>	80
<i>Quercus ilex fruit</i>	22	<i>Quercus ilex fruit</i>	20	<i>Pistacia lentiscus</i>	10
<i>Phillyrea media</i>	5	<i>Phillyrea media</i>	15	<i>Cistus albidus</i>	5
<i>Cistus albidus</i>	4	<i>Arbutus unedo</i>	5	Grass plants	5
Grass plants	2	Grass plants	10		

The mineral content of the browse species grazed by the goat in forest pasture

17 collected samples of shrubs and tree species from forest pastures in the mountainous region of northwest Morocco were collected to determine their Ca and P content. The collected samples were dried in an oven until a constant weight at 40°C temperature. Then, the dried samples were milled with a sieve mesh size of 1 mm to evaluate their Ca and P content. The analyses were conducted on triplicate from each species, which were subjected to dry ashing at 600 °C for 4 hours and then prepared for mineral analysis using the wet ashing (HCl-HNO3) procedure. The Ca content analysis was performed using the manganometric method according to AOAC (1997), and for P determination, the nitrovanadomolybdate technique was employed (AOAC, 1997).

Data analysis

The Ca and P content were analyzed using a general linear model (GLM) with the SAS software 9.4 version. To determine the variability between forage species, the variance of the means was analyzed according to a one-factor variation model $Y_{ij} = \mu + T_i + e_{ij}$ with μ : overall mean, T_i : forage species, and e_i : residual error. Differences between mean values were tested using the LSD "Last Square Deviation" test.

Estimation of the macromineral deficit of the goats

A) The macromineral requirement for goats

The primary mineral (Ca and P) requirements for goats are expressed in absorbable elements obtained by the equations of Meschy (2007) (1 and 2) based on the dry matter intake (DMI) in kg/day and body weight (BW) in kg. They are expressed by g per day.

$$P \text{ abs} = 0.905 \text{ DMI} + 0.3 + 0.002 \text{ BW} \quad (1)$$

$$Ca \text{ abs} = 0.67 \text{ DMI} + 0.01 \text{ BW} \quad (2)$$

B) The mineral deficit of the goat's diet

With the data collected previously (daily feed intake, diet composition, and Ca and P requirement), the potential mineral intake was calculated based on their daily dry matter intake and Ca and P absorbable of the feed using the real absorption coefficients (RAC) for Ca and P by Meschy (2007). Then, we define the deficit by subtracting the intakes from the calculated needs based on each animal's body weight. Finally, the deficit is estimated as a percentage of the requirements.

RESULTS

Mineral content of pastoral fodder

The mineral content of the pastoral fodder selected by indigenous grazing goats is presented in Table 4. The P concentration ranged from 0.99 ± 0.07 to 2.30 ± 0.01 g/kg DM, which varies significantly between the forage species ($P < 0.001$). The pastoral species with the highest P content are *Olea europaea* L. (2.30 g/kg DM), *Rosmarinus officinalis* L. (2.25 g/kg DM), *Ceratonia siliqua* L., and *Erica arborea* L. (1.93 and 1.91 g/kg DM, respectively). The other species' content varies between 1.78 and 0.99 g/kg DM. Moreover, the average Ca content of the selected plants is 1.79 g/kg DM. *Erica arborea* L. and *Genista scorpius* L. showed the lowest content (0.86 and 0.75 g/kg DM, respectively). At the same time, the highest values were observed in *Olea europaea* L., *Arbutus unedo* L., and grass plants (2.63, 2.61, and 2.42 g/kg DM, respectively). The Ca and P ratio oscillates between 0.45 and 2.39 (Table 4). The higher ratios were observed in grass plants (2.39), *Cistus ladanifer* L. (1.88), and *Arbutus unedo* L. (1.71). In contrast, *Vaccinium myrtillus* L., *Genista scorpius* L., and *Erica arborea* L. showed lower ratios (0.62, 0.58, and 0.45, respectively).

Table 1 - Average content of macrominerals in species browsed by goats in forest pasture of Northwest Morocco (g/kg DM)

Pastoral species	Phosphorus (g/kg DM)	Calcium (g/kg DM)	Ca:P ratio
<i>Olea europaea</i> L. (Olive tree)	2.30 ^a	2.63 ^a	1.14 ^{fg}
<i>Cistus albidus</i> L. (White-Leaf Rockrose)	1.78 ^c	2.17 ^{cd}	1.21 ^{fg}
<i>Cistus ladanifer</i> L. (Gum Rockrose)	1.02 ^j	1.92 ^e	1.88 ^b
<i>Quercus canariensis</i> (Algerian Oak)	1.15 ⁱ	1.35 ^{fg}	1.17 ^{fg}
<i>Phillyrea media</i> L. (Mock privet)	1.41 ^g	2.31 ^{bcd}	1.64 ^{cd}
<i>Quercus ilex</i> (Holm Oak)	1.44 ^{fg}	1.91 ^e	1.32 ^{ef}
<i>Quercus ilex</i> (Fruit)	1.64 ^{de}	1.18 ^{gh}	0.72 ^{jk}
<i>Arbutus unedo</i> L. (Strawberry tree)	1.53 ^{ef}	2.61 ^a	1.71 ^{bc}
<i>Pistacia lentiscus</i> L. (Mastic tree)	1.60 ^{de}	2.37 ^{bc}	1.48 ^{de}
<i>Ceratonia siliqua</i> L. (Carob tree)	1.93 ^b	2.19 ^{bcd}	1.14 ^{fg}
<i>Erica arborea</i> L. (White heather)	1.91 ^b	0.86 ^{ij}	0.45 ^l
<i>Quercus suber</i> L. (Cork oak)	1.68 ^{cd}	1.46 ^f	0.87 ^{ij}
<i>Genista scorpius</i> L. (Mediterranean broom)	1.30 ^h	0.75 ⁱ	0.58 ^{kl}
<i>Vaccinium myrtillus</i> L. (European blueberry)	1.78 ^c	1.10 ^h	0.62 ^{kl}
<i>Lavandula stoechas</i> L. (Butterfly lavender)	0.99 ^j	1.06 ^{hi}	1.07 ^{gh}
<i>Rosmarinus officinalis</i> L. (Rosemary)	2.25 ^a	2.09 ^{de}	0.93 ^{hi}
Grass plants	1.01 ^j	2.42 ^{ab}	2.39 ^a
Mean	1.57	1.79	1.20
SEM	0.06	0.09	0.07
Probability (P) value	<0.0001	<0.0001	<0.0001

^{a,b,c}: Mean values in the same row with different letters are significantly different. DM: Dry Matter SEM: Standard error of the means

Macro mineral balance of goats

Based on the previous results, we derived the Ca and P intake (Table 5). The female goat's intake from forest pasture resulted in a moderately high average daily P intake of approximately 2.04 g/day, representing a deficit of about 6% of requirements. As for Ca, the forage intake was 1.07 g/day, representing a deficit of approximately 29% of requirements. On the other hand, the intake by male goats from forest pastures resulted in an average daily P intake of 1.63 g/day (i.e., a deficit of about 10% of requirements). Regarding Ca, the male goat's intake from forest pastures was about 1.06 g/day (i.e., a deficit of about 26% of requirements). The male kids' daily absorbable P intake was 1.15 g/day, corresponding to a 14% deficit of requirements. In contrast, female kids had an intake of 1.27 g/day of absorbable P (i.e., a deficit of 11% of requirements) and an intake of 0.81 g/day of absorbable Ca, reflecting a 19 % deficit of requirements compared to an intake of 0.75 g/day in absorbable Ca for the male kids (i.e., a deficit of 17% of requirements).

Table 5 - Phosphorus and Calcium intake by goats in forest pastures (g/day).

Mineral intake and deficit	Male goats		Female goats		Male kids		Female kids	
	P	Ca	P	Ca	P	Ca	P	Ca
Daily potential mineral intake per day ^a (g/day)	1.63	1.06	2.04	1.07	1.15	0.75	1.27	0.81
Daily potential mineral requirements ^b (g/day)	1.82	1.44	2.18	1.51	1.33	0.91	1.42	1
Deficit (g/day)	-0.19	-0.37	-0.13	-0.44	-0.18	-0.16	-0.15	-0.19
Deficit (% of daily requirements)	-10%	-26%	-6%	-29%	-14%	-17%	-11%	-19%

a: assumed daily DM intake by the method cited in Delagarde et al. (2019) (in g/day). b: Recommended average requirements by Meschy (2007) equations (in g/day).

DISCUSSION

The phosphorus content in the studied plant species oscillates between 0.23 % and 0.10 % DM. In contrast, Dione et al. (2022) showed that the highest P content in forage plants in the agro-pastoral zone of Senegal varies between 0.82 % and 0.06 % DM. Additionally, Abdelkefi et al. (2004) reported a P concentration among some pastoral species in semi-arid and arid North Africa that varied from 0.05 to 0.52 % DM. However, Abdullah et al. (2013) found a lower P concentration in some browse species used as feed for livestock (0.016%).

Phosphorus has been known as a “master mineral” given that it affects the majority of metabolic processes (Rasby et al., 1997). The National Research Council (1984) recommended a P range of 0.12 to 0.48% for all ruminant classes, which aligns with present findings regarding the P concentration in the pastoral species used as feed for the northern goats of Morocco. The soil's P status, the plant's maturation stage, and the climate all impact the P content of forages, which varies widely (Underwood and Suttle, 1999).

The average Ca concentration in the studied pastoral plants was 1.79 g/kg DM. However, present findings were lower than the results reported by Abdullah et al. (2013) for browse species fed to livestock (1.79 vs 3 g/kg DM). Mirzaei (2012) revealed Ca concentration ranging from 4.17 to 2.42 g/kg in grass plants grazed by ruminants, which the results of the present experiment for grass plants (24.2 g/kg) are consistent with the findings of these researchers. Moreover, Chhabra et al. (2015), reported higher Ca content, approximately 0.77% (equivalent to 7 g/kg), in winter fodder in India.

These pastoral plants have Ca concentrations lower than the levels recommended by the NRC literature cited in Ghazanfar et al. (2011) and by Kessler (1991) in Ramirez-Orduña et al. (2005) for goat requirement. An exceeding of 1% of Ca content can decrease DM intake and reduce the absorption of trace minerals, especially zinc. However, the Ca requirements in grazing animals are a widely debated subject, as they are influenced by factors such as the type of animal, age, and production level (Khan et al., 2007).

The variations in Ca levels between the results of this study and values published in the literature are attributed to differences in forage species, species composition, seasonal and maturational stages, and changes in soil properties (Mirzaei, 2012). However, information about minerals in pastoral species, particularly those browsed by ruminants, is limited. These findings will provide a comprehensive knowledge of the mineral composition of grazing forage, thereby improving and ensuring goats' welfare and growth. In cases of deficiency, coupled with previous research regarding energy and protein content, they will enable the optimization of goat diets through informed adjustments.

Ca and P were studied together due to their close metabolic association, as an excess of either in the diet restricts the availability of both nutrients. The Ca:P ratio ranged from 0.45 to 1.88, which is in line with the recommended ratio by Abdulrazak et al. (2000), except for the grass plants with the highest Ca:P ratio (2.39). A higher ratio might interfere with the animal's ability to use Ca effectively (Fadel Elseed et al., 2002), and can also decrease livestock P absorbance.

The mineral elements are not produced in the body; the feed usually provides them. The concentrations of these elements in bodily fluids will vary depending on the availability of minerals, the quantity of dietary sources consumed, and the mineral content of feed (Suttle, 2010). The mineral concentrations of fodder plants are influenced by a wide range of environmental and plant parameters, such as the type of soil, species or strain/variety, seasonal circumstances during plant growth, plant maturity stage, and other management techniques (Underwood and Suttle, 1999).

P is a crucial mineral for animals, essential for their nutritional requirements. Approximately 80% of P is found in their skeleton, a critical bone and teeth component. Moreover, it plays a vital function in the transfer and utilization of energy. A P deficiency can decrease ruminant appetite, reduce fiber digestibility, and lower growth rates, weight gain, and reproduction (Drogoul et al., 2004). In cattle and small ruminants, a severe P deficiency (less than 1g/kg DM) can cause locomotor abnormalities, followed by paralysis of the rear end and spontaneous fractures (Meschy, 2010). P deficiency can also lead to losing appetite and consuming abnormal materials, such as bones, soil, wood, and flesh (Underwood and Suttle, 1999). Ramírez-Orduña et al. (2005) reported that the potential P intake of goats consuming shrubs in Mexico didn't fulfill their requirement, especially during years of low rainfall, which can harm goat performances.

Calcium is the most abundant mineral in the body, comprising 99% of the skeleton. A Ca deficiency can cause soft, weak, or deformed bones, leading to lameness, a condition known as osteomalacia or rickets. Ca is also required for blood clotting, nerve conduction, and muscular contraction (Hart, 2009). It was advised that the Ca requirements for maintaining, growing, and lactating sheep should be 1.2 to 2.6 g/kg (Mirzaei, 2012), which is higher than the potential mineral intake of the studied goats.

An excess or deficiency of these macrominerals can cause disruptions, slow growth, and limit the digestion of nutrients (NRC, 2007). To optimize animal well-being, it is preferable to provide them with the precise amounts they need based on their species and body weight. Mineral deficiency can also be caused by the feeding behavior of goats and the quality of their forage. As mentioned by Chebli et al. (2022a), there was a notable decline in forage production in the forested rangeland of Beni Arouss, located in the northern region of Morocco, with a 31% reduction in summer and a 47% decrease in autumn compared to the spring season. Furthermore, the intake rate was lower in the summer and autumn compared to the spring (4.94, 4.52 vs 5.57 g DM/min). The intake rate is influenced by season, as the goats tend to extend the duration of their grazing during summer days in comparison to the rainy season, as they strive to meet their intake requirements (Safari et al., 2011).

In this study, it is imperative to acknowledge several limitations that may have influenced the results to ensure the transparency and integrity of this research's findings. Methodological constraints, particularly concerning the estimation of dry matter intake on pasture and the estimation of mineral requirements, may have contributed to uncertainties in the results. In summary, present findings contribute to identifying the deficiencies in P and Ca within the goats' diet, thus emphasizing the necessity of considering them in ration formulation.

CONCLUSION

Mineral concentrations in browse plants differ significantly. Most plants had higher Ca levels than P during the rainy season (0,15 vs 0,10 g/kg DM). The levels of these macrominerals found in the browse shrubs grazed by goats in the northern region of Morocco were insufficient, resulting in a deficiency in their estimated daily mineral requirement, especially during the cold season (-0,23% deficit in Ca vs - 0,10 % deficit in P). To fulfill the mineral needs of goats, it is essential to formulate feeding strategies and implement grazing management. To ensure a balanced diet for goats, it is essential to include supplements rich in calcium (Ca) and phosphorus (P), such as a vitamin-mineral complex. The diet can also be enhanced by incorporating concentrated feed options like bitter vetch, barley, wheat bran, and sorghum.

This approach aims to meet their mineral requirement and improve the production and performance of goats. The findings could offer valuable and specific information for herders to design supplementary diet formulations, considering grazing activities and the quality of the consumed plant species.

DECLARATIONS

Corresponding author

Correspondence and requests for materials should be addressed to Asma Al Rharad; E-mail: asma.alrharad@etu.uae.ac.ma; ORCID: <https://orcid.org/0009-0000-1595-1472>

Ethical approval

All procedures involving animals in this study have been conducted according to the ethical standards of the Regional Center of Agricultural Research of Tangier. All the authors complied with the ARRIVE guidelines

Authors' contribution

Conceptualization: A. Al Rharad, A. Bouassab, M. Acherkouk, M. Ayadi

Data curation: M. Ayadi, M. Acherkouk, A. Al Rharad, A. Bouassab
Formal analysis: A. Al Rharad, A. Bouassab, M. Acherkouk, M. Ayadi
Investigation: M. Ayadi, M. Acherkouk, A. Al Rharad, A. Bouassab
Methodology: A. Al Rharad, A. Bouassab, M. Acherkouk, M. Ayadi
Validation: A. Al Rharad, A. Bouassab, M. Acherkouk, M. Ayadi
Writing – original draft: A. Al Rharad, M. Ayadi, A. Bouassab, M. Acherkouk,
Writing – review & editing: A. AL RHARAD, M. Ayadi, A. BOUASSAB, M. Acherkouk

Acknowledgements

The National Center for Scientific and Technical Research of Morocco, through the Research Excellence Grants Program, financially supported this research.

Consent to publish

All authors agree to the publication of this manuscript.

Competing Interests

The authors have not declared any conflict of interest.

REFERENCES

- Abdelkefi A, Ben Fadhel N, Ben Salah A, Boussaid M and Zaouali Y (2004). Pastoral plants in the arid environments of North Africa. In : Ferchichi A. (comp.), Ferchichi A. (collab.). Réhabilitation des pâturages et des parcours en milieux méditerranéens. Zaragoza : CIHEAM, 2004. (Cahiers Options Méditerranéennes ; n. 62), p. 55-59. <http://om.ciheam.org/om/pdf/c62/04600128.pdf>
- Abdullah M, Khan R, Yaqoob S and Ahmad M (2013). Mineral profile of browse species used as feed by grazing livestock in Cholistan rangelands, Pakistan. *Pakistan Journal of Nutrition*, 12: 135-143 <https://doi.org/10.3923/pjn.2013.135.143>
- Abdulrazak SA, Fujihara T, Ondiek JK and Ørskov ER (2000). Nutritive evaluation of some Acacia tree leaves from Kenya. *Animal Feed Science and Technology*, 85(1-2): 89-98. [https://doi.org/10.1016/S0377-8401\(00\)00133-4](https://doi.org/10.1016/S0377-8401(00)00133-4)
- Acherkouk M, Ayadi M and Al Gharad A (2022). Feed intake of goats from high-mountain pastures in the North of Morocco. *African and Mediterranean Agricultural Journal - Al Awamia*, 136 (Special Issue): 159-171. <https://doi.org/10.34874/IMIST.PRSM/afirmed-i136.34958>
- AOAC (1997). Official Methods of Analysis of AOAC International, 16th edition. Association of Official Analytical Chemists. Washington, DC, USA. pp. 2000. <https://www.cabidigitallibrary.org/doi/full/10.5555/19951414840>
- Ayadi M, Gharad A, Bouassab A, Jaber A and Acherkouk M (2022). Perennial pastoral fodder plants from the high mountains of the northern region of Morocco. *African and Mediterranean Agricultural Journal - Al Awamia*, 137: 1-21. <https://doi.org/10.34874/IMIST.PRSM/afirmed-i137.36751>
- Chebli Y, El Otmani S, Chentouf M, Hornick JL and Cabaraux JF (2021). Temporal variations in chemical composition, in vitro digestibility, and metabolizable energy of plant species browsed by goats in southern mediterranean forest rangeland. *Animals*, 11(5): 1441. <https://doi.org/10.3390/ani11051441>
- Chebli Y, El Otmani S, Hornick JL, Bindelle J, Cabaraux JF, and Chentouf M (2022a). Estimation of grazing activity of dairy goats using accelerometers and global positioning system. *Sensors*, 22(15): 5629. <https://doi.org/10.3390/s22155629>
- Chebli Y, El Otmani S, Hornick JL, Keli A, Bindelle J, Chentouf M, et al. (2022b). Using GPS Collars and Sensors to Investigate the Grazing Behavior and Energy Balance of Goats Browsing in a Mediterranean Forest Rangeland. *Sensors*, 22(3). <https://doi.org/10.3390/s22030781>
- Chebli Y, El Otmani S, Hornick JL, Keli A, Bindelle J, et al. (2022 c). Forage availability and quality, and feeding behaviour of indigenous goats grazing in a Mediterranean silvopastoral system. *Ruminants*, 2(1):74-89. <https://doi.org/10.3390/ruminants2010004>
- Chhabra S, Randhawa S and Bhardwaj S (2015). Macro and micro mineral profile in forage and blood plasma of water buffaloes with respect of seasonal variation. *Buffalo Bulletin*, 34 :45-50. <https://www.researchgate.net/publication/282938102>
- Delagarde R, Bonneau M, Boval M, Chapuis H, Fleurance G, et al. (2019). Methods for estimating individual feed intake in group-raised animals. *Défis scientifiques Phase*, Nov 2019, Rennes, France. <https://hal.inrae.fr/hal-02735680>
- Dione A, Bathily A, Ngom S, Sarr O, Diarra AR, Ngom D, et al. (2022). Bromatological and nutritional characterization of forage woody plants in the agro-pastoral zone of Ngouye in Senegal. *Journal of Applied Biosciences*, 177: 18471-18498. <https://doi.org/10.35759/JABs.177.10>
- Drogoul C, Gadoud R, Joseph MM, Jussiau R, Lisberney MJ, et al. (2004). *Nutrition et alimentation des animaux d'élevage*, Volume 2. 3e éd. Dijon: Educagri, France, p. 270. https://books.google.co.ma/books/about/Nutrition_et_alimentation_des_animaux_d.html?id=qGB9NygckoYC&redir_esc=y
- Fadel Elseed A, Amin AE, Alhadead K, Ati A, Sekine J, and Hamana K. (2002). Nutritive evaluation of some fodder tree species during the dry season in central Sudan. *Asian-Australasian Journal of Animal Sciences*. 15: 844-850. <https://doi.org/10.5713/ajas.2002.844>
- Ghazanfar S, Latif A, Mirza IH, and Nadeem MA (2011). Macro-minerals concentrations of major fodder tree leaves and shrubs of District Chakwal, Pakistan. *Pakistan Journal of Nutrition*, 10(5): 480-484. <https://doi.org/10.3923/pjn.2011.480.484>
- Glasser TA, Landau SY, Ungar ED, Perevolotsky A, Dvash L, Muklada H, et al. (2012). Foraging selectivity of three goat breeds in a Mediterranean shrubland. *Small Ruminant Research*, 102: 7-12. <https://doi.org/10.1016/j.smallrumres.2011.09.009>
- Hart S (2009). Meat Goat Nutrition. Proceedings of the 24th Annual Goat Field Day, Langston University. <https://pressbooks.umn.edu/app/uploads/sites/7/2019/08/Goat-nutrition-for-health.pdf>
- Jimenez LE, Avalos JO, Ortega OA, Hernandez JC, and Ronquillo MG (2024). Forage yield, chemical composition and potential milk yield using maize silage from asia, europe, north and south american continents: A systematic review. *Tropical and Subtropical Agroecosystems*, 27(3): 113. <http://dx.doi.org/10.56369/tsaes.5406>

- Kessler J (1991). Mineral nutrition of goats. In: Morand-Fehr, P. (ED), Goat Nutrition, Vol. 46. EAAP publication, pp 104-119. <https://ira.agroscope.ch/fr-CH/publication/14014>
- Khan IZ, Ashraf M and Hussain A (2007). Evaluation of Macro Mineral Contents of Forages: Influence of Pasture and Seasonal Variation. Asian-Australasian Journal of Animal Sciences, 20(6): 908-913. <https://doi.org/10.5713/ajas.2007.908>
- Manousidis T, Kyriazopoulos AP, Parissi ZM, Abraham EM, Korakis G and Abas Z (2016). Grazing behavior, forage selection and diet composition of goats in a Mediterranean woody rangeland. Small Ruminant Research, 145: 142-153. <https://doi.org/10.1016/j.smallrumres.2016.11.007>
- Manousidis T, Parissi ZM, Kyriazopoulos AP, Malesios C, Koutroubas SD and Abas Z (2018). Relationships among nutritive value of selected forages, diet composition and milk quality in goats grazing in a Mediterranean woody rangeland. Livestock Science, 218: 8-19. <https://doi.org/10.1016/j.livsci.2018.10.002>
- MAPMDREF (Ministère de L'Agriculture, de la Pêche Maritime, du Développement Rural et des Eaux et Forêts). Forêts en Chiffre. Département des Eaux et Forêts. 2018 [Forests in Figures. Department of Water and Forests. 2018]. Available online: <http://www.eauxetforets.gov.ma/ForetsMarocaines/ForetsChiffres/Pages/Forets-En-Chiffres.aspx>
- Meschy F (2007). Alimentation minérale et vitaminique des ruminants : actualisation des connaissances [Re-assessment of mineral and vitamin nutrition in ruminants]. INRAE Productions Animales, 20(2): 119-128. <https://hal.science/hal-01173433>
- Meschy F (2010). Nutrition minérale des ruminants [Mineral nutrition of ruminants]. Editions Quae, Collection Savoir-Faire, p. 208. <https://hal.science/hal-01173578>
- Meuret M, Bartiaux-Thill N, Bourbouze A, Rosenberger S, Vernery M, Sourbier Y, et al. (1985). Evaluation de la consommation d'un troupeau de chèvres laitières sur parcours forestier [An enzymatic procedure for estimating horse forage digestibility]— Méthode d'observation directe des coups de dents — Méthode du marqueur oxyde de chrome. Annales de Zootechnie, 34(2): 159-180. <https://doi.org/10.1051/animres:19850203>
- Mirzaei F (2012). Minerals profile of forages for grazing ruminants in Pakistan. Open Journal of Animal Sciences, 02(03): 133-141. <https://doi.org/10.4236/ojas.2012.23019>
- National Research Council, NRC (1984). Nutrient Requirements of Beef Cattle. 3th Rev. EDN., National Academy Press, National Research Council, Washington DC., USA. <https://doi.org/10.17226/19398>
- National Research Council, NRC (2007). Nutrient Requirements of Small Ruminants: Sheep, Goats, Cervids, and New World Camelids. The National Academies Press, Washington DC. <https://doi.org/10.17226/11654>
- Ramírez-Orduña R, Ramírez RG, González-Rodríguez H and Haenlein GFW (2005). Mineral content of browse species from Baja California Sur, Mexico. Small Ruminant Research, 57(1): 1-10. <https://doi.org/10.1016/j.smallrumres.2004.03.004>
- Rasby RJ, Brink DR and Rush IG (1997). EC97-277 Minerals and Vitamins for Beef Cows. Nebraska Cooperative Extension EC97-277 <https://www.researchgate.net/publication/228540932>
- Safari J, Mushi DE, Kifaro GC, Mtenga L A and Eik LO (2011). Seasonal variation in chemical composition of native forages, grazing behaviour and some blood metabolites of Small East African goats in a semi-arid area of Tanzania. Animal Feed Science and Technology, 164(1-2): 62-70. <https://doi.org/10.1016/j.anifeedsci.2010.12.004>
- Suttle NF (2010). Mineral Nutrition of Livestock. 4th Edition, CABI, Cambridge. <http://dx.doi.org/10.1079/9781845934729.0000>
- Underwood EJ and Suttle NF (1999) The mineral nutrition of livestock. 3rd Edition, CABI Publishing, Wallingford, UK. 3rd edition p. 614. <https://doi.org/10.1079/9781789240924.0011>

Publisher's note: Scienceline Publication Ltd. remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access: This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <https://creativecommons.org/licenses/by/4.0/>.

GROWTH PERFORMANCE AND PROFITABILITY OF WEANLING PIGS (*Sus scrofa domesticus* L.) FED PRE-STARTER DIET SUPPLEMENTED WITH NUCLEOTIDE

Arwin VERDIJO¹  and Hershey MONDEJAR²  

¹UltraBio Corporation, UBC Stock Farm, Inc., Barangay Singasing, Balamban, Cebu, 6317 Philippines

²College of Veterinary Medicine, Cebu Technological University Barili Campus, Barili, Cebu, 6317 Philippines

 Email: hersheymondejar75@gmail.com

 Supporting Information

ABSTRACT: Nucleotides can improve intestinal health by modulating the local immune response and intestinal mucosa development in weaned piglets. This study was conducted to evaluate the growth performance of post weaned piglets and evaluated the economic analysis of nucleotides supplementation for 30 days. A total of 120 mixed breed piglets were selected at the weaning stage and were used in the experiment as control group vs. treatment 2 supplemented with nucleotide. Each treatment consisted of 60 heads with three replications with 20 heads per replication arranged in Complete Randomized Design. Results were analyzed through Pairwise T and LSD tests. In terms of growth performance, results showed that supplementation of nucleotide had significantly increased the average daily gain, feed conversion ratio and weight gain by 0.50, 1.20 and 15.02 kg, respectively. However, there was no significant difference in terms of the average daily feed intake. With regards to the economic analysis, total production input had no effect with or without nucleotide supplementation but surprisingly, it had a gross margin of Php 95,900 (Philippine peso money) which was 5% more as with those that have supplementation. As to the net income, supplementation of nucleotides increased about 40.7% in comparison to control. Furthermore, a peso of investment could have a return of about 18 cents (1050 Php) more returns with supplementation, which apparently had 0.11 cents leverage compared to control group (0.07 cents). In conclusion, nucleotide supplementation not only improved the growth performance of post-weaned piglets but also enhanced profitability, offering a significant return on investment for swine producers. This makes nucleotide supplementation a promising strategy for improving both animal health and economic outcomes in swine production.

Keywords: Benefit Cost Ratio, Daily weight gain, Feed conversion ratio, Net Income, Nucleotides.

INTRODUCTION

Major adjustments in technologies in swine management and nutritional programs have been significantly improved for efficiency and quality in commercial swine production. Providing the primary needs of the weanling pigs, like feeds, water, and air, is crucial to their growth performances (Gaillard et al., 2020). One of the stressful moments in pig's life is on weaning, which is often accompanied by physiological variations in the gastrointestinal tract (GIT) (Pluske, 2016). Weaning is also challenging because of not only, the new atmosphere but also the transition to dry feed from sow's milk causing nutritional stress (Van Kerschave et al., 2023). One of the most health challenging experiences for pigs after weaning, is post-weaning lag (Vasa et al., 2024). Post-weaning lag is the condition where changes in the weaned pig's intestinal biochemistry can contribute to diarrhea, weight loss, a decline in appetite, and reduced growth after a new environment and the initial separation from the sow and the littermates (Dunshea et al., 2003; Muro et al., 2023). Also, after weaning, nutrient digestibility is reduced by the collapse of the intestinal barriers due to intestinal inflammation and oxidative stress (Lallès et al., 2004; Moeser et al., 2017). Thus, reducing oxidative stress by dietary nucleotides can be associated with improved health status and growth performance in nursery pigs (Jang and Kim, 2019; Duarte et al., 2019).

Dietary nucleotide supplementation to pre-starter feeds (after weaning) may provide a readily available nucleotide needed by the animals that can be used as a precursor for the maturation of intestinal mucosa, which can help alleviate the effect of early nutritional stress, brought about commonly by weaning (Correa et al., 2021). Thus, improved intestinal maturation when nucleotides are included in the diet can reduce the onset of diarrhea in weaned piglets (Martinez-Puig et al., 2007).

Nucleotides are organic molecules made up of a nitrogenous base, a pentose sugar, and a phosphate. They serve as the building blocks of nucleic acids. Nucleic acids such as deoxyribonucleic acids (DNA) and ribonucleic acids (RNA) are essential in cellular growth (Minchin and Lodge, 2019). Nucleotides can be synthesized in the body via two pathways; the first type is the de novo pathway which starts with metabolic precursors such as amino acids, ribose, carbon dioxide, and

RESEARCH ARTICLE
 PII: S222877012500002-15
 Received: September 11, 2024
 Revised: January 14, 2025
 Accepted: January 15, 2025

ammonia (Watson and Crick, 1953). The synthesis using the de novo pathway is energy-requiring. It utilizes many metabolic pathways that require a large amount of energy to proceed. In addition, some tissues in the body, like intestinal mucosa, have a limited capacity to synthesize purine nucleotides via the de novo pathway (Boza et al., 2002). Therefore, there can be a need for an exogenous supply of bases that can be utilized via salvage pathway for optimal function (Uauy et al., 1990). If the nucleotide requirements of the intestinal mucosa are quickly met, this may result in rapid intestinal development and maturation. On the other hand, the second type is the salvage pathway, which retrieves free purine and pyrimidine bases, as well as nucleosides, from the degradation of nucleic acids or from the diet (Moffatt and Ashihara, 2002; Dinardo et al., 2022). The salvage pathway is much more efficient than the de novo pathway, and this is what the study aims to develop.

The nucleosides and nucleotides are identified as bioactive compounds with significant potential for application in food and nutrition (Bezerra et al., 2024). These compounds could be integrated into functional foods to improve metabolic processes, enhance immune function, and support cellular regeneration. For infant nutrition, nucleotide fortification in infant formulas offers a strategy to replicate the immune and gut health benefits of human breast milk. Additionally, immune-boosting foods, such as soups or fortified beverages, could benefit from nucleotide inclusion to strengthen immune responses (Prakash et al., 2020). Lastly, the development of nucleotide-based dietary supplements presents an opportunity for promoting cellular health and DNA repair, highlighting the broader potential of these compounds as bioactive ingredients in food products designed for health enhancement. Moreover, previous studies found that supplementation of dietary nucleotides in nursery diets enhanced intestinal morphology (Jang and Kim, 2019; Valini et al., 2021) and intestinal immunity (Waititu et al., 2017). A research report has shown that the inclusion of a nucleotide base at a concentration of 0.5% in diets resulted in enhanced weight gain and increased feed intake among weaned pigs (Zomborszky-Kovacs et al., 2000). Moreover, it is still speculative if the efficiency of absorption of free nucleotides dissolved in water is different from that of bound nucleotides in feed ingredients (Sauer et al., 2012). Meanwhile, separate from the pig's environment, feeding strategies, age at injecting, and pig genotype and age are mediated by psychological and behavioral stress. A study has indicated that the addition of nucleotide supplementation within the range of 50 to 250 mg/kg in diets can potentially benefit newly weaned pigs. These benefits include improved growth performance, potentially attributed to reduced intestinal inflammation and oxidative stress, as well as enhanced intestinal villi structure and energy digestibility (Jang and Kim, 2019). However, little is known about the information on the effects of nucleotide on the growth of newly weaned piglets. Thus, present study was conducted to evaluate the effects of nucleotide supplementation on the nutrient digestibility and growth performance of pigs when added to pre-starter feedings over a period of 30 days.

MATERIALS AND METHODS

Time and place of study

The experiment was conducted at the Nursery Section, Department of the Research and Development Farm of UBC Stock Farm, Incorporated at Sitio Dao, Brgy. Singasing, Balamban, Cebu, Philippines from October to November 2023.

Experimental design and layout

The 120 heads total of mixed-breed newly weaned pig (Landrace × Duroc × Large White) were chosen at the weaning stage. A Completely Randomized Design (CRD) was used in the experiments with two experimental treatments, the control group (T1) consisted of weaned pigs given pure pre starter diet (without nucleotide supplementation) and treatment T2 were pigs supplemented with nucleotides added in the formulated feeds. Each group contained 60 heads which was further divided into 3 subgroups representing the three replications. The selection of the newly weaned piglets was based on their size and uniformity, as well as their sex, to ensure a fair distribution of males and females in each treatment group.

Care and management

Before conducting the experiment, the pen underwent thorough cleaning, disinfection, and sanitation process for approximately 14 days. This was done prior to transferring the pigs from the farrowing section to the weaning area. Each pen was provided with rotary feeders and one linear feeder. Feeds and clean water were provided in an ad libitum scheme. Vitamins and mineral supplements were administered via drinking water upon loading via medicator tank and during stressful periods such as challenging weather (extreme cold and extreme heat) and disturbance stress. Curtains and lighting were also installed on a case-by-case basis to provide the most comfortable conditions for the newly weaned pigs. Vaccination of Hog Cholera at 35 days old and *Mycoplasma pneumonia* and *Haemophilus parasuis* 2 at 42 days old was administered to all the experimental pigs.

Experimental diets

Both groups were administered the same feed formulations of the pre-starter diet. Group 2 animals received the pre-starter diet with 500 grams of nucleotide per ton of feeds. The nucleotides were included during the mixing of feeds in the feed mill where the feeds were produced. The source of nucleotide was provided from the yeasts extracts through hydrolysis (Bioiberica, Spain, Table 1).

Table 1 - Nucleotide concentration (ppm) in some commonly used feed ingredients (as-is basis) (Mateo et al., 2004)

Ingredient	Nucleotide: 5' -AMP	5'- CMP	5'-GMP	5'-IMP	5'-UMP
Barley	1	2	1	1	0
Casein	0	1	0	0	0
Corn	2	3	3	1	0
Fishmeal	11	26	2	35	1
Naked oats	3	3	3	1	1
Non-fat dried milk	0	65	0	195	106
Plasma protein, spray dried	2	2	2	1	0
Red blood cells, spray dried	44	0	3	6	2
Soybean meal, 44%	8	16	3	2	9
Soy protein concentrate	1	0	2	1	0
Whey dried	19	270	0	4	1
Whey protein concentrate	0	34	0	159	89

Adenosine 5' monophosphate (5' AMP), cytidine 5' monophosphate (5' CMP), guanosine 5' monophosphate (5' GMP), inosine 5' monophosphate (5' IMP), and uridine 5' monophosphate (5' UMP)

Data Collection

Body weight and feed measurements

From the first day to the day 30 of feeding, both feed consumption and body weights of the experimental animals were monitored. The collected data was analyzed, summarized, and categorized according to metrics like the ADG or Average Daily Gain, the ADFI or Average Daily Feed Intake, and the FCR or Feed Conversion Ratio (FCR) during the entire feeding duration. Additionally, calculations were made: 1) feed price per pig; 2) feed price per kilo of gain; 3) income over feed and 4) pig price.

Body weight and weight gain

The body weight at weaning (start of feedings) and after 30 days of feedings (end of feedings) was determined and recorded. The average body weight for each replicate was determined by dividing the group weight by the number of pigs in each pen. To determine the body weight gain for each replicate, the final weight at the end of the experiment was subtracted from the initial body weight.

Feed consumption

The feed intake of the pigs was calculated by subtracting the number of left-over feeds from the amount of feed offered divided by the corrected number of pigs (less mortality).

Feed conversion ratio

The feed conversion ratio was determined by dividing the feed consumption by the corresponding weight gain at the conclusion of the pre-starter feeding period for each replicate.

Economic analysis

Variable cost of the treatments

Cost of Treatment 1 = total cost of compounded feeds given + other cost, if any

Cost of Treatment 2 = total cost of compound feeds + nucleotide supplements, other cost, if any.

To calculate the financial return, the total expenses were subtracted from the total expected sales.

Net profit = Total sales - Total Expenses

Where:

Total sales = were calculated by multiplying the final weights of the pigs by the current selling price.

Total expenses = were expenses incurred throughout the experiments.

Benefit-cost ratio

The discounted cash inflows and outflows ratios, which must be equal to or larger than one, are known as the Benefit-cost ratio (BCR). The ratio must be at least 1:1, indicating that the expense incurred and the benefit received are equal. If the benefits outweigh the costs, the ratio should be greater than 1. This parameter shows the rate of return and is worked out by dividing the total gross return by the total cost return.

$$BCR = \frac{\text{Total Gross Return}}{\text{Total Cost}}$$

Statistical analysis

The data were analyzed using STAR version 2.0.1 or Agricultural Statistics software. Pairwise t-test analysis was used to compare treatments, while Least Significant Difference (LSD) test was used to determine significant differences between responses variables among different treatments in the study to ensure the reliability of the results obtained.

RESULTS AND DISCUSSION

Growth performance of pigs

The effectivity of nucleotide supplementation on post-weaned pigs has been studied earlier, showing contradicting results including but not limited to its dietary effects (Perricone et al., 2020), physiological attributes (Reina et al., 2014), and immune system (Superchi et al., 2012). Nonetheless, the effectiveness of nucleotide supplementation on post-weaned piglets has been the focus of the study.

Different growth performance or test variables have been tested in the study, including the average daily gain (ADG), feed conversion ratio (FCR), and weight gain (WG). As presented in Table 2, the effect of supplementation of nucleotide to the pre-starter on post-weaned had significantly improved their ADG, FCR, and WG, respectively. Results showed that nucleotide supplementation had significantly increased the daily gain by approximately 0.07 kg compared to the treatment without nucleotide supplementation. Regarding this, the results may be attributed to the positive effects of nucleotide supplementation to enhance nutrient absorbability in the small intestine by promoting the development of enhanced villi. This, in turn, leads to improved performance in pigs. The findings of this study are consistent with previous literature that has reported varying outcomes concerning nucleotide supplementation in pigs. For instance, Perricone et al. (2020) found that nucleotide supplementation can positively affect dietary intake and growth metrics, albeit with differing results based on the specific composition and dosage of nucleotides used. Similarly, Reina et al. (2014) reported improvements in physiological attributes associated with nucleotide intake, while Superchi et al. (2012) highlighted enhancements in immune response. While some studies have reported contradictory outcomes regarding the effectiveness of nucleotides on growth performance, this research supports the notion that, when utilized correctly, nucleotides can significantly enhance growth metrics. The discrepancies observed in previous studies may be attributed to various factors, including differences in experimental design, genetic backgrounds of the pigs, and environmental conditions, which can all influence nutrient metabolism and growth performance (Rocadembosch et al., 2016; Pierozan et al., 2016).

Table 2 - Growth performance of pigs during the pre-starter stage in the post-weaning period (PWP), comparing those feds with and without nucleotide supplementation.

Treatment	Average daily gain (kg)	Feed conversion ratio	Average daily feed intake (kg)	Weight gain (kg)
T1 (Without Nucleotide)	0.430 ^b	1.47 ^a	0.64	13.04 ^b
T2 (With Nucleotide)	0.500 ^a	1.20 ^b	0.63	15.02 ^a
P-Value	0.03	0.04	0.56	0.03
F Value	9.50	8.74	0.40	9.52
CV %	5.41	8.49	2.04	5.60

Means with the same letter designations were not significantly different at 5% level LSD test.

Feed conversion ratio served as the feed used per pound of weight gain. The results of the study demonstrated that the supplementation of nucleotides effectively reduced the amount of feed required to produce a kilogram of live weight. It takes only 1.20 kg of feeds to have a kilogram of its equivalent live weights. A lower FCR indicates that pigs are efficiently converting feed into body weight. The Feed Conversion Ratio (FCR) in pigs can differ across countries (Rocadembosch et al., 2016), stages of growth (Pierozan et al., 2016), environmental conditions (Agostini et al., 2014), and genetic factors (Bereskin, 1986). However, as a general guideline, a pig's FCR should ideally fall within the range of 3:1. In terms of the weight gain, results showed that the higher the average daily gain, this also resulted to a higher weight gain at harvesting (Table 2). Statistical evidence demonstrated that nucleotide supplementation resulted in a significant increase in live weight by 15.02 kg, which corresponds to a 7% higher weight compared to the group without nucleotide supplementation.

Statistical analysis showed no significant difference between the two treatments, indicating no effects on the daily feed intake. Based on the findings, the addition of nucleotide supplementation increased the average daily gain, feed conversion, and ultimately the live weight gain of post-weaned pigs. This indicates that nucleotide supplementation positively impacted the growth and performance of the pigs. Other scientists have also confirmed the effectiveness of

nucleotides, not only in terms of improving growth performance but also in boosting the immune system and reducing environmental stresses. This indicated that nucleotides have multiple beneficial effects beyond just promoting growth in animals. Nucleotides have important effects on the growth and development of rapid turnover cells, such as those in the immune system and the gastrointestinal (GI) tract (Dancey et al., 2006) However, it has been observed that animals fed diets lacking in nucleotides exhibit lower immune responses, as noted by Chandra and Kumari (1994). This suggests that the absence of nucleotides in the diet can negatively impact the immune system of animals. Dietary nucleotides recognized a potential antibiotic alternative, have been found to exhibit beneficial effects on intestinal hyperemia, systemic immunity, small-intestinal growth, and hepatic composition in pigs (Jang and Kim, 2019). Moreover, Sauer et al. (2011) summarized that supplementation of nucleotides affects immune function, nutrient metabolism, hepatic morphology and function and accelerates T-cell-dependent antibody production (Grimble et al., 2001).

Economic analysis of piglets

Economic analysis of pigs differs due to several reasons like breed type, cost of feeds fed, operating expenses, vitamins and medicine for diseases of pigs, market niche, current buying price, location, and demand pool in the society. In this study, the computation of the total variable cost is based on the existing care and maintenance practices that are uniformly applied to all blocks (Table 3). This means that the cost calculations consider the standard care and maintenance procedures that are consistently implemented across all blocks. The variable cost includes the price of pigs (20 heads per block), feeds, electricity, water, managerial pay/labor cost, as well as vaccines and vitamins. According to the analysis, there is no significant difference observed in terms of variable cost. This means that the addition of nucleotides does not result in any significant variation in the production inputs required for raising pigs. In terms of its gross income, calculation includes the total live weight of 20 pigs at harvest times at a price of each pig at 210 per kilogram, which was applied and computed to all blocks.

Table 3 - Economic analysis of swine production at the pre-starter stage during the post-weaning period (PWP), comparing the outcomes with and without the inclusion of nucleotides.

Treatment	Total per Philippine Peso		Net income	Benefit cost ratio
	Variable cost	Gross income		
T1 (Without nucleotide)	81,497.67	87,535 ^b	6,037.33 ^b	1.07 ^b
T2 (With nucleotide)	81,574.33	95,900 ^a	14,325.49 ^a	1.18 ^a
P Value	0.82	0.05	0.04	0.04
F Value	0.06	7.67	9.20	8.98
CV %	0.49	4.03	32.86	3.75

Means with the same letter designations were not significantly different at 5% level LSD test.

The study results indicate that nucleotide supplementation leads to higher live weight, resulting in a higher gross income during harvesting. The statistical results indicated that nucleotide supplementation has significantly increased income compared to the non-supplemented group. A gross income of Php 95,900 (1648.39 USD) was incurred with nucleotide supplementation, which is 3 percent (3.84%) higher compared to Php 87,535 (1504.61 USD) without nucleotide supplementation. It simply means that the higher the gross income compared to variable cost, the higher is the net income. It is true that this study has significant implications, as the supplementation of nucleotides in T2 resulted in a 40.70 percent leverage compared to T1 (without nucleotide supplementation). Moreover, in terms of its benefit-cost ratio, the use of nucleotide supplementation incurred 1050 Php (18 cents) return on a peso of investment. This demonstrated a significant difference, with a lower return on the treatment without supplementation, yielding only 0.07 cents of return on each peso of investment.

Generally, the study implies that the addition of nucleotide supplements does not affect the overall production cost. However, it positively impacts the gross income by increasing pig live weights, net income, and the return on peso investment by 18 cents per peso, respectively.

CONCLUSION

The addition of nucleotide supplementation resulted in developments in average daily gain, feed conversion ratio, and the weight gain, although it did not have a significant impact on the average daily feed intake. Also, in terms of its economic analysis, supplementation of nucleotide had shown no effect on the total production cost but had higher gross income with 5% leverage as compared to control group, 40.70% increase in net income, and a higher benefit of 18 cents for a peso spending compared to treatment without supplementation.

DECLARATIONS

Corresponding author

Correspondence and requests for materials should be addressed to Hershey MONDEJAR; E-mail: hersheymondejar75@gmail.com; ORCID: <https://orcid.org/0000-0002-6123-7779>

Data availability

The datasets used and/or analysed during the current study available from the corresponding author on reasonable request.

Ethical considerations

This study adhered to the Animal Welfare Act of the Philippines 1998 ([The Philippine Animal Welfare Society, 1998](#)) to ensure the humane treatment of the post-weaned piglets. Proper care, including adequate housing, nutrition, and veterinary support, was provided throughout the experiment. Stress and pain were minimized by handling the piglets gently and using trained personnel for all procedures. Clear criteria for humane endpoints were established. The number of animals used was kept to the minimum necessary to achieve valid results, following the principle of reduction. The study-design ensured that the use of animals was ethically justified by the potential benefits to swine production ([FAO, 2015](#)).

Authors' contribution

Both authors contribute on conceptualization of the study, data analysis and the write up of the manuscript

Competing interests

The authors declare no competing interests in this research and publication.

REFERENCES

- Agostini PS, Fahey AG, Manzanilla EG, O'Doherty JV, De Blas C, and Gasa J (2014). Management factors influencing mortality, feed intake, and feed conversion ratio of grow-finishing pigs. *Animal*, 8(8):1312-1318. <https://doi.org/10.1017/S1751731113001912>
- Bereskin B (1986). A genetic analysis of feed conversion efficiency and associated traits in swine. *Journal of Animal Science*, 62(4):910-917. <https://doi.org/10.2527/jas1986.624910x>
- Bezerra FW, Moraes VS, Pantoja GV, Salazar MD, da Silva MP, Ferreira BF, da Silva Martins LH (2024). Nucleosides and Nucleotides as Classes of Bioactive Food Compounds. In: *Bioactive Compounds from Food* (pp. 205-219). CRC Press. <https://www.taylorfrancis.com/chapters/edit/10.1201/9781003386247-15/nucleosides-nucleotides-classes-bioactive-food-compounds-fernanda-wariss-figueiredo-bezerra-vinicius-sidonio-vale-moraes-gabriela-vieira-pantoja-marielba-de-los-angeles-rodriguez-salazar-marcilene-paiva-da-silva-brunna-fernanda-zahlouth-ferreira-luiza-helena-da-silva-martins>
- Boza JJ, Moënnos D, Bournot CE, Blum S, Zbinden I, Finot PA, et al. (2000). Role of glutamine on the de novo purine nucleotide synthesis in Caco-2 cells. *European journal of Nutrition*, 39:38-46. <https://doi.org/10.1007/s003940050074>
- Chandra RK, and Kumari S (1994). Nutrition and immunity: an overview. *Journal of Nutrition*, 124 (Suppl. 8): 1433S-1435S. https://doi.org/10.1093/jn/124.suppl_8.1433S
- Correa F, Luise D, Archetti I, Bosi P, and Trevisi P (2021). Investigation of early supplementation of nucleotides on the intestinal maturation of weaned piglets. *Animals*, 11(6):1489. <https://doi.org/10.3390/ani11061489>.
- Dancey CP, Attree EA, Brown KF (2006). Nucleotide supplementation: a randomised double-blind placebo-controlled trial of IntestAidIB in people with Irritable Bowel Syndrome [ISRCTN67764449]. *Nutrition Journal*, 5:16. <https://doi.org/10.1186/1475-2891-5-16>
- Dinardo FR, Maggiolino A, Martinello T, Liuzzi GM, Elia G, Zizzo N, et al. (2022). Oral administration of nucleotides in calves: Effects on oxidative status, immune response, and intestinal mucosa development. *Journal of Dairy Science*, 105(5):4393-409. <https://doi.org/10.3168/jds.2021-20804>
- Duarte ME, Zhou FX, Dutra Jr WM, and Kim SW (2019). Dietary supplementation of xylanase and protease on growth performance, digesta viscosity, nutrient digestibility, immune and oxidative stress status, and gut health of newly weaned pigs. *Animal Nutrition*, 5(4):351-8. <https://doi.org/10.1016/j.aninu.2019.04.005>
- Dunsha FR, Kerton DK, Cranwell PD, Campbell RG, Mullan BP, King RH, et al. (2003). Lifetime and post-weaning factors influencing performance indices of pigs. *Australian Journal of Agricultural Research*, 54(4): 363-370. <https://doi.org/10.1071/AR02111>
- FAO (2015). Administrative Circular No. 04 Series of 2015 on Registration of Animal Control Facility, Rome. <https://www.fao.org/faolex/results/details/en/c/LEXFAOC019221/#:~:text=It%20is%20the%20purpose%20of,trade%20or%20as%20household%20pets.>
- Gaillard C, Brossard L, and Dourmad JY (2020). Improvement of feed and nutrient efficiency in pig production through precision feeding. *Animal Feed Science and Technology*, 268:114611. <https://doi.org/10.1016/j.anifeeds.2020.114611>.
- Grimble GK, and Westwood OM (2001). Nucleotides as immunomodulators in clinical nutrition. *Current Opinion in Clinical Nutrition & Metabolic Care*, 4(1):57-64. https://journals.lww.com/co-clinicalnutrition/abstract/2001/01000/nucleotides_as_immunomodulators_in_clinical.11.aspx
- Jang KB, and Kim SW (2019). Supplemental effects of dietary nucleotides on intestinal health and growth performance of newly weaned pigs. *Journal of Animal Science*, 97(12):4875-4882. <https://doi.org/10.1093/jas/skz334>
- Lallès JP, Sève B, Pié S, Blazy F, Laffitte J, and Oswald IP (2007). Weaning is associated with an upregulation of expression of inflammatory cytokines in the intestine of piglets. *The Journal of nutrition*, 134(3): 641-647. <https://doi.org/10.1093/jn/134.3.641>

- Martinez-Puig D, Manzanilla EG, Morales J, Borda E, Pérez JF, Piñeiro C, et al. (2007). Dietary nucleotide supplementation reduces occurrence of diarrhoea in early weaned pigs, *Livestock Science*, 108 (1-3): 276-279. <https://doi.org/10.1016/j.livsci.2007.01.099>
- Mateo CD, Peters DN, and Stein HH (2004). Nucleotides in sow colostrum and milk at different stages of lactation. *Journal of Animal Science*, 82(5):1339-1342. <https://doi.org/10.2527/2004.8251339x>
- Minchin S and Lodge J (2019). Understanding biochemistry: structure and function of nucleic acids. *Essays in Biochemistry*, 63(4):433-56. <https://doi.org/10.1042/EBC20180038>
- Moeser AJ, Pohl CS, and Rajput M (2017). Weaning stress and gastrointestinal barrier development: Implications for lifelong gut health in pigs. *Animal Nutrition*, 3(4):313-21. <https://doi.org/10.1016/j.aninu.2017.06.003>
- Moffatt BA, and Ashihara H (2002). Purine and pyrimidine nucleotide synthesis and metabolism. *The Arabidopsis Book/American Society of Plant Biologists*. 2002;1. <https://doi.org/10.1199/tab.0018>
- Muro BB, Carnevale RF, Monteiro MS, Yao R, Ferreira FN, Neta CS, et al. (2023). A systematic review and meta-analysis of creep feeding effects on piglet pre-and post-weaning performance. *Animals*, 13(13):2156. <https://doi.org/10.3390/ani13132156>
- Perricone V, Comi M, Bontempo V, Lecchi C, Cecilian F, Crestani M, et al. (2020). Effects of nucleotides administration on growth performance and immune response of post-weaning piglets. *Italian journal of Animal Science*, 19(1):295-301. <https://doi.org/10.1080/1828051X.2020.1738966>
- Pierozan CR, Agostini P, and Gasa J (2016). Factors affecting daily feed intake and feed conversion rates of pigs in feeding houses: a company case study. *Porcine Health Management*, 2: article no. 7. <https://doi.org/10.1186/s40813-016-0023-4>
- Pluske JR (2016). Invited review: aspects of gastrointestinal tract growth and maturation in the pre-and postweaning period of pigs. *Journal of Animal Science*, 94(suppl_3): 399-411. <https://doi.org/10.2527/jas.2015-9767>
- Prakash S, Jyoti P, Srinivasa NH, and Shamsudeen P (2020). Importance of dietary nucleotides and its impact on immunomodulation: a review. *Journal of Immunology and Immunopathology*, 22 (1): 10-18. <http://dx.doi.org/10.5958/0973-9149.2020.00002.7>
- Reina R, Tzvetanov M, and Jørgensen H (2014). The effects of dietary nucleotides on growth performance and health of pigs. *Journal of Animal Science*, 92(2):850-857. <https://doi.org/10.2527/jas.2013-6887>
- Rocadambosch J, Fernández C, and Casanova N (2016). Factors affecting the feed conversion ratio in growing pigs: A review. *Livestock Science*, 183:136-146. <https://doi.org/10.1016/j.livsci.2015.11.003>
- Sauer N, Eklund M, Bauer E, Gänzle MG, Field CJ, Zijlstra RT, et al. (2012). The effects of pure nucleotides on performance, humoral immunity, gut structure and numbers of intestinal bacteria of newly weaned pigs. *Journal of Animal Science*, 90(9):3126-3134. <https://doi.org/10.2527/jas.2011-4417>
- The Philippine Animal Welfare Society (1998). The animal welfare act of 1998 as amended (Ra 8485 As Amended by Ra10631). https://paws.org.ph/downloads/ra8485_as_amended_by_ra10631.pdf
- Uauy R, Quan R, and Gil A (1994). Role of nucleotides in intestinal development and repair: Implications for infant nutrition. *Journal of Nutrition*, 124:1436S-1441S. https://doi.org/10.1093/jn/124.suppl_8.1436S
- Valini GA, Duarte MS, Calderano AA, Teixeira LM, Rodrigues GA, Fernandes KM, et al. (2021). Dietary nucleotide supplementation as an alternative to in-feed antibiotics in weaned piglets. *Animal*, 15(1):100021. <https://doi.org/10.1016/j.animal.2020.100021>
- Van Kerschaver C, Turpin D, Michiels J, and Pluske J. (2023). Reducing weaning stress in piglets by pre-weaning socialization and gradual separation from the sow: a review. *Animals*, 13(10):1644. <https://doi.org/10.3390/ani13101644>
- Vasa SR, Gardiner GE, Arnaud EA, O'Driscoll K, Bee G, and Lawlor PG (2024). Effect of supplemental milk replacer and liquid starter diet for 4 and 11 days postweaning on intestinal parameters of weaned piglets and growth to slaughter. *Animal*, 18(9): 101271. <https://doi.org/10.1016/j.animal.2024.101271>
- Waititu SM, Yin F, Patterson R, Yitbarek A, Rodriguez-Lecompte JC, Nyachoti CM (2017). Dietary supplementation with a nucleotide-rich yeast extract modulates gut immune response and microflora in weaned pigs in response to a sanitary challenge. *Animal*, 11:2156-2164. <https://doi.org/10.1017/S1751731117001276>
- Watson JD, and Crick FH (1953). Molecular structure of nucleic acids: a structure for deoxyribose nucleic acid. *Nature*, 171(4356):737-738. <https://doi.org/10.1038/171737a0>
- Zomborszky-Kovács M, Bardos L, Biro H, Tuboly S, Wolf-Táskai E, Toth A, et al. (2000). Effect of beta-carotene and nucleotide base supplementation on blood composition and immune response in weaned pigs. *Acta Veterinaria Hungarica*, 48(3):301-311. <https://doi.org/10.1556/avet.48.2000.3.7>

Publisher's note: Scienceline Publication Ltd. remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access: This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <https://creativecommons.org/licenses/by/4.0/>.

SARCOCYSTOSIS IN ALPACAS AND LLAMAS: REGIONAL, MARKET, AND MUSCLE-SPECIFIC PREVALENCE PATTERNS

Edgar GARCIA-OLARTE¹ , Jordan NINAHUANCA¹ , Wilder SUAREZ-REYNOSO¹ , Yakelin MAURICIO-RAMOS² , Maria Flores GUILLEN¹ , Ide Unchupaico PAYANO¹ , Armando Aquino TACZA¹ , and Wilhelm Guerra CONDOR³ 

¹Departamento Académico de Zootecnia, Universidad Nacional del Centro del Perú, Av. M.C. 3909, Huancayo, 12000, Perú

²Laboratorio de Sanidad Animal, Universidad Nacional del Centro del Perú, Av. M.C. 3909, Huancayo, 12000, Perú

³Facultad de Ciencias de la Salud, Universidad Peruana Los Andes, Huancayo, 12002, Perú

✉ Email: jninhuanca@unpc.edu.pe

➤ Supporting Information

ABSTRACT: The objective of this study was to determine the effect of species (alpacas and llamas), markets in the city of Huancayo (Ferrocarril Commercial Center, Nueva Esperanza, and Nazareth), and muscle groups on the prevalence of *Sarcocystis* sp. Between January and October 2023, a total of 2,211 carcasses were inspected, comprising 1,716 alpacas and 495 llamas. The results indicated a prevalence of 21% (104/495 carcasses) in llamas and 8% (138/1,716 carcasses) in alpacas. By region of origin, the prevalence in alpacas was reported as follows: Huancavelica (7.7%) with 14/181 carcasses, Junín (6.7%) with 55/820 carcasses, and Lima (9.7%) with 69/715 carcasses. For llamas, the Lima region exhibited the highest prevalence of sarcocystosis (33.9%) with 72/212 carcasses, followed by Huancavelica (14.7%) with 14/102 carcasses, and Junín (9.8%) with 18/181 carcasses. Regarding the markets, the Ferrocarril market presented the highest risk of contamination, serving as the reference group for comparison. In contrast, the Nazareth and Nueva Esperanza markets showed significantly lower odds of *Sarcocystis* sp. presence, with Odds Ratios (ORs) of 0.38 and 0.25, respectively. For muscle groups, the anatomical distribution of *Sarcocystis* sp. cysts revealed a preferential localization in the leg (OR = 1.65) and neck (OR = 1.20) compared to the shoulder. This investigation provides significant data on the prevalence of *Sarcocystis* sp. in alpacas and llamas, highlighting a higher prevalence in llamas despite their smaller sample size. These findings emphasize the need for targeted interventions to address this parasitic infection in camelid production systems.

Keywords: Animal products, Camelids, Carcass quality, Mantaro valley, Parasite.

INTRODUCTION

Peru boasts a diverse range of climatic conditions, coupled with ample availability of natural pastures (Estremadoyro et al., 2024), which serve as the primary food source for South American camelids. These animals exhibit remarkable efficiency in converting feed, requiring only 1.5 - 2% of dry matter (DM) relative to their body weight for survival (Coleman et al., 2010). With approximately 4.3 million alpacas and 1.2 million llamas, Peru sustains a substantial population of these valuable species (Catoa et al., 2016).

Sarcocystosis is a parasitic disease caused by protozoa of the genus *Sarcocystis* sp. (Fayer et al., 2015), this protozoan belongs to the domain Protozoa, phylum Apicomplexa, class Sporozoa, suborder *Eimeriorina*, and family *Sarcocystidae* (Yang et al., 2005), which affects a wide range of intermediate hosts, including domestic and wild animals (Amalfitano et al., 2017). In camelids, particularly alpacas (*Vicugna pacos*) and llamas (*Lama glama*), this parasitosis is of growing concern due to its significant impact on meat quality (Gareh et al., 2020), food safety, and public health (Fernandez-F et al., 2022; Rodríguez et al., 2023). The life cycle of *Sarcocystis* sp., involves both definitive and intermediate hosts, with the parasite undergoing asexual reproduction in the musculature of the intermediate host (Lucas et al., 2019; Wu et al., 2022), forming cysts that can compromise meat suitability for human consumption. Definitive hosts, such as domestic and wild canids, play a crucial role in the dissemination of the parasite, perpetuating its cycle (Lindsay and Dubey, 2020). Despite the cultural and economic importance of alpacas and llamas in the Andean regions, there is limited information on the prevalence and distribution of sarcocystosis in their carcasses (Shams et al., 2022), particularly in commercial markets. Previous studies have documented prevalence rates varying widely between regions and production systems, suggesting that environmental conditions, animal management practices, and market dynamics may significantly influence the epidemiology of this disease (Valentine and Martin, 2007; Condori-Quispe et al., 2019). However, these studies often lack granular insights into the role of specific factors, such as geographic location, market practices, and anatomical distribution of cysts, which are essential for designing targeted control strategies.

This study addresses a critical gap in knowledge by evaluating the prevalence of *Sarcocystis* macrocysts in alpaca (*Vicugna pacos*) and llama (*Lama glama*) carcasses marketed in three key regions of Peru: Lima, Junín, and Huancavelica

(Bartl et al., 2023). These regions are not only pivotal for livestock production but also showcase diverse environmental and commercial dynamics, which may influence the epidemiology of sarcocystosis (Ayala Vargas, 2018). Additionally, this research explores the association between the prevalence of *Sarcocystis* sp., and specific commercial markets, as well as the distribution of cysts across different muscle groups, providing a nuanced understanding of this parasitic disease. In recent years, the province of Huancayo has experienced a marked increase in the consumption of alpaca and llama meat (Caulfield et al., 2022). However, the scarcity of authorized slaughterhouses and suboptimal sanitary conditions in animal processing has created significant information gaps. These include limited data on the prevalence of *Sarcocystis* sp., as well as the economic losses associated with carcass seizures due to parasitic contamination.

In this context, the objective of this study was to determine the prevalence of *Sarcocystis* sp. in llama and alpaca carcasses marketed in Huancayo. Furthermore, it sought to evaluate the economic implications of carcass seizures caused by this parasitosis. The findings aim to provide valuable insights for the better management of camelid resources and to support the implementation of effective strategies for controlling this parasitic disease.

MATERIALS AND METHODS

Area study

The study was conducted in three markets located in the city of Huancayo, situated in the southern part of the Mantaro Valley at an altitude of 3,200 meters above sea level. The selected markets were Centro Comercial Ferrocarril ($12^{\circ}4'18.80''S$, $75^{\circ}12'17.47''W$), Nueva Esperanza ($12^{\circ}4'12.28''S$, $75^{\circ}12'16.35''W$), and Nazaret ($12^{\circ}4'12.94''S$, $75^{\circ}12'16.18''W$) (Senamhi, 2023). These markets primarily offer llama and alpaca carcasses, which are sourced from various regions, including Lima, Huancavelica, and Junín. The climate of the study area is characterized by an average annual temperature of $11^{\circ}C$ and a total annual rainfall of 625 mm. A map indicating the location of the markets is presented in Figure 1.

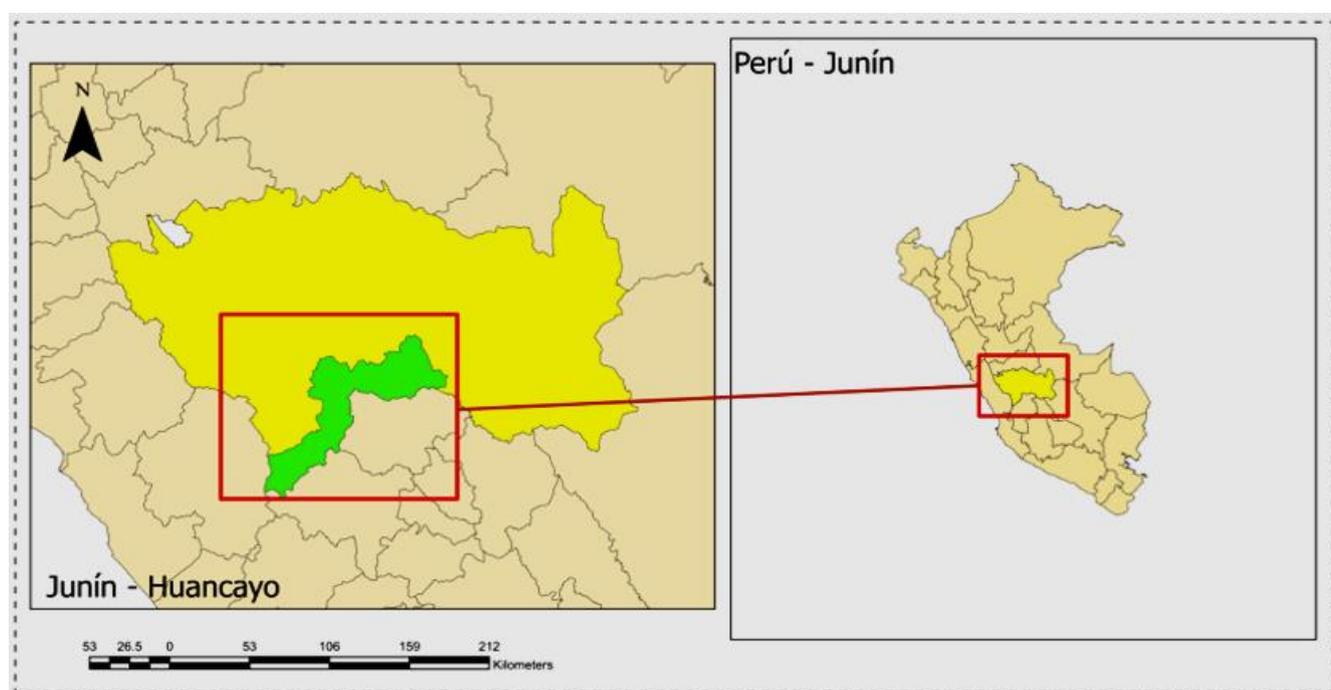


Figure 1- Location of the study

Samples

During the period from January to October 2023, a total of 2211 carcasses were collected, of which 1716 corresponded to alpacas and 495 to llamas. These were from 3 regions: Huancavelica (181/alpaca and 95/llama), Junín (820/alpaca and 108/llama), and Lima (715/alpaca and 292/llama). These carcasses from the 3 regions were received by 3 markets: Centro Comercial Ferrocarril ($n = 572$ alpaca, with 223, 126, and 223 for the neck, shoulder, and leg muscles, respectively; and 160 llama, with 52, 61, and 47 for the same muscles, respectively), Nazareth ($n = 482$ alpaca, with 154, 149, and 179 for the neck, shoulder, and leg muscles, respectively; and 182 llama, with 58, 68, and 56 for the same muscles, respectively), and Nueva Esperanza ($n = 662$ alpaca, with 249, 168, and 245 for the neck, shoulder, and leg muscles, respectively; and 153 llama, with 53, 52, and 48 for the same muscles, respectively). This collection was carried out in collaboration with the sanitary control personnel from the Bromatology area of the Provincial Municipality of Huancayo.

Parasitological analysis

Macroscopic examinations were conducted on the neck, shoulder, and legs of each camelid (alpacas and llamas) to assess infection rates of macroscopic cysts (Regensburger et al., 2015). The detection of cysts involved meticulous and thorough visual inspections of the carcasses, ensuring accuracy and consistency in observations (Apaza Jimenez and Chipana Mendoza, 2021). These examinations were performed in designated markets and at the processing center managed by the National Agricultural Health Service (SENASA), adhering to standardized protocols to ensure the reliability of the results.

Statistical analysis

The prevalence of macrocysts in alpacas and llamas were calculated using the equation:

$$Prevalence (\%) = \frac{\# \text{ positive cases}}{\text{Total number of individuals}} \times 100$$

A generalized linear mixed model (GLMM) with binomial distribution was applied, with the response variable being the presence of the parasite (macrocysts). Random effect variables included Species (Alpaca and Llama), Region (Huancavelica, Junín and Lima) and arm (BRAZO), neck (CUELLO) and leg (PIERNA) muscles. Additionally, the logit function was used to calculate Odds Ratios (OR), providing a measure of association between the variables of interest. All statistical analyses and calculations were performed using Excel (Microsoft Office® v2013) and the open-source software R (Team et al., 2018), using the packages descTools, ROCR, agricolae and stats to ensure accurate and reproducible results.

RESULTS AND DISCUSSION

Table 1 presents the total number of carcasses examined per camelid. The results showed the presence of *Sarcocystis* sp. in both species, with a prevalence of 8.0% (138 carcasses) for alpacas and 21.0% (104 carcasses) for llamas. The results of this study provide valuable insights into the prevalence and distribution of *Sarcocystis* sp. in alpaca and llama carcasses, emphasizing species, regional, and market-specific differences, as well as muscle-specific predilections. These findings highlight critical epidemiological patterns and their implications for food safety and public health. Table 2 shows that the highest prevalence of sarcocystosis in alpacas was recorded in the Lima region (9.7%) with 69 out of 715 carcasses, followed by Huancavelica (7.7%) with 14 out of 181 carcasses, and Junín (6.7%) with 55 out of 820 carcasses. Similarly, concerning llamas, the Lima Region exhibited the highest prevalence of sarcocystosis (33.9%) with 72 out of 212 carcasses, followed by Huancavelica (14.7%) with 14 out of 102 carcasses, and Junín (9.9%) with 18 out of 181 carcasses. A significant difference ($p < 0.05$) in the prevalence of sarcocystosis was observed between Lima and Junín, as well as between Lima and Huancavelica in the case of alpacas. Similarly, significant differences ($p < 0.05$) in the prevalence of sarcocystosis were found between the Huancavelica region and the Lima and Junín regions in llama carcasses. It was observed that the regions of Lima are the most at-risk places for consumption. These results are similar to those reported by Castro and Leguía (1992) in Lima and by Santiago and Leguía (2018) in the Junín region. These findings highlight that, despite being recognized as prominent livestock areas, the measures implemented for the management and prevention of parasitosis have not yielded satisfactory results in terms of food safety. In this situation, it is imperative to prioritize control efforts, such as health education and implementing authorized slaughterhouses for more effective resource management, thus ensuring food safety and public health.

Table 1 - Prevalence of *Sarcocystis* sp. in alpaca and llama carcasses.

Carcasses	Total examined	Positive infected	
		N	%
Alpaca	1716	138	8.0
Llama	495	104	21.0

Table 2 - Prevalence of *Sarcocystis* sp. in alpaca and llama carcasses.

Animal	Region	Total	Infected	%
Alpaca	Huancavelica	181	14	7.7 ^a
	Junin	820	55	6.7 ^a
	Lima	715	69	9.7 ^a
P-value				NS
Llama	Huancavelica	102	14	14.7 ^a
	Junín	181	18	9.9 ^a
	Lima	212	72	33.9 ^b
P-value				**

Equal letters in the same column do not differ significantly ($p > 0.05$).

Table 3 - Prevalence of *Sarcocystis* sp. in alpaca and llama carcasses.

Variable		N	Odds ratio	References	P values
SPECIES	ALPACA	1716	■	Ref.	
	LLAMA	495	■	2.82 (2.06, 3.84)	<0.001
REGION	HUANCAVELICA	276	■	Ref.	
	JUNIN	928	■	1.45 (0.85, 2.52)	0.179
	LIMA	1007	■	3.79 (1.68, 8.69)	0.001
MARKET	FERRO	732	■	Ref.	
	NAZA	664	■	0.38 (0.22, 0.63)	<0.001
	NESPE	815	■	0.25 (0.12, 0.51)	<0.001
MUSCLES	ARM	624	■	Ref.	
	NECK	789	■	1.20 (0.81, 1.79)	0.364
	LEG	798	■	1.65 (1.17, 2.35)	0.005

The OR (Odds ratio) is a statistical indicator that measures the probability of an event occurring in one group compared to another (Reference).

In Table 3, the Odds Ratios (OR) are observed, with the Llama species obtaining a value of 2.82, indicating that llamas have a higher chance of the presence of *Sarcocystis* sp. macrocysts compared to Alpacas. Regarding the region, Lima shows the highest chance of reporting macrocysts, reaching an OR of 3.79. This suggests a significantly higher presence compared to other regions. The higher prevalence of *Sarcocystis* sp. in llamas (21.0%) compared to alpacas (8.0%) reflects significant differences in susceptibility between the two species. This discrepancy may be attributed to llamas' greater exposure to definitive hosts, such as domestic and wild canines, and their distinct grazing behaviors, which may increase the risk of infection (Rosenthal, 2021). Additionally, management practices, herd density, and environmental conditions likely play a role in the higher parasitic burden observed in llamas. These findings align with existing literature, which suggests that llamas are often more vulnerable to parasitic infections due to less intensive management systems compared to alpacas (Wu et al., 2022). A notable finding of this study is the significant regional disparity in the prevalence of *Sarcocystis* sp. Lima exhibited the highest prevalence among regions, with an OR of 3.79 compared to other locations, such as Junín and Huancavelica. This suggests that factors specific to Lima, including urbanization, higher population density, and suboptimal slaughterhouse conditions, may contribute to the elevated risk of contamination, an idea also shared by Rene et al. (2019). Moreover, the environmental conditions in Lima, characterized by more intensive camelid trade and market activity, likely exacerbate the exposure of animals to parasitic contamination (Raymond et al., 2020). The lower prevalence observed in Junín and Huancavelica may be explained by differences in climatic conditions, such as cooler temperatures and reduced rainfall, which may limit the survival of *Sarcocystis* sp. in the environment (Baitzel et al., 2022). The results also highlight significant differences in prevalence within regions for alpacas and llamas. For instance, Lima showed a prevalence of 9.7% in alpacas and 33.9% in llamas, compared to Junín, which had lower rates for both species. These regional variations underscore the need for tailored interventions that account for the specific epidemiological and environmental characteristics of each location (Jauregui et al., 2024).

The prevalence of *Sarcocystis* sp. also varied significantly among markets (Table 3). The Ferrocarril market showed the highest risk of contamination, serving as the reference group with which other markets were compared. Conversely, the Nazareth and Nueva Esperanza markets demonstrated significantly lower odds of *Sarcocystis* sp. presence, with ORs of 0.38 and 0.25, respectively. These differences may be attributed to variations in sourcing practices, transportation conditions, and sanitary measures implemented at each market (Fernandez-F et al., 2022; Rodríguez et al., 2023). Markets with better infrastructure and stricter quality control measures are likely to exhibit lower prevalence rates of parasitic infections (Baitzel et al., 2022). The findings suggest that market-specific factors, such as hygiene standards and meat handling protocols, play a critical role in determining the risk of contamination. The anatomical distribution of *Sarcocystis* sp. cysts revealed a preferential localization in the leg (OR = 1.65) and neck (OR = 1.20) compared to the shoulder. This finding is consistent with previous studies that report a higher prevalence of macrocysts in muscle groups with greater vascularization and proximity to infected tissues (Regensburger et al., 2015). The preferential accumulation of cysts in the leg and neck muscles may reflect physiological differences in blood flow or tissue composition, which facilitate parasite development and cyst formation (Lucas et al., 2019; Wu et al., 2022). From a practical standpoint, this information is essential for meat inspection processes, as it highlights the importance of focusing on high-risk anatomical sites during routine examinations.

CONCLUSION

The research reveals significant data on the prevalence of *Sarcocystis* sp. in alpacas and llamas, highlighting a higher presence in llamas despite their smaller number of examined samples. This finding suggests a potential risk to public

health, especially in areas like Lima, where the prevalence is notably high. The results also underscore the importance of addressing risk factors such as extreme weather conditions and commercial practices that may influence the spread of parasitosis. There is a clear need to implement effective control and resource management measures to ensure food safety and public health in these regions. These findings provide a solid foundation for future research and preventive actions in the region.

DECLARATIONS

Corresponding author

Correspondence and requests for materials should be addressed to Jordan Ninahuanca Carhuas. E-mail: jninahuanca@uncp.edu.pe; ORCID: <https://orcid.org/0000-0002-0137-0631>

Authors' contribution

Edgar Garcia-Olarte: Execution of the research; Jordan Ninahuanca Carhuas: Statistical analysis and editing; Wilder Suarez-Reynoso and Wilhelm Guerra Condor: laboratory analysis; Yakelin Maurico-Ramos: data collection; María Flores Guillen: macroscopic analysis of cysts; Ide Unchupaico Payano: macroscopic analysis of cysts; Armando Aquino Tacza: carcasses non-monitoring.

Acknowledgments

The authors would like to thank the staff involved, the laboratory employees.

Ethical approval

The procedures and ethics of this research work were based on the international and national guidelines for the care and use of animals in the scientific research.

Consent to publish

All authors agree to the publication of this manuscript.

Competing interests

The authors have not declared any competing interest.

REFERENCES

- Amalfitano G, Petrigh R, Loos J and Fugassa M (2017). New parasites information to camelids increase the knowledge about Cerro Casa de Piedra 7 archaeological site, Santa Cruz province, Argentina. *Anales del Instituto de la Patagonia. Universidad de Magallanes*, pp. 101-108. <https://www.cabidigitalibrary.org/doi/full/10.5555/20183079497>
- Apaza Jimenez YK and Chipana Mendoza GJ (2021). Types of methodologies for the diagnosis of *Trichinella spiralis* in pork (*Sus scrofa domestica*). *Revista Estudiantil Agro-Vet* 5: 49. <https://agrovet.umsa.bo/index.php/AGV/article/view/61>
- Ayala Vargas C (2018). Sarcocistiosis (Arrocillo, Falsa triquina, Falso cisticercos, Sarcosporidiosis); Revisión literaria. *Revista de Investigación e Innovación Agropecuaria y de Recursos Naturales* 5(ESPECIAL): 193-206. http://www.scielo.org.bo/pdf/riarn/v5nEspecial/v5_a21.pdf
- Baitzel SI, y La Borda MP, Konecky BL, Sae-Lim J and Rivera Infante AF (2022). RETRACTED ARTICLE: Pastoral Paleoclimate Palimpsests of the South-Central Andes: High-Altitude Herder Dwellings in the 2nd Millennium ad. *Journal of Field Archaeology* 47(5): 341-359. <https://doi.org/10.1080/00934690.2022.2072161>
- Bartl K, Mogrovejo P, Dueñas A and Quispe I (2023). Cradle-to-grave environmental analysis of an alpaca fiber sweater produced in Peru. *Science of The Total Environment* 905: 167023. <https://doi.org/10.1016/j.scitotenv.2023.167023>
- Castro J and Leguía G (1992). Prevalence of *Sarcocystis* sp. in cattle, sheep and goats slaughtered in Lima's animal feedlots. *Revista Peruana de Biología* 4(1-2): 21-24. <https://doi.org/10.15381/rpb.v4i1-2.8336>
- Catoa RM, Gallegos R, Mamani TH and García JPGJRIA (2016). Estructura Genética de la Población de Llamas (*Lama glama* del Banco de Germoplasma del Instituto Nacional de Innovación Agraria-Perú. 18(1): 55-60. <http://doi.org/10.1827/ria.2016.178>
- Caulfield ME, Vanek SJ, Meza K, Huaraca J, Loayza JL, Palomino S, Olivera E, Ccanto R, Scurrah M and Vigil LJE (2022). Drivers of farmer involvement in experimental forage trials in the Peruvian Andes and implications for participatory research design. 58: e39. <https://doi.org/10.1017/S0014479722000357>
- Coleman J, Berry D, Pierce K, Brennan A and Horan B (2010). Dry matter intake and feed efficiency profiles of 3 genotypes of Holstein-Friesian within pasture-based systems of milk production. *Journal of Dairy Science*, 93(9): 4318-4331. <https://doi.org/10.3168/jds.2009-2686>
- Condori-Quispe R, Loza-Murguía MG, Gutiérrez-Ramírez L and Condori-Condori C (2019). Prevalence of *Sarcocystis* spp. in cardiac muscle of llamas (*Lama glama*) and alpacas (*Vicugna pacos*). *Journal of the Selva Andina Anima* 6(2): 39-46. <https://doi.org/10.36610/j.jsaas.2019.060200039>

- Estremadoyro LJG, Salome PH, Carhuas JN, Guzman SO, Tacza AA, Guillen MAF and Garcia-Olarte E (2024). Effects of Different Seasons on Milk Quality: A Study on Two Cattle Breeds in Rainy and Drought Contexts. *World's Veterinary Journal* 14(2): 213-219. <https://doi.org/10.54203/scil.2024.wvj26>
- Fayer R, Esposito DH and Dubey JP (2015). Human infections with *Sarcocystis* species. *Clinical microbiology reviews* 28(2): 295-311. <https://doi.org/10.1128/cmr.00113-14>
- Fernandez-F F, Gutiérrez-A R, Pacheco-S V, Chirinos-T J, Lombardo DM, Olivera LV, Bernabe-Ortiz JC and López-Casaperalta P (2022). Determination of *Sarcocystis lamacanis* microcysts in the cardiac muscle of alpacas (*Vicuña pacos*) and their correlation with troponin cTnl. A study performed in the high Andean region of southern Peru. *Veterinary and Animal Science* 18: 100270. <https://doi.org/10.1016/j.vas.2022.100270>
- Gareh A, Soliman M, Saleh AA, El-Gohary FA, El-Sherbiny HM, Mohamed RH and Elmahallawy EK (2020). Epidemiological and histopathological investigation of *Sarcocystis* spp. in slaughtered dromedary camels (*Camelus dromedarius*) in Egypt. *Veterinary Sciences* 7(4): 162. <https://doi.org/10.3390/vetsci7040162>
- Jauregui Z, Salas-Fajardo MY, Puicón V and Lucas JR (2024). Prevalence and distribution pattern of *Sarcocystis* spp. in slaughtered cattle from the Peruvian tropical Andes, Peru. *Veterinary Parasitology: Regional Studies and Reports* 48: 100990. <https://doi.org/10.1016/j.vprsr.2024.100990>
- Lindsay DS and Dubey JP (2020). Neosporosis, toxoplasmosis, and sarcocystosis in ruminants: an update. *Veterinary Clinics: Food Animal Practice* 36(1): 205-222. <https://doi.org/10.1016/j.cvfa.2019.11.004>
- Lucas JR, Barrios-Arpi M, Rodríguez J, Balcázarnakamatsu S, Zarria J, Namiyama G, Taniwaki N and Gonzales-Viera O (2019). Ultrastructural description of *Sarcocystis* sp. in cardiac muscle of naturally infected alpacas (*Vicuña pacos*). *Iranian Journal of Parasitology* 14(1): 174-179. <https://doi.org/10.18502/ijpa.v14i1.733>
- Raymond C, Horton RM, Zscheischler J, Martius O, AghaKouchak A, Balch J, Bowen SG, Camargo SJ, Hess J and Kornhuber K (2020). Understanding and managing connected extreme events. *Nature Climate Change* 10(7): 611-621. <https://doi.org/10.1038/s41558-020-0790-4>
- Regensburger CD, Gos ML, Ctibor J and Moré GA (2015). Morphological and molecular characteristics of *Sarcocystis aucheniae* isolated from meat of guanaco (*Lama guanicoe*). available online: <https://ri.conicet.gov.ar/handle/11336/55022>
- Rene C-Q, Gregorio L-MM, Luis G-R and Cirilo C-C (2019). Prevalence of *Sarcocystis* spp. in cardiac muscle of llamas (*Lama glama*) and alpacas (*Vicuña pacos*). *Journal of the Selva Andina Animal Science* 6(2): 39-46. <https://doi.org/10.36610/jjsaas.2019.060200039>
- Rodríguez A, Quispe-Solano M, Rodríguez J-L and Lucas JR (2023). The occurrence of *Sarcocystis* spp. in the myocardium of alpacas (*Vicuña pacos*) with associated risk factors in the Peruvian Andes. *Tropical Animal Health and Production* 55(2): 66. <https://doi.org/10.1007/s11250-023-03498-3>
- Rosenthal BM (2021). Zoonotic *Sarcocystis*. *Research in veterinary science* 136: 151-157. <https://doi.org/10.1016/j.rvsc.2021.02.008>
- Santiago B and Leguía G (2018). Prevalencia de *Sarcocystis* en alpacas (*Lama pacos*) y en perros pastores de una ganadería de la Sierra central del Perú. *Biotempo* 15(1): 59-62. <https://doi.org/10.31381/biotempo.v15i1.1696>
- Senamhi (2023). Hydrometeorological Data at the national level in Peru. Datos Hidrometeorológicos a nivel nacional en Perú. Retrieved 28 de abril, 2023, from <https://www.senamhi.gob.pe/?p=estaciones>
- Shams M, Shamsi L, Asghari A, Motazedian MH, Mohammadi-Ghalehbin B, Omidian M, Nazari N and Sadrebazzaz A (2022). Molecular epidemiology, species distribution, and zoonotic importance of the neglected meat-borne pathogen *Sarcocystis* spp. in cattle (*Bos taurus*): a global systematic review and meta-analysis. *Acta parasitologica* 67(3): 1055-1072. <https://doi.org/10.1007/s11686-022-00563-z>
- Team RC, Team MRC, Suggests M and Matrix S (2018). Package stats. The R Stats Package. <https://prs.ism.ac.jp/~nakama/Rjp/stats-manual.pdf>
- Valentine BA and Martin JM (2007). Prevalence of neoplasia in llamas and alpacas (Oregon State University, 2001-2006). *Journal of Veterinary Diagnostic Investigation* 19(2): 202-204. <https://doi.org/10.1177/104063870701900213>
- Wu Z, Sun J, Hu J, Song J, Deng S, Zhu N, Yang Y and Tao J (2022). Morphological and Molecular Characterization, and Demonstration of a Definitive Host, for *Sarcocystis masoni* from an Alpaca (*Vicuña pacos*) in China. *Biology* 11(7): 1016. <https://doi.org/10.3390/biology11071016>
- Yang Z-Q, Wei C-G, Zen J-S, Song J-L, Zuo Y-X, He Y-S, and et al. (2005). A taxonomic re-appraisal of *Sarcocystis nesbitti* (Protozoa: Sarcocystidae) from the monkey *Macaca fascicularis* in Yunnan, PR China. *Parasitology international* 54(1): 75-81. <https://doi.org/10.1016/j.parint.2004.12.004>

Publisher's note: Scienceline Publication Ltd. remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access: This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <https://creativecommons.org/licenses/by/4.0/>.

INFLUENCE OF FEATHER GENOTYPE, STORAGE DURATION AND TEMPERATURE ON THE EXTERNAL AND INTERNAL QUALITIES OF CHICKEN TABLE EGGS

Dawolor Nusue KANASUAH¹ , Kwaku ADOMAKO¹ , Bernard Ato HAGAN²  and Oscar Simon OLYMPIO¹ 

¹Department of Animal Science, Kwame Nkrumah University of Science and Technology, Kumasi, Ghana

²Department of Animal Production and Health, School of Agriculture and Technology, University of Energy and Natural Resources, Sunyani, Ghana

✉ Email: bernard.hagan@uenr.edu.gh

➤ Supporting Information

ABSTRACT: A study was carried out to determine the influence of the feather genotype, storage duration, temperature and method on the internal and external qualities of chicken table eggs. A total of 864 table eggs collected from naked neck (*Nanaff*), frizzle (*nanaFf*) and normal feathered (*nanaff*) birds were used in the study. A Completely Randomized Design of four factors namely, feather genotypes, storage temperatures (5°C and 26°C), storage duration (0, 7, 14, 21 and 28 days) and storage methods (with or without vegetable oil application) was used. The GLM procedure of GenStat (17th Edition) was used to determine the effects of the four factors and their interactions on external qualities (egg weight, length, and width, shell weight and thickness) and internal qualities (albumen height and weight, yolk height, weight, diameter and colour and Haugh unit) of table eggs. The effect of chicken genotype on proximate composition and nutritional values of table eggs were also determined. Feather genotype had significant ($P < 0.05$) effect on yolk colour and weight whilst storage duration, temperature and method had significant ($P < 0.05$) effects on all the internal qualities of eggs studied except effect of storage duration on yolk colour. The 2-way and 3-way interactions of the factors studied were important sources of variation for many of the internal qualities of eggs studied. With the exception of storage temperature, the other factors studied had significant ($P < 0.05$) effects on many of the external qualities of eggs. The interactions of the factors were not significant ($P > 0.05$) sources of variation for most of the external qualities of eggs. Mutant feather genes (*Na* and *F*) positively influence egg qualities which could be utilised to segment the commercial chicken egg market.

Keywords: Feather, Frizzle, Naked neck, Nutritional value, Yolk colour,.

INTRODUCTION

Eggs contain nutrients which are essential for improving human health. Proper functioning of the body is impeded if essential amino acids, which are the main nutrients in eggs, are lacking. Chicken egg albumen and yolk are reported to contain essential amino acids (Ali et al., 2019; Attia et al., 2020). The United States Department of Agriculture (USDA) (2008) reported that eggs are rich foundations of minerals and vitamins.

A complete egg is composed of parts such as the shell (exterior and interior shell membranes), albumen, air cell, cuticle or bloom, chalazae, germinal disk, nucleus of pander, yolk and vitelline membrane (EDINFORMATICS, 2013). Egg quality is built around a number of traits including albumen height, albumen weight, yolk height, yolk diameter, yolk index, yolk weight, shell ratio, shell thickness, shell weight, egg length, egg weight, egg width and Haugh Unit (Murshed and Qaid, 2024). Several studies have reported significant and positive relationships among egg quality parameters of poultry (Zhang et al., 2005; Inca et al., 2020; Guni et al., 2021). Farooq et al. (2001a) reported positive and significant relationships among egg weight, egg width and egg length for eggs from Fayoumi birds. Similarly, largely positive correlations were reported among egg quality traits of two layer chicken breeds in South Africa (Tyasi et al., 2022). In Japanese quails, Farooq et al. (2001b) reported that there were positive correlations among shell weight, egg weight and shell thickness of quail eggs. Several external factors such as cleanliness, freshness, egg weight and shell weight are important for consumers' acceptability of eggs (Hamilton, 1982; Sonaiya and Swan, 2004; Batkowska et al., 2023). Internal characteristics such as yolk index, Haugh Unit and chemical composition are also important in poultry breeding because of their influence on growth of chicks, breeding performance and egg quality for consumption (Yahaya et al., 2021). The external and internal quality traits of eggs of hens have influence on the hatchability of fertile eggs, and the weight and development of chicks (Şahan et al., 2014; Iqbal et al., 2016; Hegab and Hanafy, 2019).

The external and internal egg qualities are also influenced by storage duration and storage temperature. Eggs stored at low temperature maintain better egg quality (Samli et al., 2005). Egg weight, shell weight, albumen height, albumen viscosity, Haugh Unit and yolk colour decreased with increasing storage temperature of hens (Lee et al., 2016; Martínez et al., 2021). Eggs maintain qualities better when stored for a short period of time (Jin et al., 2011). Prolonged length of egg

RESEARCH ARTICLE
 PII: S222877012500004-15
 Received: August 06, 2024
 Revised: January 24, 2025
 Accepted: January 25, 2025

storage deteriorates egg quality of chicken eggs (Nasri et al., 2019; Melo et al., 2020). Tebesi et al. (2012) reported that eggs were able to maintain higher yolk height when stored within 7 days.

The naked neck (*Na*) and frizzle (*F*) genes are two mutant thermoregulatory genes that aid chickens to adapt to high ambient temperatures in, especially, the tropics (Asumah et al., 2022). Layer chickens carrying the *Na* or *F* alleles have been reported to record higher percentage of fertile eggs (Asumah et al., 2022), increased egg production (Fathi et al., 2013; Adomako et al., 2014) and improved egg shell quality (Salahuddin and Howliger, 1991). However, El-Rahman and Makled (2006) reported reduction in shell quality in birds carrying *Na* alleles compared to birds with only *na* alleles.

Whilst there have been several studies on egg quality traits of frizzle, naked neck and normal feathered birds in Sub-Saharan Africa and other parts of the world (Salahuddin and Howliger, 1991; Abou-Emera et al., 2017; Fathi et al., 2022), these studies have barely focused on the nutrient contents of the eggs produced by the birds. In addition, information is scanty on the interactions between feather genotype and egg storage methods on the egg quality traits of chicken eggs. The objective of this study therefore was to determine the influence of feather genotype, storage duration, storage temperature and storage method on external and internal egg quality characteristics, amino acid profile and proximate composition of chicken table eggs.

MATERIALS AND METHODS

Location and duration

The research was conducted at Akate Farms and Trading Company Limited (AFTC) at Saaman, Kumasi, Ghana and the Department of Animal Science, Kwame Nkrumah University of Science and Technology within a period of six months.

Experimental birds and eggs

The experimental birds kept at the AFTC were offspring of crosses between naked neck and frizzle feathered cocks and hybrid commercial Lohmann hens. The naked neck and frizzle feathered, both heterozygotes, were bred with normal feathered Lohmann Brown classic layers in two separate matings to produce offspring which were heterozygous naked neck, heterozygous frizzle feathered and normal feathered chickens in the first filial (F₁) generation. Eight hundred and sixty-four (864) table eggs were collected from the naked neck (*Nanaff*), frizzle (*nanaFf*) and normal feathered (*nanaff*) layer chickens (288 per genotype) kept as experimental birds by AFTC, Kumasi, Ghana. The layer birds were 28 weeks old at the start of the experiment. The external and internal egg qualities were determined after collection, using the procedures described by Fayeye et al. (2005).

Experimental design

A Completely Randomized Design in a 3x2x5x2 factorial was applied. Eggs were obtained from three genotypes being *Nanaff*, *nanaFf* and *nanaff*, stored at two storage temperatures (26°C and 5°C) for four storage durations with a control of 0 days (0, 7, 14, 21 and 28 days) using two storage methods (with or without the application of vegetable oil to the egg shells). For eggs which received oil treatment, Sunny vegetable oil manufactured in Ghana was applied by immersion.

The experiment was conducted in three phases and eggs were collected from the chicken genotypes which were housed in deep litter pens. The three chicken genotypes were placed into nine different pens, with each bird genotype put into three different pens labelled as treatments (T₁, T₂ and T₃) with about 20 birds in each pen. A total of 864 table eggs from the three genotypes were further used in the study with 288 table eggs obtained from each genotype.

Parameters studied and their measurement

- a. The egg width and length was measured using a pair of vernier calipers in centimetres.
- b. Egg weight was measured with a digital electric balance in grams.
- c. Egg shell thickness was measured with a micrometer screw gauge in mm. Shell thickness was calculated from the average of three measurements taken at the middle, broad end and the small end of the eggs.
- e. Yolk diameter was measured with a vernier caliper in centimeters.
- f. Yolk colour was determined with the DSM yolk colour fan (formerly Roche Yolk Color Fan). Higher figures indicate deeper yolk colour while lower figures indicate lighter yolk colour.
- g. Yolk weight was determined with a digital weighing scale in grams.
- h. Yolk height was determined by the use of a tripod spherometer.
- i. Albumen weight was also determined by the use of a digital weighing scale.
- j. Albumen height was determined with a tripod spherometer in mm.
- k. Egg weight loss was determined by subtracting the final weight from the initial weight and expressed as a percentage.
- l. Haugh unit was determined using the formula, $HU = 100 \times \log (H + 7.57 - 1.7W^{0.37})$ introduced by Haugh (1937). where HU = Haugh Units; H = Observed albumin height (mm); W = Observed weight of egg (g) (Roush, 1981).

Twenty-four table eggs from each of the three genotypes were analysed on each storage period (0, 7, 14, 21 or 28), storage method (with or without vegetable oil application).

Proximate composition of the eggs from the three genotypes was determined by drying egg samples (albumen and yolk) in an oven at 65°C for 72 hours. The dried samples were transferred to the Crops and Soil Science Laboratory, Faculty of Agriculture, Kwame Nkrumah University of Science and Technology for the proximate composition analyses. The nutritional values of egg albumen from the three genotypes were analyzed by Evonik Nutrition South Africa Limited. Similar to the proximate analysis, the albumen was also dried in an oven at 65°C for 72 hours and later transferred to South Africa for the amino acid profile analyses.

Data analysis

The data on external, internal quality characteristics of eggs, and proximate composition and amino acid profiles of the albumen using the general linear model procedure of GenStat (17th Edition). The model used for the analysis of the data collected is presented below.

$$Y_{ijklm} = \mu + G_i + T_j + D_k + M_l + GT_{ij} + GD_{ik} + GM_{il} + TD_{jk} + TM_{jl} + DM_{kl} + GTD_{ijk} + GTM_{ijl} + TDM_{jkl} + e_{ijklm}$$

Where Y_{ijklm} = measured or calculated variables;

μ = overall mean;

G_i = fixed effect of the i^{th} chicken genotype (naked neck, frizzle or normal feathered);

T_j = fixed effect of the j^{th} storage temperature (26°C and 5°C);

D_k = fixed effect of the k^{th} egg storage duration (0, 7, 14, 21 or 28 days);

M_l = fixed effect of the l^{th} egg storage method (with or without cooking oil treatment);

GT_{ij} = fixed interaction of the i^{th} genotype and the j^{th} storage temperature;

GD_{ik} = fixed interaction of the i^{th} genotype and the k^{th} storage duration;

GM_{il} = fixed interaction of the i^{th} genotype and the l^{th} storage method;

TD_{jk} = fixed interaction of the j^{th} storage temperature and the k^{th} storage duration;

TM_{jl} = fixed interaction of the j^{th} storage temperature and the l^{th} storage method;

DM_{kl} = fixed interaction of the k^{th} storage temperature and the l^{th} storage method;

GTD_{ijk} = fixed interaction of the i^{th} genotype, j^{th} storage temperature and the k^{th} storage duration;

GTM_{ijl} = fixed interaction of the i^{th} genotype, j^{th} storage temperature and the l^{th} storage method;

TDM_{jkl} = fixed interaction of the j^{th} storage temperature, k^{th} storage duration and l^{th} storage method;

e_{ijklm} = random error term associated with each observation $\sim N(0, \sigma_e^2)$ where σ_e^2 is residual variance.

Differences between means were separated using Tukey's Test at 5% probability level.

RESULTS AND DISCUSSION

Internal qualities of table eggs as influenced by genotype

The effect of chicken genotype on internal egg qualities of table eggs are presented in Table 1. There were no significant differences ($P>0.05$) among various genotypes in relation to albumen height, albumen weight, yolk diameter, Haugh unit and yolk height. The absence of significant differences for these parameters agrees with the findings of Rajkumar et al. (2009) who observed no significant differences in albumen height, albumen weight, yolk height (at 28 weeks old) and Haugh unit for NaNa, Nana and nana chicken genotypes in India. Udoh et al. (2012) also reported no significant difference ($P>0.05$) among three local genotypes in terms of yolk weight, albumen height and yolk height in Nigeria. Frizzle genotype recorded significantly ($P<0.05$) heavier yolk weight than normal feathered genotype, with the normal feathered showing the lowest value in this trait. The higher yolk weight for the frizzle eggs could probably be due to their efficient feed conversion ratio in converting protein for feather production into their eggs. The heavier yolk weight of eggs from frizzle feathered genotype in this study is contrary to the report of Yakubu et al. (2008) who reported that naked neck chicken eggs had heavier yolk weight compared to eggs from normal and frizzled feathered birds. However, Rajkumar et al. (2009) recorded a significantly ($P<0.05$) heavier yolk weight for normal feathered birds than naked neck ones in India, and noted that lower yolk weight in naked neck birds indicated lower fat percentage in these birds than their normal feathered counterparts. Non-significant ($P>0.05$) effect of feather genotype on yolk weight has also been reported by Udoh et al. (2012) and Ogundero et al. (2019) in Nigerian local indigenous chickens.

The yolk colour for naked neck and the normal feathered bird eggs were not significantly ($P>0.05$) different but both were significantly ($P<0.05$) different from the frizzle hens which recorded a lower yolk colour value (Table 1). Islam et al. (2011) recorded higher yolk colour values from Bangladesh naked neck chicken which is in agreement with the results of the current study. However, Rajkumar et al. (2009) reported higher yolk colour in normal feathered (8.00) and naked neck (7.49) than observed in the present findings. Yolk colour is probably controlled mainly by nutrition than genetics (Grashorn, 2016), hence the varying results for yolk colour as influenced by feather genotype in literature and this study.

Table 1 - Internal quality characteristics of table eggs as influenced by chicken feather genotype, egg storage duration, egg storage temperature and storage method

Items	Albumen height (mm)	Albumen weight (g)	Haugh unit (%)	Yolk colour	Yolk diameter, (cm)	Yolk height, (mm)	Yolk weight, (g)
Genotype							
<i>Nanaff</i>	4.52	33.74	60.36	4.94 ^a	4.21	13.38	17.37 ^a
<i>nanaFf</i>	4.52	33.69	60.47	4.52 ^b	4.23	12.88	17.48 ^a
<i>nanaff</i>	4.43	32.30	59.92	4.93 ^a	4.22	13.18	17.15 ^b
SEM	0.092	0.295	0.737	0.098	0.019	0.013	0.098
<i>P-value</i>	0.544	0.512	0.855	0.003	0.822	0.116	0.042
Storage duration (days)							
0	4.82 ^a	34.57 ^a	62.25 ^a	4.99	4.05 ^c	14.62 ^a	17.36
7	4.49 ^b	34.28 ^{ab}	60.94 ^b	4.80	4.19 ^c	13.15 ^b	17.33
14	4.44 ^b	33.75 ^b	59.95 ^c	4.83	4.23 ^b	12.91 ^c	17.25
21	4.40 ^b	32.58 ^c	59.80 ^c	4.60	4.25 ^b	12.93 ^c	17.31
28	4.31 ^c	32.70 ^c	58.30 ^d	4.76	4.41 ^a	12.13 ^d	17.38
SEM	0.084	0.354	0.873	0.116	0.022	0.015	0.112
<i>P-value</i>	<0.001	<0.001	0.005	0.216	<0.001	<0.001	0.938
Storage temperature							
Refrigeration (0°C)	5.21 ^a	34.22 ^a	68.27 ^a	5.07 ^a	4.00 ^b	15.13 ^a	17.16 ^b
Room temperature (26°C)	3.77 ^b	32.93 ^b	52.32 ^b	4.52 ^b	4.46 ^a	11.15 ^b	17.51 ^a
SEM	0.247	0.059	0.617	0.082	0.015	0.144	0.079
<i>P-value</i>	<0.001	0.001	<0.001	<0.001	<0.001	<0.001	0.008
Storage method							
Vegetable oil	4.79 ^a	33.60 ^a	63.77 ^a	4.89 ^a	4.08 ^b	14.08 ^a	17.44 ^a
No oiling	3.85 ^b	32.86 ^b	54.00 ^b	4.55 ^b	4.39 ^a	11.75 ^b	17.15 ^b
SEM	0.059	0.247	0.617	0.116	0.015	0.144	0.079
<i>P-value</i>	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001

^{abcd} Means within the same sub-column with different subscripts are significant at P<0.05

Internal qualities of table eggs as influenced by storage duration

The effect of storage duration on the internal qualities of table eggs is presented in Table 1. Storage duration of eggs did not have any significant ($P>0.05$) effect on yolk colour and yolk weight. However, Jin et al. (2011) reported significant effect of storage time on yolk colour of laying hens at peak production. Duration of egg storage significantly ($P<0.05$) influenced albumen height, albumen weight, Haugh unit, yolk diameter and yolk height. Albumen height, albumen weight, Haugh unit and yolk height largely decreased with increase in length of egg storage. The importance of storage duration on albumen height in this study corroborates the findings of Raji et al. (2009) and Santos et al. (2019) who observed decline in albumen height with increase in storage length. Akinola and Ibe (2014) and Abioja et al. (2021) also reported similar findings to the present study. Tebesi et al. (2012), however, reported different findings with eggs stored for 14 days showing higher albumen height. Yolk diameter significantly ($P<0.05$) increased with increase in storage length. This could be due to the expansion of yolk as storage length increases. This finding agrees with results of Abioja et al. (2021) in FUNAAB- α chicken eggs. The reduction in yolk height with increase in storage length could be attributed to loss in moisture from the yolk resulting in shrinkage of the yolk. The current result agrees with Raji et al. (2009) and Tebesi et al. (2012) who reported higher yolk height for day 7. Similarly, the decline in Haugh unit with increase in storage duration is an indication of deterioration of egg quality. USDA (2000) reported that higher Haugh unit determines the protein content and freshness of eggs. Several authors (Rajkumar et al., 2009; Raji et al., 2009; Akinola and Ibe, 2014; Abioja et al., 2021) have presented results of higher Haugh unit with reduced storage duration of eggs which corroborate the findings from this work.

Internal qualities of table eggs as influenced by storage temperature

Results of the influence of storage temperature on internal egg quality is presented in Table 1. Albumen height, albumen weight, Haugh unit, yolk height, yolk colour, yolk diameter and yolk weight were significantly ($P<0.05$) affected by storage temperatures.

Albumen height was higher for eggs stored in a refrigerator than those stored under room temperature. This finding corroborates the results of Scott and Silversides (2000) and Samli et al. (2005) who observed increased albumen height for refrigerated eggs compared to eggs stored at room temperature. This could be attributed to the fact that eggs stored in a refrigerator maintain better albumen quality than those stored at room temperature. Refrigeration of eggs enhance the ability to retard carbon dioxide loss and breakdown of carbonic acid to carbon dioxide leading to the maintenance of egg quality (Qin et al., 2024). The heavier weight of the albumen for refrigerated eggs could be due to the prevention of evaporation of moisture from eggs stored in a refrigerator as a result of low temperature. The retention of Mucin fiber in the albumen of eggs stored in a refrigerator could have prevented the albumen from becoming watery and losing weight (Mountney, 1976). Khan et al. (2013) noted that albumen quality deterioration could be due to the effect of evaporation of moisture and carbon dioxide from the egg when stored under room temperature.

The mean yolk weight of eggs stored in a refrigerator was significantly heavier ($P<0.05$) than those stored at room temperature. This could be due to the retention of moisture in the yolk of eggs stored in a refrigerator. Samli et al. (2005) observed that there was a decrease in yolk weight with increase in storage temperature. Eggs stored at room temperature showed significantly lower yolk colour value (4.52) than eggs stored in a refrigerator (5.07) and this agrees with Jin et al. (2011) who reported significant effect of storage temperature on yolk colour. Yolk diameter also exhibited a significant difference ($P<0.05$) with eggs stored at room temperature showing higher yolk diameter than the ones stored in a refrigerator. Yolk height was significantly higher ($P<0.05$) for eggs stored in a refrigerator than eggs stored at room temperature. This result was similar to the finding of Raji et al. (2009) who recorded higher yolk height for eggs stored in a refrigerator compared to those stored at room temperature. Haugh unit showed a significant difference ($P<0.05$) with eggs stored in a refrigerator recording higher Haugh unit (68.27) than eggs stored at room temperature (52.32). The higher Haugh unit indicates the freshness of eggs stored in a refrigerator as Haugh unit value determines the changes of the interior qualities of eggs. Park et al. (2003) and Grashorn et al. (2016) also recorded a decrease in Haugh unit for eggs stored under high temperature. The present result is similar to that of Dudusola (2009) and Raji et al. (2009) who recorded higher Haugh unit values for eggs stored in a refrigerator compared to eggs stored under room temperature.

Internal qualities of table eggs as influenced by storage method

The effect of storage method on internal egg quality traits is presented in Table 1. There were significant ($P<0.05$) differences in internal egg qualities between eggs coated with vegetable oil and those that were not coated with vegetable oil. This difference could be attributed to the fact that oil has the ability to seal egg pores, preventing evaporation of moisture and carbon dioxide from the eggs during storage. Eggs coated with vegetable oil had heavier albumen weight and higher albumen height values than those that were not coated with vegetable oil. This might be due to the retention of moisture within the albumen of oiled eggs in the absence of osmotic pressure as observed by Orji et al. (1981). Eggs coated with vegetable oil were significantly higher ($P<0.05$) in yolk weight and yolk height than those eggs stored without vegetable oil and these values could be due to increase of fat within the yolk through absorption or reduction in moisture evaporation from the yolk. Raji et al. (2009) also observed higher yolk height in eggs of laying hens in a dry climate stored with vegetable oil.

The yolk colour was significantly ($P<0.05$) higher for oiled eggs (4.89) compared to eggs stored without oil application (4.55). The significantly higher yolk colour value observed for oiled eggs indicates that eggs stored after oil application maintained better yolk colour than eggs stored without oil application. Yolk colour has effect on the nutritional value of eggs. Eggs stored without oil application showed significantly higher ($P<0.05$) yolk diameter than those stored after vegetable oil application. The higher yolk diameter indicates the spread of yolk as a result of moisture loss from the yolk. This could be attributed to the evaporation of moisture from the eggs during storage as a result of high temperature. Oil helps to seal the various pores on the eggs preventing evaporation of moisture during storage.

Haugh unit showed a significant difference ($P<0.05$) with eggs coated with vegetable oil showing higher value (63.77) than eggs without vegetable oil (54.00). Güçlü et al. (2008) observed Haugh unit similar to the current results in Table 1, with eggs stored with fish oil showing higher ($P<0.05$) Haugh unit than other storage methods. Dudusola (2009) reported results that were similar to the current study. The current result also agrees with the finding of Grobas et al. (2001) and Eke et al. (2013) who reported significantly ($P<0.05$) higher Haugh unit for eggs stored with oil.

Internal qualities of table eggs as influenced by the two-way and three-way interactions of feather genotype, storage duration, storage temperature and storage method

The P values of the influence of the two and three-way interactions among genotype, storage duration, storage temperature and storage method on internal egg quality traits are presented in Table 2. No significant interaction of genotype by storage duration on internal egg qualities was observed except for yolk weight (0.008). This suggests that variation in yolk weight of chicken genotypes is dependent on the storage durations of the eggs. Significant interaction (0.034) of genotype and storage method on yolk colour of eggs was also observed.

Storage duration x storage temperature had significant effect on all the internal egg qualities studied except yolk colour. This is in agreement with other studies who have reported important storage duration x temperature influence on Haugh unit (Chung and Lee, 2014), albumen height (Samli et al., 2005), albumen weight and yolk weight (Jin et al., 2011). However, Jin et al. (2011) reported significant storage duration x storage time interaction on yolk colour which was contrary to the finding in this study. The variation between the two studies could be attributed to the differences in the breeds of chicken used.

Yolk height was significantly higher ($P<0.05$) for eggs coated with vegetable oil during storage; although the yolk height values slowly decreased with increase in storage time. Eggs stored without oil application rapidly deteriorated in yolk height as storage time increased. This result agrees with Tebesi et al. (2015) and Raji et al. (2009) who recorded higher yolk height values for eggs stored with oil application for shorter periods of time.

Storage temperature x storage method had significant ($P<0.05$) effect on albumen height, Haugh unit, yolk diameter, yolk height and yolk weight. There were significantly higher ($P<0.05$) albumen height values for oiled eggs stored in refrigerator as compared to eggs stored without oil application under room temperature. The eggs coated with oil stored at room temperature or refrigerator also recorded higher albumen height values than eggs stored at room temperature or refrigerator without oil application. This indicates that eggs coated with vegetable oil and stored in a refrigerator maintained better albumen quality possibly due to the prevention of moisture loss by evaporation thus retention of moisture in the albumen as the oil seals the egg pores. Dudusola (2009) reported that eggs coated with oil and refrigerated eggs did not lose much solvent as compared with those in polythene bag and uncoated. The significant variations in yolk weight and diameter due to storage temperature x storage method corroborate the findings of Dudusola (2009) and Orji et al. (1981) respectively who observed increased in yolk weight and diameter as a result of increase in storage temperature and storage time. The increased yolk weight during storage at room temperature could be due to movement of water from albumen to the yolk due to some high pressures. In addition, the significant variation in yolk height due to storage temperature x storage method agrees with Raji et al. (2009) who recorded higher yolk height values for oiled eggs stored at low temperature. Similarly, the important variation in Haugh unit due to storage temperature x storage method is an indication that changes in egg quality due to storage temperature is dependent on the presence or absence of oil application on the egg.

The three-way interactions of the factors studied were significant sources of variations for some of the internal egg parameters except yolk colour (Table 2). The explanation of some of these complex interactions could be quite complicated.

External qualities of table eggs as influenced by chicken genotype

The effect of genotype on external egg qualities are presented in Table 3. Chicken genotype had significant ($P<0.05$) effect on all the external qualities of eggs studied except egg weight. The differences in the external qualities could be attributed to the differences in the alleles controlling the genotypes. These results corroborate the findings of Egahi et al. (2013) who also found significant ($P<0.05$) effect of genotype on all the external egg qualities studied including egg weight. Rajkumar et al. (2009) however observed no significant differences in shell weight and egg weight for NaNa, Nana, and nana genotypes in India. Udoh et al. (2012) also reported no significant difference ($P>0.05$) among three local genotypes in terms of shell thickness in Nigeria.

Naked neck showing higher shell thickness value could be attributed to the result of their feed intake and feed conversion ratio. As the naked neck take in more feed, calcium from the feed is being converted into the egg shell thereby making their shell thicker than the other birds. However, the frizzle and the normal feathered birds showed no significant difference ($P>0.05$) in terms of shell thickness. The current result for naked neck was similar to that of Nwachukwu et al. (2006), who also recorded shell thickness between 0.30 mm to 0.34 mm in naked neck, frizzle and normal feathered birds. Yakubu et al. (2008) observed 0.38 mm of shell thickness in naked neck chickens from Nigeria which was higher than the values realized in the present study (0.31 mm). Egahi et al. (2013) also reported shell thickness of 0.33 mm in naked neck, 0.36 mm in frizzle, and 0.32 mm in normal feathered birds.

Table 2 - P-values of the 2 and 3-way interactions among the effects of feather genotype, storage duration, storage temperature and storage method on internal egg quality characteristics

Source of variation	Albumen height (mm)	Albumen weight (g)	Haugh unit (%)	Yolk colour	Yolk diameter, (cm)	Yolk height, (mm)	Yolk weight, (g)
Genotype*SD	0.994	0.384	0.505	0.986	0.099	0.112	0.008
Genotype*ST	0.937	0.408	0.945	0.936	0.958	0.715	0.856
Genotype*SM	0.467	0.467	0.580	0.034	0.397	0.583	0.568
SD*ST	0.052	<0.001	0.035	0.623	<0.001	<0.001	0.003
SD*SM	0.830	0.493	0.830	0.643	1.000	<0.001	0.126
ST*SM	0.017	0.696	<0.001	0.144	<0.001	<0.001	<0.004
Genotype*SD*SM	0.183	0.018	0.308	0.657	0.375	0.010	0.488
Genotype*SD*ST	0.077	0.140	0.111	0.423	0.506	0.020	0.173
Genotype*ST*SM	0.300	0.850	0.350	0.670	0.040	0.561	0.402
SD*ST*SM	0.043	0.824	0.008	0.655	<0.001	<0.001	0.008

¹SD: Storage duration; ST: Storage time; SM: Storage method

Table 3 - External qualities of table eggs as influenced by genotype, storage duration, storage temperature and storage method

Factors	Shell thickness (mm)	Shell weight (g)	Egg weight (g)	Egg length (cm)	Egg width (cm)
Genotype					
<i>Nanaff</i>	0.30 ^a	6.01 ^a	61.46	5.90 ^a	4.32 ^a
<i>nanaFf</i>	0.25 ^b	6.09 ^a	61.21	5.81 ^b	4.33 ^a
<i>nanaff</i>	0.22 ^b	5.95 ^b	60.95	5.80 ^b	4.28 ^b
SEM	0.004	0.042	0.333	0.016	0.010
p-value	<0.001	0.043	0.569	0.035	0.003
Storage duration					
0	0.27 ^a	6.16 ^a	63.21 ^a	5.95 ^a	4.37 ^a
7	0.27 ^a	6.02 ^b	61.45 ^b	5.88 ^a	4.31 ^a
14	0.27 ^a	5.96 ^b	61.34 ^b	5.87 ^a	4.31 ^a
21	0.26 ^a	5.96 ^b	60.04 ^c	5.83 ^b	4.28 ^b
28	0.23 ^b	5.99 ^b	59.98 ^c	5.85 ^b	4.28 ^b
SEM	0.005	0.049	0.395	0.019	0.012
p-value	<0.001	0.043	<0.001	<0.001	<0.001
Storage Temperature					
Refrigeration	0.26	6.02	61.45	5.86	4.30
Room Temperature	0.25	6.01	60.96	5.86	4.32
SEM	0.004	0.034	0.279	0.013	0.009
p-value	0.901	0.943	0.467	0.326	0.643
Storage method					
Vegetable oil	0.27 ^a	6.05	61.50 ^a	5.87	4.30
No oiling	0.24 ^b	5.98	60.40 ^b	5.86	4.31
SEM	0.003	0.035	0.300	0.013	0.009
p-value	<0.001	0.176	<0.001	0.709	0.752

^{abc}Means within the same sub-column with different subscripts are significant at $P<0.05$.

External qualities of table eggs as influenced by storage duration

Storage duration significantly ($P < 0.05$) influenced shell thickness and weight, egg weight, length and width (Table 3). Shell thickness was lower ($P < 0.05$) for eggs stored for 28 days compared to all the other storage durations. The significant variation in shell thickness due to storage duration is in agreement with the report of Grashorn et al. (2016) but contrary to the finding of Lee et al. (2016) who reported non-significant effect of storage duration on shell thickness. Shell weight and egg weight significantly ($P < 0.05$) decreased with increase in length of storage duration and this corroborates the findings of Samli et al. (2005), Jin et al. (2011), Akinola and Ibe (2014) and Lee et al. (2016). The loss in weight is attributed to water loss through evaporation from the pores in the egg shell and escape of carbon dioxide from the egg albumen (Samli et al., 2005). Dudusola (2009) also indicated that the loss of egg weight due to prolonged storage might be due to loss of carbon dioxide, ammonia, nitrogen, hydrogen sulphide gas and water from the eggs.

External qualities of table eggs as influenced by storage temperature and storage method during storage

The external egg qualities were not significantly ($P > 0.05$) different between the two storage temperatures. This finding does not agree with report of Raji et al. (2009) who observed higher egg weight for eggs stored in the refrigerator than those stored under room temperature. Oil application during egg storage had important ($P < 0.05$) effect on shell thickness and egg weight (Table 3). Eggs with oil application had thicker shells and heavier egg weight than those without oil application. The higher shell thickness of oil coated eggs is in agreement with Raji et al. (2009) who also reported higher shell thickness values for eggs coated with oil during storage. The high shell thickness of oiled eggs is due to the layer of oil applied on the shells. In addition, the heavier egg weights of oil applied eggs compared to non-oil applied eggs is probably due to the reduction in moisture loss through the pores on the shells.

External qualities of table eggs as influenced by the two-way and three-way interactions of genotype, storage duration, storage temperature and storage method

All the two-way and three-way interactions of genotype, storage duration, storage temperature and storage method on the external qualities of eggs were not significant ($P > 0.05$) except the interaction of storage duration x storage method on shell thickness, storage temperature x storage method on egg weight and genotype x storage duration x storage temperature on egg weight (Table 4). The significant ($P < 0.05$) interaction of storage duration x storage method observed in this study corroborates the findings of Tebesi et al. (2012) and Akinola and Ibe (2014) but contrary to the report of Raji et al. (2009).

Table 4 - P-values of the 2 and 3-way interactions among the effects of genotype, storage duration, storage temperature and storage method on external egg qualities

Factors	Shell thickness (mm)	Shell weight (g)	Egg weight (g)	Egg length (cm)	Egg width (cm)
Genotype*SD	0.989	0.702	0.477	0.156	0.550
Genotype*ST	0.870	0.876	0.939	0.577	0.045
Genotype*SM	0.910	0.736	0.986	0.069	0.580
SD*ST	0.513	0.841	0.401	0.256	0.034
SD*SM	0.007	0.386	0.237	0.632	0.590
ST*SM	0.100	0.279	0.031	0.765	0.502
Genotype*SD*SM	0.390	0.500	0.420	0.464	0.886
Genotype*SD*ST	0.756	0.763	0.009	0.164	0.142
Genotype*ST*SM	0.820	0.930	0.460	0.402	0.511
SD*ST*SM	0.453	0.142	0.166	0.685	0.642

¹SD: Storage duration; ST: Storage time; SM: Storage method

Effect of genotype on the proximate composition of egg albumen and egg yolk (as-fed basis)

Table 5 shows the effect of chicken genotype on proximate composition of chicken egg albumen. There were no significant ($P > 0.05$) difference among chicken genotypes for all the proximate compositions of egg albumen except ether extract (EE). The EE content of egg albumen from frizzle feathered hens were significantly lower (0.08%) than those of normal feathered and naked neck birds. There was no significant ($P > 0.05$) difference among chicken genotypes with respect to the proximate composition of egg yolk except for ash content. Eggs from frizzle feathered hens recorded higher levels of ash compared to those from the naked neck and normal feathered birds.

Effect of chicken genotype on the amino acid profile of table egg albumen and egg yolk

There were no significant differences ($P > 0.05$) among the chicken genotypes with respect to amino acid profile of the egg albumen (Table 6) and egg yolk (Table 7). The absence of significant differences among frizzle, naked neck and normal feathered birds with regard to amino acid profiles in the albumen and yolk might be due to the similarity of diet fed to the birds and the same environmental conditions under which they were raised.

Table 5 - Proximate composition on egg albumen and egg yolk as influenced by chicken genotype

Egg part	Moisture (%)	NFE (%)	Ash (%)	EE (%)	CF (%)	CP (%)
Genotype						
Egg albumen						
<i>Nanaff</i>	89.04	5.14	0.27	0.18 ^a	0.02	5.35
<i>nanaFf</i>	89.39	4.93	0.19	0.08 ^b	0.03	5.38
<i>nanaff</i>	88.66	5.49	0.17	0.20 ^a	0.02	5.45
SEM	0.27	0.23	0.04	0.01	0.04	0.13
P-value	0.18	0.24	0.12	<0.01	0.21	0.83
Egg yolk						
<i>Nanaff</i>	57.06	6.78	1.24 ^b	27.08	0.06	7.78
<i>nanaFf</i>	57.00	6.95	1.58 ^a	26.86	0.08	7.53
<i>nanaff</i>	57.17	6.74	1.16 ^b	27.05	0.07	7.81
SEM	0.09	0.18	0.09	0.18	0.03	0.05
P-value	0.40	0.70	0.01	0.67	0.86	0.20

^{ab}Means with different superscripts within the same column indicate a significant difference ($P < 0.05$). SEM: Standard Error of Means; P-value: Probability Value; NFE: Nitrogen Free Extract; EE: ether extract; CF: crude fibre; CP: crude protein.

Table 6 - Effect of feather genotype on amino acid profile as a percentage of egg albumen

Genotype	<i>Nanaff</i>	<i>nanaFf</i>	<i>nanaff</i>	SEM	P-value
Amino acid profile					
ALA (%)	0.59	0.59	0.59	0.01	0.95
ARG (%)	0.43	0.44	0.45	0.01	0.41
ASP (%)	1.02	1.03	1.03	0.01	0.78
CYS (%)	0.15	0.14	0.15	0.04	0.58
GLU (%)	1.29	1.31	1.32	0.02	0.50
GLY (%)	0.35	0.35	0.35	0.03	0.90
HIS (%)	0.24	0.24	0.25	0.03	0.76
ILE (%)	0.50	0.50	0.50	0.01	0.86
LEU (%)	0.82	0.84	0.84	0.01	0.92
LYS (%)	0.54	0.53	0.56	0.01	0.38
MET (%)	0.31	0.31	0.31	0.06	0.94
MET + CYT (%)	0.45	0.45	0.46	0.06	0.57
PHE (%)	0.57	0.59	0.59	0.08	0.82
PRO (%)	0.34	0.34	0.35	0.04	0.32
SER (%)	0.64	0.64	0.65	0.08	0.62
THR (%)	0.43	0.42	0.43	0.03	0.75
VAL (%)	0.67	0.67	0.67	0.01	0.90

¹SEM - Standard Error of Means; P-value: Probability Value; ALA: Alanine, ARG: Arginine; ASP: Aspartic acid; CYS: Cystine; GLU: Glutamic acid; GLY: Glycine; HIS: Histidine; ILE: Isoleucine; LEU: Leucine; LYS: Lysine; MET: Methionine; MET+CYS: Methionine+Cystine; PHE: Phenylalanine; PRO: Proline; SER: Serine; THR: Threonine; VAL: Valine.

Table 7 - Effect of genotype on amino acid profile of egg yolk

Genotype	<i>Nanaff</i>	<i>nanaFf</i>	<i>nanaff</i>	SEM	P-value
Amino acid profile					
ALA (%)	0.77	0.75	0.75	0.01	0.23
ARG (%)	0.56	0.56	0.57	0.01	0.72
ASP (%)	1.33	1.31	1.31	0.01	0.08
CYS (%)	0.19	0.18	0.18	0.01	0.66
GLU (%)	1.68	1.67	1.68	0.01	0.48
GLY (%)	0.46	0.46	0.45	0.01	0.45
HIS (%)	0.32	0.31	0.31	0.03	0.40
ILE (%)	0.65	0.64	0.63	0.01	0.63
LEU (%)	1.07	1.07	1.06	0.01	0.24
LYS (%)	0.70	0.68	0.71	0.01	0.39
MET (%)	0.40	0.39	0.39	0.01	0.48
MET + CYT (%)	0.59	0.57	0.58	0.01	0.31
PHE (%)	0.76	0.75	0.75	0.01	0.39
PRO (%)	0.45	0.43	0.45	0.03	0.19
SER (%)	0.84	0.82	0.83	0.01	0.39
THR (%)	0.55	0.55	0.55	0.03	0.08
VAL (%)	0.87	0.86	0.85	0.01	0.49

¹SEM: Standard Error of Means; P-value: Probability Value; ALA: Alanine, ARG: Arginine; ASP: Aspartic acid; CYS: Cystine; GLU: Glutamic acid; GLY: Glycine; HIS: Histidine; ILE: Isoleucine; LEU: Leucine; LYS: Lysine; MET: Methionine; MET+CYS: Methionine+Cystine; PHE: Phenylalanine; PRO: Proline; SER: Serine; THR: Threonine; VAL: Valine.

CONCLUSION

Naked neck and frizzle genes had positive influence on egg quality traits. Shorter storage duration had positive influence on egg qualities during storage. Eggs stored at low temperature showed positive results in terms of internal egg qualities. Eggs coated with vegetable oil also showed better egg quality during storage. Naked neck recorded heavier egg weight than frizzle and normal feathered in their interactions with storage duration and temperature. Refrigerator and vegetable oil showed better yolk quality in their interactions with storage duration. Information from this study could be used in the preservation of the internal and external qualities of table eggs from chicken.

DECLARATIONS

This study is part of the thesis of the first author. The complete thesis is available in the library of of the Kwame Nkrumah University of Science and Technology (KNUST). No part of this thesis is however published in any journal or conference proceedings.

Corresponding author

Correspondence and requests for materials should be addressed to ; E-mail: bernard.hagan@uenr.edu.gh; ORCID: <https://orcid.org/0000-0003-2902-5271>

Data availability

The datasets used and/or analysed during the current study available from the corresponding author on reasonable request.

Funding

None.

Ethics Approval

All experimental procedures which involved the use of birds were conducted in compliance with the ARRIVE guidelines.

Authors' contribution

Kanasuah DN performed the field work, analysed the data and drafted the manuscript, Adomako K designed the study, edited that manuscript and approved the final manuscript, Hagan BA designed the study, analysed the data and wrote the manuscript. Olympio OS edited the manuscript.

Acknowledgements

The authors are grateful to the management and staff of Akate Farms and Trading Company Limited (AFTC) and Department of Animal Science, KNUST for offering their facilities for this study and helping with data collection.

Competing Interests

None of the authors of this article has any competing interests in the publication of this article.

REFERENCES

- Abioja MO, Abiona JA, Akinjute OF and Ojoawo HT (2021). Effect of storage duration on egg quality, embryo mortality and hatchability in FUNAAB- α chickens. *Journal of Animal Physiology and Animal Nutrition*, 105(4):715-724. <https://doi.org/10.1111/jpn.13480>
- Abou-Emera OK, Ali U, Galal A, El-Safty S, Abdel-Hameid EF and Fathi MM (2017). Evaluation of genetic diversity of naked neck and frizzle genotypes based on microsatellite markers. *International Journal of Poultry Science*, 16:118-124. <http://dx.doi.org/10.3923/ijps.2017.118.124>
- Adomako K, Olympio OS, Hagan JK and Hamidu JA (2014). Effect of frizzle gene (F) on egg production and egg quality of laying hens kept in the tropical villages. *British Poultry Science*, 55(6):709-714. <http://dx.doi.org/10.1080/00071668.2014.963026>
- Akinola LA and Ibe GC (2014). Effect of colour, source and storage on quality of table eggs in Port Harcourt Metropolis, Rivers State, Nigeria. *Journal of Research in Agriculture and Animal Science*, 2(11): 1-6. <https://www.questjournals.org/jraas/papers/vol2-issue11/A2110106.pdf>
- Ali AA, Noor HSM, Chong PK, Babji AS and Lim SJ (2019). Comparison of amino acids profile and antioxidant activities between edible bird nest and chicken eggs. *Malaysian Applied Biology*, 48(2): 63-69. <https://jms.mabjournal.com/index.php/mab/article/view/1900>
- Asumah C, Adomako K, Olympio OS, Hagan BA and Yeboah ED (2022). Influence of thermoregulatory (Na & F) genes on performance and blood parameters of F2 and F3 generations of crosses of local and commercial chickens. *Tropical Animal Health and Production*, 54: 207. <https://doi.org/10.1007/s11250-022-03207-6>
- Attia YA, Al-Harhi MA, Korish MA and Shiboob MH (2020). Protein and amino acid content in four brands of commercial table eggs in retail markets in relation to human requirements. *Animals*, 10(3): 406. <https://doi.org/10.3390/ani10030406>
- Batkowska J, Melnyk O, Kutrzuba M and Drabik K (2023). Selected factors affecting the table eggs quality. *Modern Poultry*, 7-8: 26-29. <https://doi.org/10.31548/poultry2023.07-08.026>

- Chung SH and Lee K-W (2014). Effect of hen age, storage duration and temperature on egg quality in laying hens. *International Journal of Poultry Science*, 13(11): 634-636. <https://doi.org/10.3923/ijps.2014.634.636>
- Dudusola IO (2009). Effects of storage methods and length of storage on some quality parameters of Japanese quail eggs. *Tropicultura*, 27(1): 45-48. <http://www.tropicultura.org/text/v27n1/45.pdf>
- Edinformatics.com (2013): The anatomy of chicken egg. Available on https://www.edinformatics.com/science_projects/egg_proteins.htm
- Egahi JO, Dim NI and Momoh OM (2013). The effect of plumage modifier genes on egg quality indices of the Nigerian local chicken. *IOSR Journal of Agriculture and Veterinary Science*, 2(2): 4-6. <http://dx.doi.org/10.9790/2380-0220406>
- Eke MO, Olaitan NI and Ochefu JH (2013). Effect of storage conditions on the quality attributes of shell (table) eggs. *Nigerian Food Journal*, 31(2): 18-24. [https://doi.org/10.1016/S0189-7241\(15\)30072-2](https://doi.org/10.1016/S0189-7241(15)30072-2)
- El-Rahman A Abd and Makled MN (2006). Productive performance of naked neck laying hens (Sharkasi) fed different dietary protein levels. *Assiut Journal of Agricultural Science*, 31(1):85-103. <https://doi.org/10.21608/ajas.2006.273946>
- Farooq MK, Aneela FR, Durrani AK, Muqarrab NC and Khurshid A (2001a). Egg and shell weight, hatching and production performance of Japanese broiler Quails. *Sarhad Journal of Agriculture*, 17: 289-293.
- Farooq MK, Durrani AK, Aleem N, Chand N and Muqarrab NC (2001b). Egg traits and hatching performance of Desi, Fayoumi and Rhode Island Red Chicken. *Pakistan Journal of Biological Sciences*, 4(7): 909-911. <https://doi.org/10.3923/pjbs.2001.909.911>
- Fathi M, Abou-Emera O, Al-Homidan I, Galal A and Rayan G (2022). Effect of genotype and egg weight on hatchability properties and embryonic mortality pattern of native chicken populations. *Poultry Science*, 101:102129. <https://doi.org/10.1016/j.psj.2022.102129>
- Fathi MM, Galal A, El-Safty S and Mahrous M (2013). Naked neck and frizzle genes for improving chicken raised under high ambient temperature: I. Growth performance and egg production. *World's Poultry Science Journal*, 69(4):813-832. <https://doi.org/10.1017/S0043933913000834>
- Fayeye TR, Adeshiyani AB and Olugbami AA (2005). Egg traits, hatchability and early growth performance of the Fulani-ecotype. *Livestock Research for Rural Development*. 17(8): Article #94. <http://www.lrrd.org/lrrd17/8/faye17094.htm>
- Grashorn M (2016). Feed additives for influencing chicken meat and egg yolk color. *Handbook on Natural Pigment in Food and Beverages. Industrial Applications for Improving Food Color* Woodhead Publishing Series in Food Science, Technology and Nutrition. Pp. 283-302. <https://doi.org/10.1016/B978-0-08-100371-8.00014-2>
- Grashorn M, Juergens A and Bessei W (2016). Effects of storage condition on egg quality. *LOHMANN Information*, 50 (1):22-27. <https://lohmanna-breeders.com/media/2020/08/VOL49-GRASHORN-Storage.pdf>
- Grobas S, Mendez J, Lazaro R, de Blas C and Mateos GG (2001). Influence of source and percentage of fat added to diet on performance and fatty acids composition of egg yolks of two strains of laying hens. *Poultry Science*, 80: 1171-1179. <https://doi.org/10.1093/ps/80.8.1171>
- Guni FS, Mbagha SH, Katule AM and Goromela EH (2021). Effects of breed and management system on egg quality traits of two improved dual-purposes chicken breeds. *Livestock Research for Rural Development*, 33(12): Article #141. <http://www.lrrd.org/lrrd33/12/33141fadh.html>
- Güçlü BK, Uyanık F and İşcan KM (2008). Effects of dietary oil sources on egg quality, fatty acid composition of eggs and blood lipids in laying quail. *South African Journal of Animal Science*, 38(2): 91-100. <https://www.ajol.info/index.php/sajas/article/view/4114>
- Haugh RR (1937). The Haugh unit for measuring egg quality. *United States Egg Poultry Magazine*, 43: 552-555.
- Hamilton RMG (1982). Methods and factors that affect the measurement of egg shell quality. *Poultry Science*, 61: 2022-2039. <https://doi.org/10.3382/ps.0612022>
- Hegab IM and Hanafy AM (2019). Effect of egg weight on external and internal qualities, physiological and hatching success of Japanese quail eggs (*Coturnix coturnix japonica*). *Brazilian Journal of Poultry Science*, 21(3). <https://doi.org/10.1590/1806-9061-2018-0777>
- Inca JS, Martinez DA and Vilchez C (2020). Phenotypic correlation between external and internal egg quality characteristics in 85-week old laying hens. *International Journal of Poultry Science*, 19(8): 346-355. <https://doi.org/10.3923/ijps.2020.346.355>
- Iqbal J, Khan SH, Mukhtar N, Ahmed T and Pasha RA (2016). Effects of egg size (weight) and age on hatching performance and chick quality of broiler breeder. *Journal of Applied Animal Research*, 44:54-64. <https://doi.org/10.1080/09712119.2014.987294>
- Islam MA, Bulbuli SM, Seeland G and Islam AB (2011). Egg quality in different chicken genotypes in summer and winter. *Pakistan Journal of Biological Science*, 4: 1411-1414. <https://doi.org/10.3923/pjbs.2001.1411.1414>
- Jin YH, Lee KT, Lee WI and Han YK (2011). Effects of storage temperature and time on the quality of eggs from laying hens at peak production. *Asian-Australian Journal of Animal Science*, 24: 279-284. <http://dx.doi.org/10.5713/ajas.2011.10210>
- Khan MJA, Khan SH, Bukhsh A, Abbass MI and Javed M (2013). Effect of different storage period on egg weight, internal egg quality and hatchability characteristics of Fayumi eggs. *Italian Journal of Animal Science*, 12: 51. <https://doi.org/10.4081/ijas.2013.e51>
- Lee MH, Cho EJ, Choi ES and Sohn SH (2016). The effect of storage period and temperature on egg quality in commercial eggs. *Korean Journal of Poultry Science*, 43(1):31-38. <http://dx.doi.org/10.5536/KJPS.2016.43.1.31>
- Martínez Y, Soliz ND, Bejarano MA, Paz P and Valdivie M (2021). Effect of storage duration and temperature on daily changes in external and internal egg quality of eggs from Dekalb White® laying hens. *European Poultry Science*, 85: 329. <https://doi.org/10.1399/eps.2021.329>
- Melo EF, Araújo ICS, Triginelli MV, Castro FLS, Baião NC and Lara LJC (2020). Effect of egg storage duration and egg turning during storage on egg quality and hatching of broiler hatching eggs. *Animal*, 15(2):100111. <https://doi.org/10.1016/j.animal.2020.100111>
- Mountney GJ (1976). *Poultry production technology*. 2nd Edition. AVI publishing company, west port Connecticut. p. 291.
- Murshed M and Qaid, MM (2024). A comparative study of internal and external quality characteristics of table eggs and the effect of storage periods and layer strain on them under summer conditions. *Indian Journal of Animal Research*, 58(4): 681-687. <https://doi.org/10.18805/IJAR.BF-1721>
- Nasri H, van den Brand H, Najjar T and Bouzouaia M (2019). Egg storage and breeder age impact on egg quality and embryo development. *Journal of Animal Physiology and Animal Nutrition*, 104 (1):257-268. <https://doi.org/10.1111/jpn.13240>
- Nwachukwu EN, Ibe SN and Ejekwu K (2006). Short term egg production and egg quality characteristics of main and reciprocal crossbred normal local, naked neck and frizzle chicken X exotic broiler breeder stock in a humid tropical environment. *Journal of Animal and Veterinary Advances*, 5 (7): 547-551.

- Ogundero AE, Adenaike AS, Balogun AO and Ikeobi CON (2019). Principal component analysis and repeatability estimates of egg production traits in Nigerian indigenous chickens divergently selected for antibody response to sheep red blood cells (SRBC). *Bulletin of Animal Health and Production in Africa*, 67:355-363. <https://www.cabidigitallibrary.org/doi/full/10.5555/20203272029>
- Orji BI, Igboeli C and Okoye PT (1981). The effect of pre-incubation storage on embryonic growth rate, mortality, hatchability and total incubation period of fowl egg. *Nigerian Journal of Agricultural Science*, 31: 99-103.
- Park YS, Yoo IJ, Jeon KH, Kim HK, Chang EJ and Oh HI (2003). Effects of various eggshell treatments on the egg quality during storage. *Asian-Australian Journal of Animal Science*, 16(8):1224-1229. <http://dx.doi.org/10.5713/ajas.2003.1224>
- Qin K, Cong X, Wang H, Yan M, Xu X, Liu M, Song F, Wang D, Xu X, Zhao J, Cheng S, Liu Y and Zhu H (2024). Effects of supplementing selenium-enriched *Cardamine violifolia* to laying hens on egg quality and yolk antioxidant capacity during storage at 4 °C and 25 °C. *Foods*, 13(5): 802. <https://doi.org/10.3390/foods13050802>
- Raji AO, Aliyu J, Igwebuike JU and Chiroma S (2009). Effect of storage methods and time on egg quality traits of laying hens in hot dry climate. *ARPN Journal of Agriculture and Biological Science*, 4:1-7. https://www.arnjournals.com/jabs/research_papers/rp_2009/jabs_0709_136.pdf
- Rajkumar U, Sharma RP, Rajaravindra KS, Niranjan M, Reddy BLN, Bhattacharya TK and Chatterjee RN (2009). Effect of genotype and age on egg quality traits in naked neck chicken under tropical climate from India. *International Journal of Poultry Science*, 8(12):1151-1155. <https://doi.org/10.3923/ijps.2009.1151.1155>
- Roush WB (1981). TI 59 calculator program for Haugh Unit Calculation. *Poultry Science*, 60:1086-1088. <https://doi.org/10.3382/ps.0601086>
- Salahuddin M and Howlinder MAR (1991). Effect of breed and season on egg quality traits of fowl. *Indian Journal of Animal Science*, 61:859-863. <https://arccjournals.com/journal/indian-journal-of-animal-research/B-657>
- Samli HE, Agha A and Senkoğlu N (2005). Effects of storage time and temperature on egg quality in old laying hens. *Journal of Applied Poultry Research*, 14:548-553. <https://doi.org/10.1093/japr/14.3.548>
- Santos RC, Segura JC and Sarmiento LF (2019). Egg quality during storage of eggs from hens fed diets with crude palm oil. *Revista MVZ Córdoba*, 24(3):7297-7304. <https://doi.org/10.21897/rmvz.1244>
- Scott TA and Silversides FG (2000). The effect of storage and strain of hen on egg quality. *Poultry Science*, 79: 1725-1729. <https://doi.org/10.1093/ps/79.12.1725>
- Sonaiya EB and Swan SEJ (2004). Small-scale poultry production. Technical Guide Manual. FAO, Rome. <https://www.fao.org/4/y5169e/y5169e00.htm>
- Şahan U, Ipek A and Sozcu A (2014). Yolk sac fatty acid composition, yolk absorption, embryo development and chick quality during incubation in eggs from young and old broiler breeders. *Poultry Science*, 93:2069-2077. <https://doi.org/10.3382/ps.2013-03850>
- Tebesi T, Madibela OR and Moreki JC (2012). Effect of Storage Time on Internal and External Characteristics of Guinea Fowl (*Numida meleagris*) Eggs. *Journal of Animal Science Advances*, 2(6): 534-542.
- Tyasi TL, Lebogang L and Hloko VR (2022). Comparative study of egg quality traits between Potchefstroom Koekoek and Hy-line silver brown layers. *Bulgarian Journal of Agricultural Science*, 28(1): 145-150. <https://www.agrojournal.org/28/01-20.pdf>
- Udoh UH, Okon B and Udoh AP (2012). Egg quality characteristics, phenotypic correlations and prediction of egg weight in three (Naked Neck, Frizzled Feather and Normal Feathered) Nigerian local chickens. *International Journal of Poultry Science*, 11(11):696-699. <https://doi.org/10.3923/ijps.2012.696.699>
- United States Department of Agriculture - USDA (2000). Egg-grading manual. Washington: Department of Agriculture. 56p. (Agricultural Marketing Service, 75).
- United States Department of Agriculture, USDA (2008). Food safety and inspection service, risk assessments of *Salmonella enteritis* in shell eggs and *Salmonella spp.* in egg products. Available from- http://www.fsis.usda.gov/PDF/SE_Risk. Accessed on March 19 2016.
- Yahaya HK, Olutunmugun AK, Mohammad YB, Shettima MM and Kabir M (2021). Evaluation of egg quality characteristics of two strains of local turkey (*Meleagris gallopavo*) in Zaria, Kaduna State. *Nigerian Journal of Animal Production*, 48(1): 1-11. <https://doi.org/10.51791/njap.v48i1.2891>
- Yakubu A, Oga OM and Barde RE (2008). Productivity and egg quality characteristics of free range Naked Neck and Normal feathered Nigerian indigenous chickens. *International Journal of Poultry Science*, 7(6):579-588. <https://doi.org/10.3923/ijps.2008.579.585>
- Zhang L-C, Ning Z-H, Xu G-Y, Hou Z-C and Yang N (2005). Heritabilities and genetic and phenotypic correlations of egg quality traits in brown-egg dwarf layers. *Poultry Science*, 84:1209-1213. <https://doi.org/10.1093/ps/84.8.1209>

Publisher's note: Sciencline Publication Ltd. remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access: This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <https://creativecommons.org/licenses/by/4.0/>.

PRIMAL CUTS OF CARCASS AND MEAT CHARACTERISTICS OF KACANG GOAT FED TOTAL MIXED RATION CONTAINING DIFFERENT SOURCES OF RUMINALLY UNDEGRADED PROTEIN

Retno ADIWINARTI¹✉ , KUSTANTINAH² , RUSMAN³ , Edy RIAN TO¹ , Agung PURNOMOADI¹ , Mukh ARIFIN¹ , SUTARYO¹ , and Vita RESTITRISNANI¹ 

¹Department of Animal Sciences, Faculty of Animal and Agricultural Sciences, Universitas Diponegoro, Indonesia

²Department of Animal Nutrition and Feeds Science, Faculty of Animal Science, Gadjah Mada University, Indonesia

³Department of Animal Product Technology, Faculty of Animal Science, Gadjah Mada University, Indonesia

✉ Email: retnoadiwinarti@lecturer.undip.ac.id

➤ Supporting Information

ABSTRACT: This study was designed to evaluate the effect of feed quality improvement using gliricidia and different sources of protein in total mixed ration (TMR) on the primal cuts, loin eye area, and fatty acids profile of goat meat. This study used twenty yearling Kacang goats weighing 17.42 ± 1.63 kg. The goats were randomly allocated into 4 different treatments in a completely randomized design. The treatments involved the use of natural grass from rangeland (NGFR; control) as well as improving the quality of feed through TMR containing various ruminally undegraded protein sources, i.e. TMR contains fish meal (TMR-FM), TMR contains soybean meal (TMR-SBM) and TMR contains formaldehyde treated soybean meal (TMR-TSBM). The parameters observed were primal cuts yield, loin eye area, meat, fat, bone of primal cuts, and fatty acids profile. Data were analyzed using a one-way analysis of variance. The results showed that the goats fed TMR-FM and TMR-TSBM produced significantly higher meat percentage than control goats. The meat yield of TMR-SBM and TMR-TSBM goats were significantly higher than those of control goats. Goats fed TMR-SBM produced the highest primal cuts yield and shoulder weight, while the weight of rib, loin, and leg of TMR-SBM goats were similar to those of TMR-TSBM goats. Loin eye area was similar between the treatments. Saturated fatty acids content in TMR groups was similar to those in control. It can be concluded that improved feed quality using TMR-SBM produced significantly higher primal cuts weight, while TMR-TSBM had better meat-to-bone ratio than control. TMR-TSBM goats produced significantly leaner meat than TMR-SBM goats. Fatty acid profiles were similar between treatments.

Keywords: Fatty acids, Fish meal, Goat meat, Meat quality, Total mixed ration.

INTRODUCTION

Goats are widely raised traditionally by farmers in Indonesia. They are usually kept in the yard and fed natural forages from the rangeland, so that they have low growth rate. Nevertheless, they produce high quality of meat (Slimani et al., 2022) due to their high content of unsaturated fatty acids (Makmur et al., 2020). Goat meat is considered a healthier meat compared to other red meat products for its leanness (Slimani et al., 2022; Gawat et al., 2023), and has a higher ratio of polyunsaturated fatty acids to saturated fatty acid (PUFA/SFA) when fed with forage-based diet (Lee et al., 2023). Lee et al. (2023) reported that dietary nutrient contents influence fatty acids composition of Korean native black goat meat, and forage-based diet increased PUFA to SFA ratio. To increase the goat production, Slimani et al. (2022) added concentrate and hay for pasture grazing goats. Low production of goats kept extensively was caused by the scarcity of good quality forages, limited of herbaceous plants and the low quality of shrubs (Slimani et al., 2022). Goats are selective eaters, preferring wild grass over soft grass (Lee et al., 2019) and showing a preference for woody plants (Chebli et al., 2022). Fish meal (FM) and soybean meal (SBM) are common protein sources in animal diet. Fish meal tends to be less degraded in the rumen than SBM (Falihatizow et al., 2015), but it is more expensive than SBM. Additionally, goats do not favor diets containing FM (Adiwinarti et al., 2016). Therefore, a better option is to use total mixed ration (TMR) for blending feed ingredients, ensuring that animals cannot selectively choose the feed based on their preference (Fluharty et al., 2017; Santana et al., 2017). Total mixed ration can be formulated based on the animals need. Santana et al. (2017) stated that TMR could maintain nutrient content in the ration. Protein in SBM is easily degraded in the rumen (Wang et al., 2021; Phesatcha et al., 2022) and provide about 65% of rumen-degradable protein (Phesatcha et al., 2022). In order to improve carcass weight, primal cuts and meat yield, protein in feed must be efficiently utilized by reducing rumen protein digestibility, so that post-rumen utilization of protein is more optimal. There have been many studies have been done to reduce the degradability of SBM in the rumen. Rooke et al. (1983) reported that degradability of formaldehyde-treated SBM was lower than that of untreated SBM. Widyobroto et al. (2010) stated that formaldehyde protected SBM increased rumen undegraded protein about 50-80%, but the formaldehyde protected SBM did not decrease the degradability in the small intestine. This study was set up to improve carcass weight, primal cuts yield, meat yield, and goat meat quality by enhancing the feed quality using TMR. Natural grass from rangeland (forage-based diet) was used as control to represent the common practice of goat rearing in the rural area, while improved feed quality involved TMR consisting of Napier grass, gliricidia, and concentrate with different protein sources (fish meal, SBM, and formaldehyde-treated SBM). The objectives of this study were to evaluate the carcass weight, primal cuts yield, meat yield, and meat

RESEARCH ARTICLE
 PII: S222877012500005-15
 Received: May 13, 2024
 Revised: January 16, 2025
 Accepted: January 18, 2025

quality of goats fed with forage-based diet compared to those fed with higher-quality feed (TMR containing different protein sources: fish meal, SBM, and formaldehyde-treated SBM).

MATERIALS AND METHODS

Animals, feeds, and experimental design

Twenty yearling Kacang goats with the initial body weight of 17.42 ± 1.63 kg were reared 95 days before being slaughtered. The goats were randomly allocated into 4 different treatments using completely randomized design. The treatments were: 100% natural grass from rangeland (NGFR) as the control and 3 improved feed quality used total mixed ration consisted of 30% Napier grass, 30% gliricidia leaves, and 40% concentrate with different sources of protein, i.e. TMR containing fish meal (TMR-FM), TRM containing soybean (TMR-SBM) and TMR containing formaldehyde treated soybean meal (TMR-TSBM) (Table 1). A 1% of formaldehyde treated soybean meal was made by using formalin and it was formulated to contain 1% of formaldehyde that was used to treat SBM in dry matter bases calculation.

Animals care and management

In order to protect from internal and external parasites, all goats were treated orally with 1.5 mL/head of Valbendazol and injected by 0.5 mL/head of Intermetctin. Goats were housed individually, provided feed and water *ad libitum* given at 8:00 AM, 12:00 PM, and 4:00 PM.

Table 1 - The composition of feed ingredients, chemical composition and Fatty acids composition (%) of the rations

Parameters	NGFR	TMR-FM	TMR-SBM	TMR-TSBM
Feed ingredients composition				
Natural grass from rangeland	100	-	-	-
Napier grass	-	30	30	30
Gliricidia leaves	-	30	30	30
Concentrate feed consisted of:	-	40	40	40
Cassava waste product		50.2	48.0	48.0
Wheat bran		34.4	34.5	34.5
Fish meal		15.4	0	0
Soybean meal		0	17.5	0
Formaldehyde treated soybean meal		0	0	17.5
Chemical composition				
Dry matter	18.6	91.3	91.5	91.4
Crude protein	10.9	15.3	15.6	14.3
Total digestible nutrients	63.2	56.2	58.0	60.1
Fatty acids composition				
Myristic acid (C14:0)	14.76	1.70	0.00	3.95
Palmitic acid (C16:0)	53.62	29.02	28.56	28.93
Stearic acid (C18:0)	12.54	11.97	9.34	10.62
Palmitoleic acid (C16:1)	0.00	0.64	0.00	0.57
Oleic acid (C18:1)	3.80	18.76	19.85	14.27
Linoleic acid (C18:2)	15.27	24.37	29.50	28.48
Linolenic acid (C18:3)	0.00	13.54	12.75	13.19
SFA (saturated fatty acids)	80.93	42.69	37.91	43.50
MUFA (mono unsaturated fatty acids)	3.80	19.40	19.85	14.84
PUFA (poly unsaturated fatty acids)	15.27	37.91	42.25	41.66
UFA (unsaturated fatty acids)	3.71	36.48	32.63	29.78

NGFR: natural grass from rangeland, TMR-FM: total mixed ration containing fish meal, TMR-SBM: total mixed ration containing soybean meal, TMR-TSBM: total mixed ration containing formaldehyde treated soybean meal

During the experimental period, two goats from the TMR-FM and TMR-TSBM groups became ill and died, resulting in only 18 goats being available for data collection. After a treatment periods of 95 days, the goats were deprived of feed and given only clean water for 12 hours. Subsequently, the goats were weighed and slaughtered. Goats were slaughtered to obtain carcass, which was then cooled at a temperature of 4 °C for 12 hours before further carcass observation were conducted. Carcass were cut and the primal cuts (shoulder, rib, loin, leg) were dissected for meat, fat, and bone separation to observe weight and percentage of carcass composition (meat, fat, and bone). Loin eye areas were measured following the method of [Rezende et al. \(2020\)](#).

Sampling and sample analysis

Samples from *Biceps femoris* muscle were frozen before fatty acids analysis. Fatty acids composition was analyzed using Gas Chromatography-mass Spectrometry ([Stashenko and Martinez, 2014](#)).

Statistical analysis

Data were analyzed by a one-way analysis of variance using significance level based on $p < 0.05$ ([Steel and Torrie, 1980](#)). If there was a significantly different between the treatments, Duncan's Multiple Range test was used for further analysis ([Steel and Torrie, 1980](#)).

Table 2 – Performance, carcass, and primal cuts characteristics of goats fed natural grass and total mixed ration

Parameters	NGFR	TMR-FM	TMR-SBM	TMR-TSBM	P-value
Performance					
Dry matter intake (g)	485.27 ^b ±58.31	620.71 ^{ab} ±47.54	740.24 ^a ±132.16	648.00 ^a ±106.29	0.007
Protein intake (g)	53.00 ^c ±6.37	94.72 ^b ±7.25	115.43 ^a ±20.61	92.40 ^b ±15.16	0.0001
Total digestible nutrients intake (g)	309.28 ^b ±42.89	349.55 ^{ab} ±40.33	438.19 ^a ±101.57	388.55 ^{ab} ±58.86	0.053
Average daily gain (g)	28.92 ^c ±7.05	57.56 ^b ±21.42	78.54 ^a ±10.23	56.19 ^b ±4.94	0.0001
feed conversion ratio	17.60 ^a ±4.67	12.52 ^{ab} ±6.35	9.45 ^b ±1.30	11.48 ^b ±1.11	0.035
Slaughter weight (kg)	20.04 ^c ±1.66	20.59 ^c ±1.72	25.09 ^a ±1.33	22.97 ^{bc} ±0.55	0.0001
Carcass weight (g)	7,174.80 ^c ±1,076.00	7,979.75 ^{bc} ±1,288.16	10,042.20 ^a ±1,021.03	8,882.75 ^{ab} ±703.26	0.005
Primal cuts (% carcass)	74.25±2.02	73.68±1.32	75.48±1.36	72.47±3.13	0.218
Primal cuts yield (g)	5,322.29^c±770.19	5,868.42^{bc}±863.04	7,573.89^a±699.17	6,434.48^b±541.31	0.002
Shoulder (g)	1,983.02 ^b ±323.04	2,271.63 ^b ±547.55	2,842.38 ^a ±252.23	2,314.69 ^b ±280.65	0.015
Rib (g)	553.98 ^b ±125.82	595.60 ^b ±57.94	850.42 ^a ±214.31	723.95 ^{ab} ±41.91	0.019
Loin (g)	636.04 ^b ±168.13	670.56 ^b ±89.49	925.54 ^a ±85.19	826.59 ^{ab} ±169.77	0.016
Leg (g)	2,149.26 ^b ±289.18	2,330.63 ^b ±227.17	2,955.55 ^a ±491.55	2,569.25 ^{ab} ±261.80	0.014
Primal cuts composition					
Meat (g)	3,758.10 ^c ±560.60	4,371.55 ^{bc} ±789.55	5,458.95 ^a ±445.56	4,815.94 ^{ab} ±384.64	0.002
Meat (%)	70.58 ^b ±1.24	74.21 ^a ±2.53	72.17 ^{ab} ±3.16	74.87 ^a ±0.51	0.039
Fat (g)	381.17 ^b ±133.70	322.29 ^b ±56.58	661.89 ^a ±186.73	420.71 ^b ±182.34	0.019
Fat (%)	7.06±2.00	5.59±1.29	8.72±2.33	6.47±2.38	0.182
Bone (g)	1,183.02±163.40	1,174.58±89.05	1,453.04±243.81	1,197.83±120.25	0.069
Bone (%)	22.36 ^a ±2.40	20.19 ^{ab} ±1.79	19.11 ^b ±1.70	18.67 ^b ±1.88	0.053
Edible portion (%)	77.64 ^b ±2.40	79.81 ^{ab} ±1.79	80.89 ^a ±1.70	81.33 ^a ±1.88	0.053
Meat-bone ratio	3.19 ^b ±0.36	3.70 ^{ab} ±0.44	3.81 ^{ab} ±0.46	4.04 ^a ±0.39	0.043
Meat+fat-bone ratio	3.51±0.47	3.98±0.44	4.27±0.46	4.40±0.55	0.058
Loin eye area (cm ²)	4.75±1.32	5.47±1.82	7.72±2.96	7.22±1.80	0.140

NGFR: natural grass from rangeland, TMR-FM: total mixed ration containing fish meal, TMR-SBM: total mixed ration containing soybean meal, TMR-TSBM: total mixed ration containing formaldehyde treated soybean meal

Table 3 - Fatty acids concentration of goat meat

Parameters	NGFR	TMR-FM	TMR-SBM	TMR-TSBM	P-value	Average
SFA (%)	52.77±1.63	43.45±8.92	42.25±6.72	42.93±10.27	0.127	45.59
Myristic acid (%)	1.24±0.89	1.92±1.69	1.38±1.04	1.29±0.64	0.798	1.44
Palmitic acid (%)	21.43±1.72	19.12±5.45	17.59±3.21	21.87±3.36	0.259	19.95
Stearic acid (%)	30.10±1.29	22.40±2.25	23.28±5.19	19.77±13.34	0.176	24.20
MUFA (%)	46.07±2.09	49.40±11.89	57.06±6.76	56.34±10.80	0.169	52.15
Palmitoleic acid (%)	1.58±0.97	1.66±0.76	1.61±0.84	1.82±1.10	0.981	1.66
Oleic acid (%)	44.50±2.32	47.74±11.14	55.45±7.47	54.52±9.83	0.153	50.49
PUFA (%)	1.16±0.60	0.77±0.91	0.69±0.34	0.73±0.55	0.602	0.85
Linolenic acid (%)	0.16±0.35	0.44±0.75	0.19±0.21	0	0.521	0.17
Arachidonic acid (%)	1.00±0.52	0.33±0.29	0.51±0.16	0.73±0.55	0.167	1.44
UFA (MUFA+PUFA) (%)	47.23±1.63	56.55±8.92	57.75±6.72	57.07±10.27	0.127	54.41
MUFA/SFA	0.88±0.07	1.19±0.45	1.41±0.45	1.43±0.68	0.235	1.22
PUFA/SFA	0.02±0.01	0.02±0.03	0.02±0.01	0.02±0.01	0.856	0.02

NGFR: natural grass from rangeland; TMR-FM: total mixed ration containing fish meal; TMR-SBM: total mixed ration containing soybean meal; TMR-TSBM: total mixed ration containing formaldehyde treated soybean meal; SFA: saturated fatty acids; MUFA: monounsaturated fatty acids; PUFA: polyunsaturated fatty acids; UFA: unsaturated fatty acids.

RESULTS

After being raised for 95 days, the performance of the goats including feed intake, growth rate, and feed conversion ratio are presented in the Table 2. Those parameters were significantly different between the treatments ($P < 0.05$). The slaughter weights of the goats were significantly different between the treatments ($P < 0.01$). Carcass weights and primal cuts yield also showed significant differences between the treatments ($P < 0.01$) (Table 2). In fact, the primal cut percentages were similar across all treatments. Additionally, there were significant differences in the meat-to-bone ratio of primal cuts between the treatments ($P < 0.05$). The fatty acids composition (%) of the rations is presented in the Table 1. The main fatty acids identified in goat meat in this study included myristic acid, palmitic acid, stearic acid, palmitoleic acid, oleic acid, linolenic acid, and arachidonic acid. Myristic acid, palmitic acid, stearic acid are grouped into saturated fatty acids (SFA), while palmitoleic acid and oleic acid belong to the category of mono-unsaturated fatty acids (MUFA), and linolenic acid and arachidonic acid are poly-unsaturated fatty acids (PUFA). There were no significant differences observed in the fatty acids content between the treatments (Table 3).

DISCUSSION

Productivity of goats fed natural grass and total mixed ration

Carcass weight of goats fed TMR containing SBM and TSBM were higher than those of control goats (NGFR) (Table 2). The higher carcass weight because of their average daily gain were higher than those of control goats. In addition, the faster growth of TMR goats was caused by the more dry matter and protein intake (Table 2).

Primal cuts were used as a parameter of goat production in this study because they represent about $74.07 \pm 2.16\%$ of the total carcass. Primal cuts consisted of shoulder, rib, loin, and leg (Rezende et al., 2020). Total primal cuts yield (g) and shoulder (g) of TMR-SBM were higher than other treatments, while the weight of rib, loin, and leg (g) of TMR-SBM were similar to those of TMR-TSBM. This indicated that improved feed quality using TMR-SBM and TMR-TSBM produced higher product than control.

Goats in TMR-SBM and TMR-TSBM produced more primal cuts meat compared to control. In fact, the meat percentage of TMR-TSBM and TMR-FM was higher than those of control (Table 2). Goats from TMR-SBM and TMR-TSBM had bone percentage that was lower than control. In addition, the meat-to-bone ratio and the percentage of edible portion in TMR-TSBM were higher compared to the control group. However, the fat content in TMR-SBM was the highest ($p < 0.05$) compared to the other treatments. These findings suggest that goats fed TMR produced better product than those in the control group, particularly when TMR containing SBM and TSBM. The meat from TMR-TSBM goats also appeared leaner compared to the meat from the TMR-SBM goats. Bambou et al. (2021) stated that diet is one of the factors influencing meat production. Goats fed with TMR-TSBM might increase the bypass protein level, thereby enhancing meat production. Rooke et al. (1983) reported that the degradation rate of formaldehyde-treated SBM decreased from 0.90 (untreated SBM) to 0.40 (formaldehyde-treated SBM). Widjibroto et al. (2010) stated that formaldehyde protected SBM can increase rumen undegraded protein by about 50-80%. However, it is essential to note that formaldehyde treatment did not reduce the protein degradation ability in the small intestine. Therefore, undegraded rumen protein can be efficiently utilized to enhance meat production. Additionally, tannin in gliricidia might reduce the digestibility of dry matter, organic matter, and crude protein as reported by Aguerre et al. (2016). Adiwintarti et al. (2019) reported that retained protein to meat conversion ratio of goat fed ration containing 50% untreated SBM+50% formaldehyde treated SBM is better than those of goats fed a diet containing untreated SBM or those of goats fed a diet containing treated SBM.

The meat content (%) in this study was relatively similar to the findings of Bambou et al. (2021), who reported values ranging from approximately 69.1% to 75.1%. The meat-to-bone ratio of primal cuts in TMR goats was higher compared to the meat-to-bone ratio reported by Cruz et al. (2023), which ranged between 3.08 to 3.28. However, Bambou et al. (2021) reported that the muscle-to-bone ratio in the left shoulder is between 2.8 and 4.2.

Loin eye area of goats was relatively similar between the treatments. Loin eye area of NGFR (4.75 cm^2) and TMR-FM (5.47 cm^2) were smaller than those reported by Cruz et al. (2023), but those of TMR-SBM (7.72 cm^2) and TMR-TSBM (7.22 cm^2) were relatively similar. Some researchers reported that loin eye areas are $7.35\text{-}8.00 \text{ cm}^2$ in goat having 21.8 to 23.4 kg of body weight (Cruz et al., 2023), $8.51\text{-}9.36 \text{ cm}^2$ in growing period and $12.81\text{-}13.01 \text{ cm}^2$ in fattening period of Nubian goats (Chen et al., 2022), $13.91\text{-}15.12 \text{ cm}^2$ in goat with body weight of 44.9 to 48.4 kg (Kafle et al., 2021).

Fatty acids of goats fed natural grass and total mixed ration

The main SFA of goat meat in this study consisted of stearic and palmitic acid, while the main UFA was oleic acid. Previous studies also reported that the dominant fatty acids in goat meat are palmitic acids, stearic acids, and oleic acids (Kafle et al., 2021; Akbas et al., 2022), along with linoleic acid and arachidonic acids (Kim et al., 2019). According to Kafle et al. (2021), palmitic acids, stearic acids, and oleic acids contribute approximately 80-85% of the total fatty acids.

In this study, goat meat had a higher content of SFA and MUFA compared to PUFA, aligning with the findings reported by Dinh et al. (2021). The high presence of SFA in ruminants was attributed to the biohydrogenation process in their

rumens. [Slimeni et al. \(2022\)](#) also reported that extensively raised goats had a higher proportion of SFA, while semi-intensively raised goats produced more MUFA in goat meat.

The SFA content in goat meat from the goats in control group was relatively similar to that in the TMR group (Table 3), although the SFA content in natural grass from rangeland (control) was higher than those in TMR diets (Table 1). The average SFA content in TMR goat meat was 42.83% that was lower than the findings in [Bambou et al. \(2021\)](#) or [Slimeni et al. \(2022\)](#), but comparable to [Lee et al. \(2023\)](#) and higher than in [Akbas et al. \(2022\)](#). [Slimeni et al. \(2022\)](#) reported that extensively raised goat had 50.3% SFA, while semi intensively raised goat had 44.6% SFA. [Bambou et al. \(2021\)](#) reported SFA content in Creole goat meat ranging from about 42.8% to 52.6%. [Lee et al. \(2023\)](#) reported an SFA concentration of 42.48%, while [Akbas et al. \(2022\)](#) reported around 38.9%. [Akbas et al. \(2022\)](#) stated that low concentration of lauric acids, miristic acids, palmitic acids, and stearic acids indicated better meat quality. The concentration of UFA in the TMR group was relatively similar to those reported by [García et al. \(2019\)](#) and [Lee et al. \(2023\)](#). The MUFA concentration in this study was higher compared to the findings reported by [García et al. \(2019\)](#): 42.9%, [Akbas et al. \(2022\)](#): 43.53%, and [Lee et al. \(2023\)](#): 36.27%. Meanwhile, the concentration of PUFA was lower compared to the results of [García et al. \(2019\)](#), [Akbas et al. \(2022\)](#), and [Lee et al. \(2023\)](#). [Akbas et al. \(2022\)](#) reported a PUFA content of 14.85%, [Lee et al. \(2023\)](#) reported 21.25%, [García et al. \(2019\)](#) reported 6.57%.

In this study PUFA/SFA ratio of forage diet (NGFR) was similar to other treatments. However, [Lee et al. \(2023\)](#) mentioned that different diets can influence the fatty acid composition of native Korean black goats meat, and a forage-based diet can enhance the PUFA/SFA ratio. The PUFA/SFA ratio in this study was lower compared to other researchers (0.08 reported by [Guzmàn et al. \(2020\)](#) and between 0.1 to 0.15 reported by [García et al. \(2019\)](#)).

The ratio of MUFA to SFA between treatments did not show significant differences. The MUFA/SFA ratio in goats fed with TMR (1.19 to 1.43) was higher compared to the findings of [García et al. \(2019\)](#), [Guzmàn et al. \(2020\)](#) and [Akbas et al. \(2022\)](#) who reported MUFA/SFA ratios ranging from 0.32 to 0.83 ([Guzmàn et al., 2020](#)), 0.81 to 0.92 ([García et al., 2019](#)), and 1.07 to 1.15 ([Akbas et al., 2022](#)).

CONCLUSION

In conclusion, improving feed quality by using TMR-SBM (total mixed ration containing soybean meal) resulted in higher primal cuts weight, while TMR-TSBM (total mixed ration containing formaldehyde treated soybean meal) displayed a better meat-to-bone ratio compared to the control. Goats fed with TMR-TSBM produced leaner meat compared to those fed with TMR-SBM. The fatty acid profiles were similar between the treatments.

DECLARATIONS

Corresponding author

Correspondence and requests for materials should be addressed to Retno ADIWINARTI; E-mails: retno_adi@yahoo.co.id; retnoadiwinarti@lecturer.undip.ac.id;  ORCID: 0000-0003-0638-4220

Ethical considerations

This experiment procedures related to the use of animals have been approved by the committee of animal ethics in the Animal and Agricultural Science, Universitas Diponegoro (59-04/A-08/KEP-FPP) and the authors have complied with the ARRIVE guidelines.

Acknowledgements

The authors are gratefully thanks to Prof. Ir. I Gede S. Budisatria, M.Sc., Ph.D. (RIP) for supervising the research, "Kacang goat" team and Meat Science Laboratory assistants for helping the data collection.

Authors' contribution

R.Adiwinarti designed the research, supervised the fieldwork, analyzed the data, prepared and wrote the manuscript.

Kustantinah and Rusman participated in designing the research, reviewed and edited the manuscript.

E.Rianto and A.Purnomoadi participated in supervising the fieldwork, reviewed and edited the manuscript.

M.Arifin and Sutaryo contributed to review and edit the manuscript.

V.Restitrisnani supervised the fieldwork and lab work.

All authors have read and approved the final manuscript.

Competing interests

There are no competing interests regarding the publication of this article.

REFERENCES

- Adiwinarti R, Kustantinah, Budisatria IGS, Rusman and Indarto E (2016). Improving the performance of local Kacang goats using ruminally undegradable protein feeds. *Asian Journal of Animal Science*, 10(4): 262-267. <https://doi.org/10.3923/ajas.2016.262.267>
- Adiwinarti R, Budisatria IGS, Kustantinah K, Rusman R and Indarto E (2019). Effects of rations containing formaldehyde-protected soybean meal on meat production in Kacang goats. *Veterinary World*, 12(6):890-895. <https://doi.org/10.14202/vetworld.2019.890-895>
- Aguerre MJ, Capozzolo MC, Lencioni P, Cabral C and Wattiaux MA (2016). Effect of quebracho-chestnut tannin extracts at 2 dietary crude protein levels on performance, rumen fermentation, and nitrogen partitioning in dairy cows. *Journal of Dairy Science*, 99:4476-4486. <http://dx.doi.org/10.3168/jds.2015-10745>
- Akbas AA, Kuleasan Ş, Üstuner H, Elmaz O, Sari M and Saatci M (2022). Some meat quality traits and fatty acid composition of Saanen, Turkish Hair × Saanen (F1) and Honamlı × Saanen (F1) crossbreed kids in tropical region of Turkey. *Research Square*. <https://doi.org/10.21203/rs.3.rs-1834997/v1>
- Bambou J-C, Cériac S, Liméa L, Arquet R, Bocage B and Alexandre G (2021). Impact of diet supplementation and age at slaughter on carcass characteristics of creole goats. *Frontiers in Veterinary Science*, 8, 671948. <https://doi.org/10.3389/fvets.2021.671948>
- Chebli Y, El Otmani S, Hornick J-L, Keli A, Bindelle J, Cabaraux J-F and Chentouf M (2022). Forage availability and quality, and feeding behaviour of indigenous goats grazing in a Mediterranean Silvopastoral system. *Ruminants*. 2:74–89. <https://doi.org/10.3390/ruminants2010004>
- Chen Y-A, Chen J-Y, Chen W-Q, Wang W-Y and Wu H-H (2022). Effects of castration age on the growth performance of Nubian crossbred male goats. *Animals*, 12:3516. <https://doi.org/10.3390/ani12243516>
- Cruz GFL, Santos EM, Araújo GGL, Azevedo PS, de Albuquerque ÍRR, Panosso NM, Perazzo AF, Zanine AM, Ferreira DJ, Lima AGVO and Oliveira JS (2023). Carcass traits and meat quality of goats fed with cactus pear (*Opuntia ficus-indica* Mill) silage subjected to an intermittent water supply. *Scientific Reports*, 13: 855. <https://doi.org/10.1038/s41598-022-25923-7>
- Dinh TTN, To KV and Schilling MW (2021). Fatty acid composition of meat animals as flavor precursors. *Meat and Muscle Biology*, 5(1):34. <https://doi.org/10.22175/mmb.12251>
- Falahatizow J, Danesh Mesgaran M, Vakili AR, Tahmasbi AM and Nazari MR (2015). The estimation of ruminal protein degradation parameters of various feeds using in vitro modified gas production technique. *Iranian Journal of Veterinary Research*, 16(1):47-52. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4789239/pdf/ijvr-16-047.pdf>
- Fluharty FL, Zerby HN, Lowe GD, Clevenger DD and Relling AE (2017). Effects of feeding corn silage, pelleted, ensiled, or pelleted and ensiled alfalfa on growth and carcass characteristics of lamb. *South African Journal of Animal Science*, 47(5):704-711. <http://dx.doi.org/10.4314/sajas.v47i5.14>
- García EM, López A, Zimmerman M, Hernández O, Arroquy JI and Nazareno MA (2019). Enhanced oxidative stability of meat by including tannin-rich leaves of woody plants in goat diet. *Asian-Australasian Journal of Animal Sciences*, 32(9):1439-1447. <https://doi.org/10.5713/ajas.18.0537>
- Gawat M, Boland M, Singh J and Kaur L (2023). Goat meat: production and quality attributes. *Foods*, 12:3130. <https://doi.org/10.3390/foods12163130>
- Guzmán JL, Delgado-Pertíñez M, Beriáin MJ, Pino R, Zarazaga LA and Horcada A (2020). The use of concentrates rich in orange by-products in goat feed and its effects on physico-chemical, textural, fatty acids, volatile compounds and sensory characteristics of the meat of suckling kids. *Animals*, 10:766. <https://doi.org/10.3390/ani10050766>
- Kafle D, Lee JH, Min BR and Kouakou B (2021). Carcass and meat quality of goats supplemented with tannin-rich peanut skin. *Journal of Agriculture and Food Research*, 5:100159. <https://doi.org/10.1016/j.jafr.2021.100159>
- Kim H-J, Kim H-J and Jang A (2019). Nutritional and antioxidative properties of black goat meat cuts. *Asian-Australasian Journal of Animal Sciences*, 32(9):1423-1429. <https://doi.org/10.5713/ajas.18.0951>
- Lee S-H, Lee J, Chowdhury MMR, Jeon D, Lee S-S, Kim S, Kim D H and Kim K-W (2019). Grazing Behavior and Forage Selection of Goats (*Capra hircus*). *Journal of the Korean Society of Grassland and Forage Science*, 39(3):189-194. <https://doi.org/10.5333/KGFS.2019.39.3.189>
- Lee J, Kim H-J, Lee S-S, Kim K-W, Kim D-K, Lee S-H, et al. (2023). Effects of diet and castration on fatty acid composition and volatile compounds in the meat of Korean native black goats. *Animal Bioscience*, 36(6):962-972. <https://doi.org/10.5713/ab.22.0378>
- Makmur M, Zain M, Marlinda Y, Khasrad K and Jayanegara A (2020). *In vitro* ruminal biohydrogenation of C18 fatty acids in mixtures of *Indigofera zollingeriana* and *Brachiaria decumbens*. *Journal of the Indonesian Tropical Animal Agriculture*, 45(2):124-135. <https://doi.org/10.14710/jitaa.45.2.124-135>
- Phesatcha K, Phesatcha B and Wanapat M (2022). Mangosteen peel liquid-protected soybean meal can shift rumen microbiome and rumen fermentation end-products in lactating crossbred Holstein Friesian cows. *Frontiers in Veterinary Science*, 8:772043. <https://doi.org/10.3389/fvets.2021.772043>
- Rezende MPG, Figueiredo GC, Araujo JIM, Campos BM, Moretti R, Bozzi R, Malhado CHM, de Souza Jr AAO and Carneiro PLS (2020). Growth curve, carcass traits and klieber ratio of Dorper crossbreed with hairless native Brazilian sheep breeds. *Small Ruminant Research*, 192(2020):106190. <https://doi.org/10.1016/j.smallrumres.2020.106190>
- Rooke J, Brookes I and Armstrong D (1983). The digestion of untreated and formaldehyde-treated soya-bean and rapeseed meals by cattle fed a basal silage diet. *The Journal of Agricultural Science*, 100(2):329-342. <https://doi.org/10.1017/S0021859600033487>
- Santana A, Cajarville C, Mendoza A and Repetto JL (2017). Combination of legume-based herbage and total mixed ration (TMR) maintains intake and nutrient utilization of TMR and improves nitrogen utilization of herbage in heifers. *Animal*, 11(4):616-624. <https://doi.org/10.1017/S1751731116001956>
- Slimeni O, Hajji H, Mekki I, Smeti S, Mahouachi M, Saidani F, et al. (2022). Is it possible to increase goat meat production under Mediterranean forest conditions using small amounts of concentrate without deterioration of its quality? *Animal Science Papers and Reports*, 40(3):335-350. Institute of Genetics and Animal Biotechnology of the Polish Academy of Sciences, Jastrzębiec, Poland. <https://www.igbzpan.pl/uploaded/FSiBundleContentBlockBundleModelTranslatableBlockTranslatableFilesElement/filePath/2206/str335-350.pdf>

Stashenko E and Martínez JR (2014). Gas Chromatography - Mass Spectrometry. Available at: <https://www.intechopen.com/chapters/46209>

Steel RGD and Torrie JH (1980). Principles and Procedures of Statistics. McGraw-Hill Book Co. Inc. New York.

Wang Z, Yu Y, Li X, Xiao H, Zhang P, Shen W, Wan F, He J, Tang S, Tan Z, Wu D and Yao H (2021). Fermented soybean meal replacement in the diet of lactating Holstein dairy cows: modulated rumen fermentation and ruminal microflora. *Frontiers in Microbiology*, 12:625857. <https://doi.org/10.3389/fmicb.2021.625857>.

Widyobroto BP, Budhi SPS and Agus A (2010). Effect of protein undegraded supplementation on production and composition of milk in dairy cows. *Journal of the Indonesian Tropical Animal Agriculture*, 35 (1): 27 - 33. DOI: <https://doi.org/10.14710/jitaa.35.1.27-33>

Publisher's note: Scienceline Publication Ltd. remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access: This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <https://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2025

HEMATOLOGICAL AND BIOCHEMICAL PARAMETERS OF CAPTIVE FALLOW DEER (*Dama dama*) IN A ZOO ENVIRONMENT

Nejra HAŽIMUSIĆ¹   , Vedad ŠKAPUR² , Dženita HADŽIJUNUZOVIĆ-ALAGIĆ¹ , and Amela LIVNJAK¹ 

¹University of Sarajevo - Veterinary faculty, Department for Veterinary Clinical Sciences, Zmaja od Bosne 90, Sarajevo, Bosnia and Herzegovina

²University of Sarajevo - Faculty of Agriculture and Food Sciences, Zmaja od Bosne 8, Sarajevo, Bosnia and Herzegovina

[✉]Email: nejra.hadzimusic@vfs.unsa.ba

[➤]Supporting Information

ABSTRACT: Accurate health assessment of wild, semi-captive, or domesticated animals is essential for their well-being. Despite this necessity, limited studies have been conducted on deer species, and there is a paucity of information on the hemato-biochemical parameters of different deer species globally. Present study aimed to fill this gap by determining the hematological and serum biochemical parameters of fallow deer (*Dama dama*) maintained in semi-captivity within zoo environments for the first time in Bosnia and Herzegovina. Present research involved six healthy male fallow deer, aged 2 to 5 years. The deer were immobilized using xylazine hydrochloride and ketamine hydrochloride, and blood samples were collected from the external jugular vein. The hematological parameters measured included RBC, PCV, HGB, MCV, MCH, MCHC, RDW, RETIC, WBC, WBC differential, PLT, MPV, PDW, and PCT. Biochemical parameters included glucose, urea, creatinine, albumin, triglycerides, cholesterol, and enzymes (AST, ALT, ALKP, and GGT) activities. The results showed the higher glucose and urea concentrations and the same values for creatinine, triglycerides, and enzyme activities when compared to some previous reports. These findings highlighted the importance of considering handling methods and environmental conditions when interpreting biochemical parameters, contributing to improved health assessments and management practices for deer in captivity.

Keywords: Biochemical and hematological parameters, Captive wildlife, Domesticated animals, Fallow deer.

Abbreviations: ALB: Albumin; ALKP: Alkaline Phosphatase; ALT: Alanine Aminotransferase; AST: Aspartate Aminotransferase; BASO: Basophils; CHOL: Cholesterol; CREA: Creatinine; EDTA K: Ethylenediaminetetraacetic Acid Potassium Salt; EOS: Eosinophils; GGT: Gamma-Glutamyl Transferase; GLU: Glucose; HGB: Hemoglobin; LYM: Lymphocytes; MCH: Mean Corpuscular Hemoglobin; MCHC: Mean Corpuscular Hemoglobin Concentration; MCV: Mean Corpuscular Volume; MONO: Monocytes; MPV: Mean Platelet Volume; NEU: Neutrophils; PCT: Plateletcrit; PCV: Packed Cell Volume; PDW: Platelet Distribution Width; PLT: Platelets; RBC: Red Blood Cells; RDW: Red Cell Distribution Width; RETIC: Reticulocytes; TG: Triglycerides; WBC differential: White Blood Cells Differential; WBC: White Blood Cells.

INTRODUCTION

Accurately assessing the health of wild, semi-captive, or domesticated animals is crucial for their wellbeing. However, there have been very few studies conducted on deer species, and limited reports exist on the haemato-biochemical parameters of different deer species worldwide (Gupta et al., 2007; Sinanović et al., 2013; Vukšić et al., 2016).

While normal hematological and serum biochemical values for several deer species are limited in the literature, some studies have established reference ranges for certain wild species. For instance, Rosef et al. (2004) provided hematological and serum biochemical reference values for free-ranging red deer (*Cervus elaphus atlanticus*) in Norway. Similarly, Miller et al. (2013) reported biochemical and hematologic reference values for free-ranging, chemically immobilized wild Norwegian reindeer (*Rangifer tarandus tarandus*) during early winter. Additionally, Karpiński et al. (2023) presented hematology, and serum chemistry values for free-ranging roe deer (*Capreolus capreolus*) in Poland. These studies provide valuable baseline data for health assessments and disease diagnosis in these species. However, for other deer species lacking specific reference values, comparisons are often made using baseline data from domestic small ruminants such as sheep and goats (Gupta et al., 2007).

The fallow deer (*Dama dama*) is a native Eurasian wild species of cervid (Pastrana et al., 2022) and among the most common cervid species in Europe and the most widely distributed cervid globally. Although the fallow deer has been introduced to most parts of Europe, it is native only to southern Anatolia, Sicily, southern Italy, and the southern Balkan peninsula. However, distribution data for the fallow deer in Bosnia and Herzegovina is not available, as they are only found in reserves (Bijl and Csányi, 2022). Fallow deer are the most common deer species found in both wild and captive environments in Bosnia and Herzegovina.

SHORT COMMUNICATION
 PII: S222877012500006-15
 Received: August 08, 2024
 Revised: March 20, 2025
 Accepted: March 22, 2025

Several studies have identified differences in blood values among deer, which can be attributed to various factors including farming conditions, management practices, and sampling techniques. Methods such as collection from pasture, yarding, drafting, indoor confinement, isolation, and catheterisation have been shown to induce stress in deer (Vengušt et al., 2006). These variations in blood values can also result from genetic, environmental, nutritional, and physiological factors, as well as the stress of capture and the influence of different blood sampling techniques (Vengušt et al., 2006).

Assessing the health of wild, semi-captive, or domesticated animals is vital, yet studies on deer species and their haemato-biochemical parameters are limited (Gupta et al., 2007). Differences in blood values among deer have been linked to farming conditions, management, and sampling techniques (Vengušt et al., 2006). In the absence of specific data, comparisons are often made with baseline values from domestic small ruminants like sheep and goats (Gupta et al., 2007).

Fallow deer are the most common deer species found in both wild and semi-captive environments in Bosnia and Herzegovina. They are residents of forested areas and are also kept in semi-captivity in zoos. The data reported here detail the typical hematological and serum biochemical parameters of clinical importance for a deer species raised in captivity. Specifically, these parameters pertain to fallow deer maintained in captivity within zoo environments.

MATERIALS AND METHODS

The research was conducted on six male fallow deer (*Dama dama*), aged between 2 and 5 years, housed in the zoo in Sarajevo – Pionirska dolina, Sarajevo, Bosnia and Herzegovina; 43° 52' 41.8"N 18° 24' 44.1"E; elevation 518 m). The animals were healthy, well-fed, and clinically healthy. The deer were immobilized using a combination of 100 mg xylazine hydrochloride and 300 mg ketamine hydrochloride. Following immobilization, blood was collected from the external jugular vein. For hematological analysis, blood samples were collected into tubes containing the EDTA K as an anticoagulant. Hematological analysis was conducted within 2 to 3 hours after sampling using an automated veterinary hematology analyzer – The ProCyte IDEXX, PRC 1025236.

The following parameters were determined: red blood cells (RBC), packed cell volume (PCV), hemoglobin (HGB), the mean corpuscular volume (MCV), the mean corpuscular hemoglobin (MCH), the mean corpuscular hemoglobin concentration (MCHC), the red blood cell distribution width (RDW), reticulocyte count (RETIC), white blood cells (WBC), white blood cells differential (WBC-D), the platelets (PLT), the mean platelet volume (MPV), the platelet distribution width (PDW), and the plateletcrit (PCT). Plain tubes were used to collect serum for the analysis of biochemical parameters, including glucose, urea, creatinine, albumin, triglycerides, cholesterol, and the activity of enzymes AST, ALT, ALKP, and GGT. Serum biochemical parameters were determined by the IDEXX Catalyst One veterinary chemistry analyzer, REF 89-92525-00.

Ethical Regulations and animal welfare

In present study, all procedures involving animals were conducted in accordance with recognized standards for animal welfare and ethical research. The fallow deer (*Dama dama*) were housed in managed care within a zoo environment, specifically in the Pionirska Dolina Zoo in Sarajevo, Bosnia and Herzegovina. Their health and well-being were closely monitored by experienced veterinarians, ensuring that all animals were kept in optimal conditions and handled according to established welfare protocols. Blood sampling was performed as part of routine veterinary care, and all immobilization and handling techniques were aimed at minimizing stress and discomfort to the animals. As the blood sampling occurred during routine health monitoring, it did not require specific approval from an ethical committee. However, all efforts were made to follow best practices for animal care and welfare, and the animals were never subjected to unnecessary pain or distress.

Statistical analysis

The results were analysed statistically. The values are presented throughout as a mean value and standard deviation (SD). Statistical analysis was performed by means of the SPSS package (SPSS Inc., Chicago, Illinois, USA).

RESULTS AND DISCUSSION

The hematological and biochemical analysis of six male fallow deer (*Dama dama*) maintained in a zoo environment revealed several notable findings. The glucose and urea concentrations were higher compared to some previous studies, while creatinine, triglyceride, and enzyme activity levels remained consistent with prior reports. The hematological parameters, including RBC, PCV, HGB, MCV, MCH, and MCHC, aligned closely with previously published values.

Table 1 - Mean values of haematology parameters in male fallow deer

Parameters	Mean \pm SD
RBC ($\times 10^{12}/L$)	10.81 \pm 2.06
PCV (L/L)	44.3 \pm 37.1
HGB (g/L)	143.3 \pm 23
MCV (fL)	41.2 \pm 1.48
MCH	14.27 \pm 0.6
MCHC	34.67 \pm 0.59
RDW	31.97 \pm 2.8
%RETIC	0
RETIC	1.13 \pm 1.27
WBC ($\times 10^9/L$)	1.95 \pm 0.94
%NEU	56.27 \pm 6.26
%LYM	23.03 \pm 2.47
%MONO	5.6 \pm 2.1
%EOS	13.23 \pm 2.43
%BASO	1.87 \pm 0.25
NEU	1.07 \pm 0.44
LYM	0.46 \pm 0.26
MONO	0.11 \pm 0.06
EOS	0.27 \pm 0.17
BASO	0.04 \pm 0.02
PLT	265.33 \pm 92.45
MPV	7.77 \pm 0.51
PDW	6.55 \pm 0.35
PCT	0.21 \pm 0.07

Table 2 - Mean values of biochemistry parameters in male fallow deer

Parameters	Mean \pm SD
GLU (mmol/L)	9.71 \pm 2.35
UREA (mmol/L)	6.31 \pm 2.04
CREA ($\mu\text{mol}/L$)	127 \pm 20.58
ALB (g/L)	48 \pm 2.67
TG (mmol/L)	0.52 \pm 0.46
CHOL (mmol/L)	1.99 \pm 0.12
AST (U/L)	99 \pm 36.78
ALT (U/L)	36 \pm 5.87
ALKP (U/L)	212 \pm 19.87
GGT (U/L)	38 \pm 6

Existing literature provides data on the blood parameters of deer which vary depending on the sampling technique used, including chemical immobilization (Peinado et al., 1999; Poljičak-Milas et al., 2004), physical restraint (Rehbein et al., 1999), or post-culling (Vengušt et al., 2002). The RBC values determined by present research are slightly lower than those determined by Tajchman et al. (2023) and the values determined by Vukšić et al. (2016), but correspond to the values determined by Vengušt et al. (2006) for deer after sedation and culling.

Present study measured PCV, HGB, MCV, MCH, and MCH levels in deer, aligning closely with the findings with the values reported earlier (Vengušt et al., 2006) following sedation and culling. Comparable values were also reported by Kováč et al. (1997), as well as by Barić Rafaj et al. (2011) in red deer. Vukšić et al. (2016) noted that the hemoglobin concentration ranged from 157.00 to 164.00 g/L, averaging 160.50 g/L, which was similar to the concentration observed in present study but higher than that reported by Vengušt (2002). Present research identified significantly lower WBC values compared to other studies (Vengušt et al., 2006; Vukšić et al., 2016). Vukšić et al. (2016) determined the average leukocyte count in adults ($7.07 \times 10^9/L$), which was higher compared to young deer, indicating that age influences the value of this parameter.

The PLT value was determined in fallow deer to be 265.33 ± 92.45 , while [Vukšić et al. \(2016\)](#) reported a mean PLT value of 161.78. Additionally, [Barić-Rafaj et al. \(2011\)](#) found a PLT value of 262 ± 118 in adult red deer. The significant difference between present findings and those of [Vukšić et al. \(2016\)](#), might be attributed to variations in sampling methods, environmental conditions, the health and physiological status of the animals, or differences in the populations studied. Interestingly, present PLT values are closer to those reported by [Barić-Rafaj et al. \(2011\)](#) for adult red deer, suggesting that species differences or age-related factors might play a role. Further investigation is needed to understand these discrepancies and to establish more comprehensive reference ranges for fallow deer and other cervids in different environments.

Red cell distribution width (RDW) measures the variation in erythrocyte size using their MCV (Mean Corpuscular Volume). [Barić-Rafaj et al. \(2011\)](#) in their study on farmed red deer, determined that the RDW was significantly higher in fawns. Although RDW determination is widely accepted in human medicine, there is limited information about this parameter in veterinary medicine, particularly concerning wild animals. The values for WBC and the differential blood count are presented in Table 1. [Vukšić et al. \(2016\)](#) determined a WBC value of 7.07 in adult fallow deer. [Barić-Rafaj et al. \(2011\)](#) found a WBC value of 15.41 ± 4.87 in 11 adult red deer. [Vengušt et al. \(2006\)](#) reported WBC values in fallow deer as follows: 9.1 ± 1.2 for restrained deer, 3.6 ± 0.9 for tranquilized deer, and 2.9 ± 1.3 for shot deer. In contrast, present research determined a WBC value of 1.95 ± 0.94 .

These differences highlight the significant variability in WBC counts depending on factors such as species, handling methods, and the physiological state of the animals. Present findings, which show lower WBC values compared to other studies ([Vengušt et al., 2006](#)) may be influenced by the specific conditions under which our samples were taken. This underscores the importance of considering these variables when interpreting hematological data and establishing reference ranges for wildlife.

Present research also determined values of some biochemical parameters (Table 2). [Vengušt et al. \(2006\)](#) determined the glucose values in deer as follows: 2.9 ± 0.4 mmol/L for restrained deer, 8.5 ± 2.1 mmol/L for tranquilized deer, and 7.5 ± 3.2 mmol/L for deer that were shot. These values indicate that glucose levels vary significantly depending on the method of restraint or sedation. The glucose values determined in present research correspond to the values found for tranquilized deer. However, in research conducted by [Vengušt and Bidovec \(2002\)](#) authors described lower glucose values compared to those determined in present research. This discrepancy highlights potential differences in environmental conditions, handling, or physiological states of the deer between the studies.

The serum glucose concentration in fallow deer may exhibit significant individual variation ([Rehbein et al., 1999](#); [Slavica et al., 2000](#)). [Wilson and Pauli \(1983\)](#) reported similar results in red deer. Compared to domestic ruminants, deer have higher serum glucose levels, which may be due to their more nervous temperament or higher metabolic rate ([Wilson and Pauli, 1983](#)).

Present research determined urea concentration of 6.31 ± 2.04 mmol/L in deer, which presents an interesting comparison to the values obtained by other studies ([Vengušt et al., 2006](#)). While present methodology also involved the use of sedation, obtained values are somewhat different from those reported by [Vengušt et al. \(2006\)](#). In previous studies, the urea values were reported as follows: 9.8 ± 3.2 mmol/L for restrained deer, 8.1 ± 0.7 mmol/L for tranquilized deer, and 6.5 ± 1.6 mmol/L for deer that were shot. These values indicate that the method of handling and sedation significantly impacts the biochemical parameters measured in deer. For instance, the highest urea concentration was observed in restrained deer, likely due to the stress response elicited by physical restraint. Tranquilized deer showed slightly lower urea levels, which can be attributed to the calming effects of sedation that reduce stress-induced metabolic changes. Deer that were shot had the lowest urea concentrations, potentially due to the rapid physiological changes occurring at the time of death, affecting metabolic waste levels. Although present research employed sedation similar to the aforementioned studies, the urea concentration we observed (6.31 ± 2.04 mmol/L) is somewhat different from the 8.1 ± 0.7 mmol/L reported for tranquilized deer in research conducted by [Vengušt et al. \(2006\)](#). This discrepancy could be due to differences in sedation protocols, environmental conditions, or the physiological status of the animals at the time of sampling. These variations underscore the importance of considering the method of animal handling and specific research conditions when interpreting biochemical parameters.

This research determined that the concentrations of creatinine and triglycerides were similar to those reported by [Vengušt et al. \(2006\)](#). This consistency suggests that, despite some variations in urea levels, the biochemical responses related to creatinine and triglycerides in present study align with existing literature ([Vengušt et al., 2006](#)). This finding reinforces the reliability of present methods and the comparability of present results with previous research. This research determined that the enzyme activities were similar to those reported in other studies ([Vengušt et al., 2006](#); [Sinanović et al., 2013](#)). This similarity indicates that the enzymatic responses observed in present study align well with existing literature ([Vengušt et al., 2006](#); [Sinanović et al., 2013](#)), further validating our methods and ensuring the comparability of present results with prior studies in this field.

Although [Sinanović](#) published a preliminary report in 2013 ([Sinanović et al., 2013](#)) on certain biochemical parameters in deer, present research is the first study in Bosnia and Herzegovina to determine both hematological and biochemical parameters in deer blood. The obtained values of hematological and biochemical parameters relate to deer

that were sedated for sampling purposes. Numerous authors mention significant differences in the frequency of the tested parameters depending on whether the animals were sedated, restrained, or shot and then sampled (Vengušt et al., 2006). Some researchers have proposed that two separate ranges of reference blood values should be established for wild animals, based on the capture method used (Vengušt et al., 2006).

CONCLUSION

This study provides essential baseline data on the hematological and biochemical parameters of captive fallow deer in Bosnia and Herzegovina. The findings highlight significant variations in these parameters depending on the method of restraint and sedation used during sampling. The glucose and urea concentrations determined in present research were higher than those reported in some previous studies, indicating potential influences from environmental conditions, handling techniques, and physiological states of the deer. Despite these variations, the consistency observed in creatinine, triglyceride concentrations, and enzyme activities with existing literature suggests that present methods are reliable and comparable to another research in the field. The hematological parameters such as RBC, PCV, HGB, MCV, MCH, and MCHC levels aligned closely with earlier findings, further validating our approach. Also, this study is the first comprehensive report in Bosnia and Herzegovina to document both hematological and biochemical parameters in deer blood, specifically under sedation. Present results emphasize the importance of standardized protocols in wildlife health assessments to ensure accurate and comparable data across different studies and regions. Establishing separate reference ranges for wild animals based on capture methods, as proposed by some researchers, could enhance the precision of health evaluations and contribute to better wildlife management and conservation practices. The results emphasize the importance of standardized protocols in wildlife health assessments to ensure accurate and comparable data across different studies and regions. Establishing separate reference ranges for wild animals based on capture methods, as proposed by some researchers, could enhance the precision of health evaluations and contribute to better wildlife management and conservation practices. In summary, present study underscores the need for continuous monitoring and evaluation of deer health in both wild and captive environments. The data generated from this study will serve as a valuable reference for veterinarians and wildlife biologists in assessing the health and well-being of fallow deer and other cervid species.

DECLARATIONS

Corresponding author

Correspondence and requests for materials should be addressed to Nejra Hadžimusić; Email: nejra.hadzimusic@vfs.unsa.ba; ORCID: 0000-0001-9278-1876

Authors' contributions

N.Hadžimusić contributed to the design of the study, data analysis, and the writing of the manuscript. N.Hadžimusić, A.Livnjak, DŽ.Hadžijunuzović-Alagić and V.Škapur were responsible for sample collection, laboratory analysis, and manuscript review. All authors have contributed to the interpretation of the data and approved the final manuscript for submission.

Ethics committee approval

As the blood sampling was performed during routine veterinary health checks and involved no invasive procedures beyond standard care. The study strictly adhered to internationally recognized animal care guidelines, ensuring minimal stress and maximum welfare for the animals throughout the study.

Acknowledgements

We would like to thank the staff of the Pionirska Dolina Zoo in Sarajevo, Bosnia and Herzegovina, for their cooperation and assistance during the study. We also appreciate the support provided by the veterinary team for their help with animal care and sample collection.

Consent to publish

All authors have read and approved the final version of the manuscript and give their consent for publication.

Competing Interests

The authors declare that there are no competing interests regarding the publication of this paper.

REFERENCES

- Barić Rafaj R, Tončić J, Vicković I and Šoštar B (2011). Haematological and biochemical values of farmed red deer haematological and biochemical values of farmed red deer (*Cervus elaphus ervus elaphus*). Veterinarski Arhiv. 81: 513-523. <https://hrcak.srce.hr/70942>
- Bijl H and Csányi S (2022). Fallow deer (*Dama dama*) population and harvest changes in Europe since the early 1980s. Sustainability. 14: 12198. <https://doi.org/10.3390/su141912198>
- Gupta AR, Patra RC, Saini M and Swarup D (2007). Haematology and serum biochemistry of chital (*Axis axis*) and barking deer (*Muntiacus muntjak*) reared in semi-captivity. Veterinary research communications. 31: 801-808. <https://doi.org/10.1007/s11259-006-0095-8>
- Karpiński M, Czyżowski P, Beeger S and Flis M (2023). Hematological and serum biochemical values of free-ranging Roe deer (*Capreolus capreolus*) in Poland. Animals. 13: 242. <https://doi.org/10.3390/ani13020242>
- Kováč G, Ciberej J, Paulikova I, and Seidel H (1997). Haematological indices in fallow deer. Acta Veterinaria Brno, 66(4):203-11. https://actavet.vfu.cz/media/pdf/avb_1997066040203.pdf
- Miller AL, Evans AL, Os Ø and Arnemo JM (2013). Biochemical and hematologic reference values for free-ranging, chemically immobilized wild Norwegian reindeer (*Rangifer tarandus tarandus*) during early winter. Journal of Wildlife Diseases. 49: 221-228. <https://doi.org/10.7589/2012-04-115>
- Pastrana CI, Gonzalez FJN, Inostroza MGP, Arbulu AA, Bermejo JVD and Aguilera MJR (2022). Study of variability of cognitive performance in captive fallow deer (*Dama dama*) through g and c factors. Journal of Veterinary Behavior. 47: 70-85. <https://doi.org/10.1016/j.jveb.2021.10.001>
- Peinado VI, Celdrán JF and Palomeque J (1999). Basic hematological values in some wild ruminants in captivity. Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology. 124: 199-203. [https://doi.org/10.1016/S1095-6433\(99\)00110-5](https://doi.org/10.1016/S1095-6433(99)00110-5)
- Poljičak-Milas N, Slavica A, Janicki Z, Robić M, Belić M and Milinković-Tur S (2004). Serum biochemical values in fallow deer (*Dama dama* L.) from different habitats in Croatia. European journal of wildlife research. 50: 7-12. <https://www.proquest.com/docview/820372839?sourceopen=Scholarly%20Journals>
- Rehbein S, Bieneschek S, Sachse M and Neubert E (1999). Hämatologische und klinisch-chemische Untersuchungen bei natürlich und bei mütterlos aufgewachsenen Damhirschen (*Dama dama* L.) [Hematological and clinical-chemical studies in naturally and motherless-raised fallow deer (*Dama dama* L.) -2. Communication: Clinical-chemical examination in blood plasma]. Zoologische Garten. 69: 89-108.
- Rosef O, Nystøyl HL, Solenes T and Arnemo JM (2004). Haematological and serum biochemical reference values in free-ranging red deer (*Cervus elaphus atlanticus*). Rangifer. 24: 79-85. <https://doi.org/10.7557/2.24.2.304>
- Sinanović N, Zahirović A, Čamo D, Čoralčić A and Čutuk A (2013). Biochemical blood parameters of fallow deer (*Dama dama*) from farms in Bosnia and Herzegovina. Veterinaria. 62: 55-60. <https://veterinaria.unsa.ba/journal/index.php/vfs/article/view/110>
- Slavica A, Janicki Z, Barić Rafaj R, Kolić E, Manojlović I and Deždek D (2000). Biochemical blood analysis of the fallow deer (*Dama dama* L.) from the Brijuni islands, Croatia. Veterinarski Arhiv. 70 (Supplement): S193-199 <https://www.croris.hr/crosbi/publikacija/prilog-casopis/111685>
- Tajchman K, Kowalik S, Janiszewski P, Licznerna K and Bogdaszewski P (2023). Basic haematological and biochemical parameters of farmed red deer and fallow deer blood. Medycyna Weterynaryjna. 79:286-290. <http://www.medycynawet.edu.pl/images/stories/pdf/pdf2023/062023/2023066775.pdf>
- Vengušt G, and Bidovec A (2002). Some serum chemistry values of fallow deer (*Dama dama* L.) in Slovenian hunting enclosures. Veterinarski arhiv. 72(4): 205-212. <https://hrcak.srce.hr/80028>
- Vengušt G, Klinkon M, Vengušt A, and Bidovec A (2002). Biochemical parameters in blood of farmed fallow deer (*Dama dama*). Zeitschrift für Jagdwissenschaft, 48(4):226. <https://www.cabidigitallibrary.org/doi/full/10.5555/20033163201>
- Vengušt G (2002). Blood parameters in deer: 1. Haematological values and leukocyte count in differential blood pictures from several species, with special emphasis on fallow deer. Veterinarske Novice, 28(10):405-411. <https://www.cabidigitallibrary.org/doi/full/10.5555/20023177785>
- Vengušt G, Žele D, Kobal S and Bidovec A (2006). Haematological and biochemical values of farmed fallow deer (*Dama dama*) after using different methods of capture. Veterinarski arhiv. 76: S189-S197. <https://vetarhiv.vef.unizg.hr/papers/2006-76-7-22.pdf>
- Vukšić N, Bešlo D, Agić D and Šperanda M (2016). Hematološki i biokemijski pokazatelji u jelena lopatara (*Dama dama* L.) na području državnog otvorenog lovišta [Hematological and biochemical indicators in fallow deer (*Dama dama* L.) in the state open hunting area]. Krndija II ~ XIV/23. In 51. croatian 11. International symposia, University of Zagreb, Zagreb, pp. 284-288. <https://repositorij.fazos.hr/islandora/object/pfos:3894>
- Wilson PR and Pauli JV (1983). Blood constituents of farmed red deer (*Cervus elaphus*). 2: biochemical values. The New Zealand Veterinary Journal. 31: 1-3. <https://doi.org/10.1080/00480169.1983.34943>

Publisher's note: Scienceline Publication Ltd. remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access: This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <https://creativecommons.org/licenses/by/4.0/>.

EFFECT OF GRADED LEVELS OF DIETARY TOMATO WASTE ON PERFORMANCE AND CARCASS CHARACTERISTICS OF JAPANESE QUAIL REARED UNDER INTENSIVE SYSTEM

Shame BHAWA , John Cassius MOREKI  and Freddy MANYEULA 

Department of Animal Science, Faculty of Animal and Veterinary Sciences, Botswana University of Agriculture and Natural Resources, Private Bag 0027, Content Farm, Sebele, Gaborone, Botswana

✉Email: jmoreki@buan.ac.bw

↳Supporting Information

ABSTRACT: This study was carried out to evaluate the effects of partial replacement of soybean meal (SM) with tomato waste (TW) in Japanese quail diets on the resulting yield, internal organs, and carcass characteristics. Eighty unsexed 1-day-old chicks were housed in battery cages with cardboard boxes used as solid floors and randomly assigned to 1 of 4 dietary groups, 46.2% SM, 44.2% SM + 2% TW, 42.2% SM + 4% TW, or 40.2% SM + 6% TW, over a 6 weeks growth period. Yields and carcass characteristics were then determined. Data were analysed using the General Linear Model (GLM) procedures followed by a response procedure for surface regression analysis (Proc RSREG; SAS 9.4) to describe the parameters' responses to graded levels of dietary tomato waste. Repeated measures analysis showed significant week × diet interaction effects on feed intake (FI, $P = 0.03$), body weight gain (WG, $P = 0.0006$), feed conversion ratio (FCR, $P = 0.002$), protein efficient ratio ($P = 0.0001$), and growth efficiency ($P = 0.0001$). By supplementing the diets of quails with a 2% inclusion level, a diet significantly affected quails' FI on weeks 1, 2, 3, and 6. A diet containing 2% TW significantly affected live weight (LW), hot carcass weight (HCW), and cold-dressed weight (CDW). It is concluded that the dietary supplementation with 44.2% SM + 2% TW seemed ideal for optimum performance in Japanese quails based on the insignificant change in feed intake and growth efficiency results compared to 46.2% SM for weeks 1 and 2. Further research is needed on the application method that could be used to enhance the utilization of tomato waste in Japanese quails.

Keywords: Carcass characteristics, Dietary replacements, Growth performance, Japanese quails, Tomato waste.

INTRODUCTION

Due to domestic birds' short gestation and generation intervals, high productivity, rapid turnover rate, higher feed efficiency, and cheap land and labour needs, poultry products including meat and eggs have been encouraged for regular intake to fulfill the protein deficit (Olawumi, 2015). In many countries, including Botswana, the Japanese quail (*Coturnix coturnix japonica*) is quickly gaining recognition as a source of meat and eggs. In contrast to other poultry species, the Japanese quail has a sexually dimorphic body structure, with females being larger than males (Sezer et al., 2006). Bonos et al. (2010) reported that female quail have heavier carcasses and bodies than their male counterparts.

As a result of the lack of animal protein caused by the declining supply of meat and eggs in developing nations, rearing Japanese quail as food is seen as another aspect of poultry farming (Illgner and Nel, 2000; Agiang et al., 2011; Dalle Zotte and Cullere, 2024). The Japanese quail produces meat and eggs that are highly prized for their distinctive flavor (Ayaşan, 2013). Furthermore, the Japanese quail serves as a laboratory animal (Ophir et al., 2005), and has unique traits notably quick growth rates which allow the bird to be sold for human consumption at 5-6 weeks of age (Hemid et al., 2010). Compared quails to chickens and found that quails reach sexual maturity earlier leading to a small generation gap, have higher laying rates, and significantly reduced space requirements (Hemid et al., 2010).

The escalating costs of cereals and imported feedstuffs for chicken diets have driven a search for substitute ingredients that will be available as by-products from domestic agricultural producers (Adeniji and Oyeleke, 2008; Okello et al., 2023; Dou et al., 2024). The increased production of soybeans is causing worry because of greenhouse gas emissions and the devastation of wildlife habitats (Siamabele, 2018). Rahman et al. (2016) posited that utilizing alternate feed materials in chicken rations is a crucial aspect of effective poultry production in countries experiencing a food shortage. In many parts of the world, using alternative feed ingredients such as tomato pomace with a high protein value of 17-24% (Lu et al., 2022) for chicken diets is crucial to the poultry industry's success (Yitbarek, 2013). Therefore, the current experiment was conducted to determine the effect of feeding graded levels of tomato waste on the growth, slaughter performance, and carcass characteristics of Japanese quail reared under an intensive system. It was

RESEARCH ARTICLE
 PII: S222877012500007-15
 Received: September 01, 2024
 Revised: March 27, 2025
 Accepted: March 28, 2025

hypothesized that replacing soybean meal (SM) with tomato waste in quail diets would not affect growth, slaughter performances, and carcass traits.

MATERIALS AND METHODS

Description of the study area

The study was conducted at BUAN farm in South-East Botswana. The farm is located at coordinates 24° 34'54.41" S 25° 58'14.64" E (Google Earth Pro, 2022). The farm is about 15 km north of Gaborone and is at an elevation of 978 m (Google Earth Pro, 2022). The vegetation type is Savannah, with tall grasses, bushes, and trees. Precipitation in Gaborone is about 457 mm per year (Climate-Data.Org, 2022). In Gaborone, the average temperature of the coldest month (July) is 13.5 °C, and that of the warmest month (January of 2022) is 26 °C.

Source and sample preparation

Tomato waste was procured from the NAFTEC Investments plant in Selebi-Phikwe, located 404 km northeast of Gaborone, the capital of Botswana. Tomato waste was dried for 4 days in February 2023 (Aragaw et al., 2021) by spreading it in the sun. After reducing the particle size of the dried tomato waste by hand, it was run through a grinder (Polymix PX-MFC 90 D model, Kinematica™, Switzerland) and passed through a 1 mm sieve. Thereafter, samples were weighed and stored at room temperature in the Nutrition Laboratory storage room at BUAN before being subjected to chemical analysis and later used in this experiment.

Proximate analyses

Tomato waste was subjected to preliminary analysis using the Association of Official Analytical Chemists (AOAC) International methods before diet formulations. After formulations were performed, subsamples of the experimental diets (TW0, TW2, TW4, and TW6) and tomato waste were analysed using methods from AOAC (2005). For dry matter (DM), method number, 930.15 was used. Ash content was determined by ashing at 550 °C for about 6 hours (AOAC, 2005; method number 924.05), whereas nitrogen was determined using the Kjeldahl method (AOAC, 2005; method number 984.13). The percentage of nitrogen was multiplied by 6.25 to determine crude protein (AOAC, 2005; method number, 920.39). Energy content was determined using a bomb calorimeter and measured in joules. Fat was determined using AOAC, (2005); method number, 920.39. Crude fiber was determined following AOAC, (2005); method number, 978.10. Neutral detergent fiber (NDF), and acid detergent fiber (ADF) were determined by refluxing 0.45 g of samples with neutral detergent and acid detergent solutions respectively, for 1 hour using the ANKOM²⁰⁰⁰ fiber analyser (ANKOM Technology, NY, USA). Acid detergent lignin (ADL) was determined by using the ADF residue that was solubilized by 72 % sulphuric acid, leaving the lignin (ADL), which was determined gravimetrically (AOAC, 2005; method number, 973.18). Table 1 shows results from a proximate analysis of tomato waste.

Table 1 - Proximate analysis (% , unless otherwise stated) of tomato waste used in this experimental trial

Parameter	Value
Dry Matter	95.84
Ash	1.58
Crude protein	20.1
Energy (J)	22
Fat	13
Crude fiber	55
Neutral detergent fiber	56
Acid detergent fiber	60
Acid detergent lignin	25

J= Joule

Diet formulations and composition of experimental diets

The following four isocaloric dietary treatments were formulated using Excel spreadsheet 2010 by partially substituting SM with tomato waste in the diets of Japanese quail: TW0 = Diet with no tomato waste (control); TW2 = Diet with 2% SM replaced with tomato waste; TW4 = Diet with 4% SM replaced with tomato waste; TW6 = Diet with 6% SM replaced with tomato waste. The ration was prepared following the National Research Council (1994). Table 2 presents the ingredients and the calculated composition of the regular diet that was in mash form.

Table 2 - Composition of experimental diets (% , unless otherwise stated) used in the experimental trial

Ingredient ¹	Diets ²				Diets ²			
	Starter (%)				Grower (%)			
	TW0	TW2	TW4	TW6	TW0	TW2	TW4	TW6
Maize	42.31	42.31	42.31	42.31	55.79	55.79	55.79	55.79
Sorghum	9.84	9.84	9.84	9.84	9.93	9.93	9.93	9.93
Soybean meal (SM)	46.20	44.20	42.20	40.20	32.70	30.70	28.70	26.70
Tomato waste (TW)	0	2	4	6	0	2	4	6
DCP	0.17	0.17	0.17	0.17	0.02	0.02	0.02	0.02
Premix	0.50	0.50	0.50	0.50	0.47	0.47	0.47	0.47
Salt	0.36	0.36	0.36	0.36	0.36	0.36	0.36	0.36
Limestone	0.10	0.10	0.10	0.10	0.20	0.20	0.20	0.20
Lysine	0.12	0.12	0.12	0.12	0.18	0.18	0.18	0.18
Methionine	0.40	0.40	0.40	0.40	0.35	0.35	0.35	0.35
Calculated nutrient content								
Metabolisable energy (MJ/kg)	12.13	12.12	12.30	13.38	13.38	13.38	13.40	13.40
Crude protein	25.90	25.40	24.90	24.40	21	20.50	20	19.60
Calcium	9.10	9	9	8.90	8.80	8.70	8.70	8.60
Phosphorous	5.40	5.30	5.20	5.10	2.20	2.10	2	1.90
Lysine	13.80	13.80	13.90	13.90	19.60	19.60	19.70	19.70
Methionine	5.20	5.20	5.20	5.20	4.90	4.90	4.90	4.90

¹Ingredient: TW = Tomato waste; DCP = Dicalcium phosphate; Premix supplying per kg feed: 12,000 IU vitamin A, 5,000 IU vitamin D3, 80 mg vitamin E, 7 mg vitamin K, 5 mg thiamine, 6 mg riboflavin, 6 mg pyridoxine, 0.02 mg vitamin B12, 60 mg niacin, 15 mg pantothenic acid, 1.5 mg folic acid, 0.25 biotin, 10 mg vitamin C, 500 mg choline chloride, 100 mg Zn, 120 mg Mn, 20 mg Fe, 15 mg Cu, 0.2 mg, Co, 1 mg I, 0.3 mg Se. ²Diets: TW0 = Diet with no tomato waste; TW2 = Diet with 2% of SM replaced with tomato waste; TW4 = Diet with 4% of SM replaced with tomato waste; TW6 = Diet with 6% of SM replaced with tomato waste.

Experimental design

A total of 80-day-old unsexed quail chicks with an initial weight of 7.6 ± 0.152 g were acquired from Makhaya Quail farm in Gaborone. Japanese quails were randomly assigned to 4 dietary treatment groups at 20 chicks per treatment in a completely randomized design. The treatments were replicated 4 times with 5 chicks in each pen (experimental unit). The one-day-old chicks were housed in battery cages with cardboard boxes used as solid floors and were randomly assigned to 1 of 4 dietary groups, 46.2% SM (TW0), 44.2% SM + 2% TW (TW2), 42.2% SM + 4% TW (TW4), or 40.2% SM + 6% TW (TW6), over a 6 weeks growth period. Yields and carcass characteristics were then determined.

Bird management

The poultry house and equipment were cleaned two weeks before the arrival of quail chicks by using clean water and disinfected with Kenosan detergent from Shield Vet Gaborone. Virocid was used to disinfect the ceiling, walls, and floors. To eradicate and sterilize bugs from the base post crevices, formalin and a salt mixture were applied to the floor, wall junctions, and the surrounding region. Throughout the experimental period, the 750 ml water fonts were used and cleaned daily, and 1 kg feed trays were used. Feed was weighed and replaced every day. Quails were kept in battery cages with firm floors made of cardboard boxes. Sunflower husks were utilised as litter to make the dwelling more comfortable as recommended by Amedu et al. (2018). Feed and water were provided *ad libitum*. The illumination schedule was 23 hours per day for the first week, and thereafter 16 hours per day until the end of the experiment (Manyeula et al., 2019). The temperature of 32 °C was maintained using electric heaters for the first two days and then decreased by 1 °C every other two days until the third week, and thereafter a temperature of 20 °C was maintained for the remainder of the feeding period. Mortality was recorded as it occurred throughout the experimental period.

Data collection procedure

Growth performance parameters

Feed was weighed using Adam's electronic scale sensitive to 0.01 g (Adam scale Pty Ltd, Gaborone, Botswana) in the morning (feed offered) and the following day the refusals were re-weighed and FI was then calculated by subtracting feed offered from the refusal (Tamburawa et al., 2018) as shown in formula 1.

$$AWFI (g/week) = \text{Feed offered (g)} - \text{feed refusal (g)} / 7 \text{days} \dots \dots \dots 1$$

Quail chicks were individually weighed at the beginning of the experimental trial using Adam's electronic scale and subsequently weekly. Weekly weight gain was calculated using formula 2.

$$AWWG (g/week) = \text{Finish weight (g)} - \text{start weight (g)} / 7 \text{days} \dots \dots \dots 2$$

The feed conversion ratio was calculated by dividing feed consumption by body weight gain (BWG) as shown in formula 3.

$$FCR = \text{Feed intake (g)} / \text{weight (g)} \dots \dots \dots 3$$

Weekly protein consumed (PC, g/bird) was calculated by multiplying the concentration of crude protein (CPd) in the diet (g/kg DM consumed) by weekly feed intake (FI) g/bird) as illustrated in formula 4.

$$PC \text{ (g/bird)} = \text{Feed intake (g)} \times \text{crude protein of the diet} \dots\dots\dots 4$$

The protein efficiency ratio (PER, g/g) was calculated by dividing mean body weight gain (AWG) by the mean protein consumed (PC) as shown in formula 5.

$$PER \text{ (g/g)} = \text{Body weight gain (g)} / \text{protein consumed} \dots\dots\dots 5$$

Growth efficiency was calculated by dividing BWG (g) by initial weight (g) as shown in formula 6.

$$GE = \text{Body weight gain (g)} / \text{initial weight (g)} \dots\dots\dots 6$$

Slaughter procedure

All the Japanese quails were humanely sacrificed to evaluate the size of internal organs and carcass characteristics following a 6-week feeding trial. The night before slaughter, feed was withdrawn to clear the digestive system. The following morning quails were weighed (pre-slaughter weight) to determine the slaughter weight (SLW). Japanese quails were then transported to the BUAN slaughterhouse where they were electrically stunned and bled for 5 minutes by cutting the jugular vein following a procedure by Mnisi et al. (2021). Quails were then soaked for 2 minutes in the heated water (60 °C) for easy de-feathering as described by Narinc et al. (2014). Quail carcasses were then weighed, and all the internal organs were removed using a sharp knife. The carcasses were then individually weighed as hot carcass weight (HCW) and stored in the chiller at -4 °C for 24 hours. The entire internal organs (liver, gizzards, proventriculus, spleen, abdominal fat, heart, small and large intestines) were then separated and weighed. After 24 hours, the carcasses were weighed again to determine cold carcass weight (CCW), and the external parts (drumstick, thighs, breast muscle, wings, and back) were dissected (Mnisi et al., 2021) and weighed individually to determine their weight. Wings were detached by cutting at the humeroscapular joint, cutting it through the rib head to the shoulder girdle, and pulling the vertebrae out intact (Alikwe et al., 2010).

Determination of carcass yield, cuts, and internal organ weights

Internal and external organs were obtained and expressed as the fraction of HCW as shown in equation 7.

$$\text{Internal and external organs} = \frac{\text{Weight (External and Internal organ)(g)}}{\text{HCW (g)}} \times 100 \dots\dots\dots 7$$

Hot carcass yield was calculated by dividing HCW by pre-slaughter weight following equation 8.

$$\text{Hot carcass yield percentage} = \frac{\text{Hot carcass weight (g)}}{\text{Live weight (g)}} \times 100 \dots\dots\dots 8$$

The dressing out percentage was calculated by dividing HCW by pre-slaughter weight as shown in equation 9.

$$\text{Dressing out (\%)} = \frac{\text{Carcass weight (g)}}{\text{Body weight(g)}} \times 100 \dots\dots\dots 9$$

The breast muscle percentage was calculated by dividing breast muscle weight by HCW as shown in equation 10.

$$\text{Breast muscle (\%)} = \frac{\text{Breast muscle (g)}}{\text{Hot carcass weight(g)}} \times 100 \dots\dots\dots 10$$

The drumstick percentage was calculated by dividing drumstick weight by HCW as shown in equation 11.

$$\text{Dumstick (\%)} = \frac{\text{Drumstick (g)}}{\text{Hot carcass weight(g)}} \times 100 \dots\dots\dots 11$$

Data analyses

The growth performance data were checked for interaction with time; where the interaction existed, data were analysed using repeated measures shown in model 1 and if no interaction existed, data were analysed using the general linear model procedure (GLM) of SAS (2013) version 9.4 for a completely randomized experimental design with pen as the experimental unit shown in model 2.

$$Y_{ijk} = \mu + \tau_i + w_j + w\tau_{ij} + e_{ijk} \dots\dots\dots \text{model 1}$$

Where, Y_{ijk} = ijk^{th} response variable; μ = General mean; τ_i = i^{th} treatment effect (tomato waste); w_j = j^{th} week effect; $(w\tau)_{ij}$ = ij^{th} interaction between week and diet effect; e_{ij} = random variation $\sim N(0, \sigma_e^2)$.

$$Y_{ij} = \mu + \tau_i + e_{ij} \dots\dots\dots \text{model 2.}$$

Where, Y_{ij} = ij^{th} response variable; μ = General mean; τ_i = i^{th} treatment effect (tomato waste); e_{ij} = random variation $\sim N(0, \sigma_e^2)$.

The Tukey's Studentized (HSD) Range Test in the SAS (2013) was used to separate the means when the Analysis of Variance indicated the existence of a significant difference between the treatment means. Furthermore, all data were evaluated for linear and quadratic effects using polynomial contrasts. Response procedures for surface regression analysis (Proc RSREG; SAS 9.4, 2013) were applied to describe responses of parameters to graded levels of tomato waste in the diets fed to Japanese quails following the quadratic model: $y = ax^2 + bx + c$, where y = response variables, a and b are the coefficients of the quadratic equation; c is the intercept; x is dietary tomato waste level (%). The significance level was declared at $P < 0.05$.

RESULTS

Growth performance

Repeated measures analyses showed overall significant week × diet interaction effects on FI ($P = 0.03$), weight gain ($P = 0.0006$), FCR ($P = 0.002$), protein efficiency ratio (PER) ($P = 0.0001$), and growth efficiency (GE) ($P = 0.0001$) but not on protein consumed ($P = 0.10$). Diet significantly affected quails' FI at weeks 1, 2, 3, and 6 only (Table 3). In weeks 1, 3, and 6 quails fed on TW0 and TW2 diets had significantly higher FI than those fed diet TW6. Quails fed on TW4 diets had significantly similar FI to those fed on TW0, TW2, and TW6 diets. In week 2, quails fed on TW0 and TW2 diets had significantly higher FI than those fed on TW4 and TW6 diets which were statistically similar. In weeks 4 ($P = 0.20$) and 5 ($P = 0.08$), diets did not affect FI. The regression analysis in Table 4 revealed that FI decreased linearly in weeks 1, 4, and 6 as tomato waste inclusion levels increased. In week 2, FI increased linearly with tomato waste. However, decreased quadratic trends in FI were observed in weeks 3 and 5 as tomato waste levels increased in the diets.

The results in Table 3 showed significant dietary effects in weeks 2, 3, and 4 but diets did not significantly affect PER in weeks 1, 5, and 6. In week 2, quails fed on TW0 and TW2 diets had significantly higher PER compared to those fed TW4 and TW6 diets, which were statistically similar. In weeks 3 and 4, quails fed on TW0 diet had significantly higher PER than those fed TW2, TW4, and TW6 diets, which did not differ significantly. Regression analysis revealed that PER linearly decreased in weeks 1 and 6 (Table 4). However, there was a linear increase in week 2 in PER as tomato waste levels increased. Increasing quadratic effects were observed in weeks 3 and 4 with increased dietary tomato waste levels. In week 5, neither linear nor quadratic trends were observed in PER as dietary tomato waste increased.

The results revealed no significant dietary effects in weeks 5 and 6 in growth efficiency (GE) (Table 3). However, significant differences were recorded in weeks 1, 2, 3, and 4. At weeks 1 and 2, quails fed on TW0 and TW2 diets had significantly higher GE followed by those fed TW4 diet and lastly, those fed on TW6 diet. In weeks 3 and 4, quails fed on TW0 diet had higher GE followed by those on TW2 and TW4 diets which were statistically similar and lastly, those fed on TW6 diet. Regression analysis results in Table 4 revealed that PER linearly decreased in week 1 and linearly increased in week 5 with increased dietary levels of tomato waste. However, a negative trend was observed on GE in response to increases in dietary levels of tomato waste at 2, 3, 4, and 6 weeks of age.

Table 3 - Weekly feed intake, protein efficiency ratio, and growth efficiency of Japanese quails fed graded levels of tomato waste as partial replacement of soybean meal

Parameter	Diets ¹				SEM ²	P value	Significance ³	
	TW0	TW2	TW4	TW6			Linear	Quadratic
Feed intake (g/quail)								
Week 1	6.56 ^a	6.54 ^a	5.96 ^{ab}	5.87 ^b	0.149	0.0091	*	NS
Week 2	8.78 ^a	8.67 ^a	7.29 ^b	7.19 ^b	0.140	< 0.0001	*	NS
Week 3	12.80 ^a	12.76 ^a	12.27 ^{ab}	12.25 ^b	0.090	< 0.0001	*	*
Week 4	17.69	17.51	17.16	16.95	0.250	0.1978	*	NS
Week 5	28.08	27.96	27.56	27.03	0.280	0.0847	*	*
Week 6	33.51 ^a	33.13 ^a	32.78 ^{ab}	31.95 ^b	0.220	0.0018	*	NS
Protein efficiency ratio								
Week 1	0.99	0.92	0.90	0.91	0.020	0.0831	*	NS
Week 2	2.61 ^a	2.58 ^a	2.26 ^b	2.24 ^b	0.050	0.0001	*	NS
Week 3	2.76 ^a	2.45 ^b	2.43 ^b	2.38 ^b	0.030	< 0.0001	*	*
Week 4	1.51 ^a	1.28 ^b	1.29 ^b	1.28 ^b	0.020	< 0.0001	*	*
Week 5	0.81	0.79	0.78	0.80	0.010	0.4425	NS	NS
Week 6	0.67	0.65	0.63	0.63	0.010	0.0579	*	NS
Growth efficiency								
Week 1	1.33 ^a	1.32 ^a	1.14 ^b	1.10 ^c	0.005	< 0.0001	*	NS
Week 2	1.87 ^a	1.86 ^a	1.80 ^b	1.74 ^c	0.010	< 0.0001	*	*
Week 3	1.00 ^a	0.81 ^b	0.80 ^b	0.78 ^c	0.005	< 0.0001	NS	*
Week 4	0.77 ^a	0.47 ^b	0.46 ^b	0.44 ^c	0.010	< 0.0001	NS	*
Week 5	0.32	0.32	0.32	0.27	0.020	0.1034	*	NS
Week 6	0.24	0.24	0.24	0.23	0.010	0.0008	*	*

^{a,b,c}In row, means with common superscripts do not differ significantly. ¹Diets: TW0 = Diet with no tomato waste; TW2 = Diet with 2% of soybean meal replaced with tomato waste; TW4 = Diet with 4% of soybean meal replaced with tomato waste; TW6 = Diet with 6% of soybean meal replaced with tomato waste. ²SEM = standard error of mean; ³Significance: * = Significant difference ($P < 0.05$); NS = Not significant.

Table 4 - The linear and quadratic trends on weekly feed intake (g/bird), weekly protein efficiency ratio, and weekly growth efficiency of Japanese quails fed graded levels of tomato waste as partial replacement of soybean meal

Parameter	Equation	P value	R ²
Feed Intake (g/quall)			
Week 1	$Y = 6.5 (\pm 0.7) - 0.8 (\pm 0.6) X$	0.0001	0.69
Week 2	$Y = 8.8 (\pm 1.2) + 0.6 (\pm 1.0) X$	0.0002	0.70
Week 3	$Y = 12.5 (\pm 0.7) + 0.5 (\pm 0.571.1) X - 0.2 (\pm 0.09) X^2$	0.0300	0.70
Week 4	$Y = 17.7 (\pm 0.5) - 1.0 (\pm 0.4) X$	< 0.0001	0.82
Week 5	$Y = 28.0 (\pm 0.4) - 0.1 (\pm 0.3) X - 0.2 (\pm 0.1) X^2$	0.0020	0.92
Week 6	$Y = 33.5 (\pm 1.2) - 1.4 (\pm 1.0) X$	0.0030	0.50
Protein efficiency ratio			
Week 1	$Y = 0.99 (\pm 0.02) - 0.05 (\pm 0.02) X$	0.0300	0.41
Week 2	$Y = 2.2 (\pm 0.1) + 0.05 (\pm 0.05) X$	0.0020	0.67
Week 3	$Y = 2.7 (\pm 0.04) - 0.2 (\pm 0.03) X + 0.02 (\pm 0.005) X^2$	< 0.0001	0.80
Week 4	$Y = 1.5 (\pm 0.02) - 0.1 (\pm 0.02) X + 0.01 (\pm 0.003) X^2$	0.0010	0.81
Week 6	$Y = 0.7 (\pm 0.01) - 0.02 (\pm 0.01) X$	0.0100	0.45
Growth efficiency			
Week 1	$Y = 1.3 (\pm 0.03) - 0.06 (\pm 0.02) X$	< 0.0001	0.76
Week 2	$Y = 1.7 (\pm 0.01) + 0.05 (\pm 0.01) X - 0.05 (\pm 0.002) X^2$	0.0100	0.85
Week 3	$Y = 0.81 (\pm 0.04) + 0.08 (\pm 0.03) X - 0.02 (\pm 0.005) X^2$	0.0100	0.50
Week 4	$Y = 0.5 (\pm 0.1) + 0.1 (\pm 0.04) X - 0.02 (\pm 0.01) X^2$	0.0100	0.50
Week 5	$Y = 0.3 (\pm 0.02) + 0.03 (\pm 0.01) X$	0.0500	0.36
Week 6	$Y = 0.2 (\pm 0.01) + 0.03 (\pm 0.01) X - 0.01 (\pm 0.001) X^2$	0.0002	0.73

The results showed that diet affected average weight gain at weeks 1, 2, 4, and 6 only but did not affect the weight gain of quails at weeks 3 and 5 (Figure 1). At week 1, quails fed on TW0 diet had significantly higher WG than those fed on TW2, TW4, and TW6 diets. In week 2, quails fed on TW0 and TW2 diets had significantly higher WG than those fed on TW4 and TW6 diets. Significantly higher WG was recorded in week 4 in quails fed on TW0 and TW2 diets than those fed on TW6 diet. However, quails on TW4 diet had statistically similar WG to those fed on TW0, TW2, and TW6 diets. In week 6, quails fed on TW0 diet had significantly higher WG compared to those fed on TW2 diet followed by TW4 and lastly TW6. At 3 and 5 weeks of age, diet did not significantly affect WG of quails.

The results showed that there were significant dietary effects in week 2 only (Figure 2). However, there were no dietary effects in FCR at weeks 1, 3, 4, 5, and 6. In week 2, quails fed on TW0 and TW2 diets had significantly lower FCR compared to those fed on TW4 and TW6 diets which were statistically similar. The regression analysis results revealed that FCR decreased linearly from week 1 to 2 and then increased linearly from week 2 to 6.

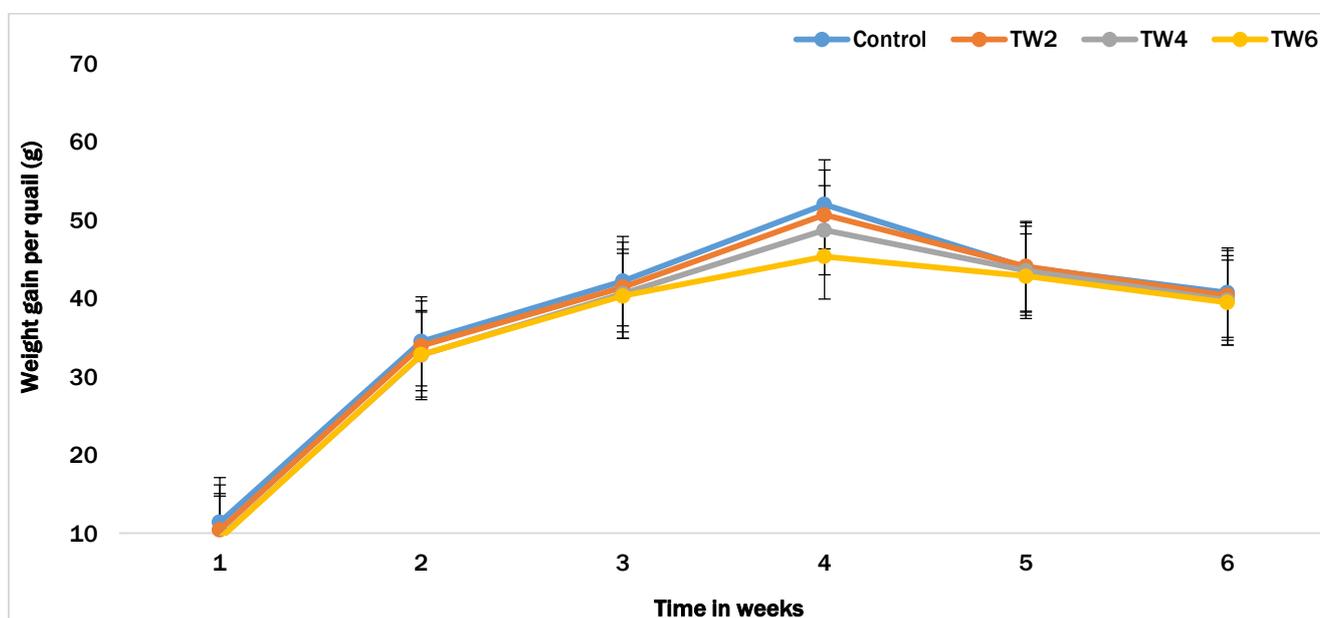


Figure 1 - Weekly weight gain of Japanese quails fed graded levels of tomato waste as a partial replacement for soybean meal.

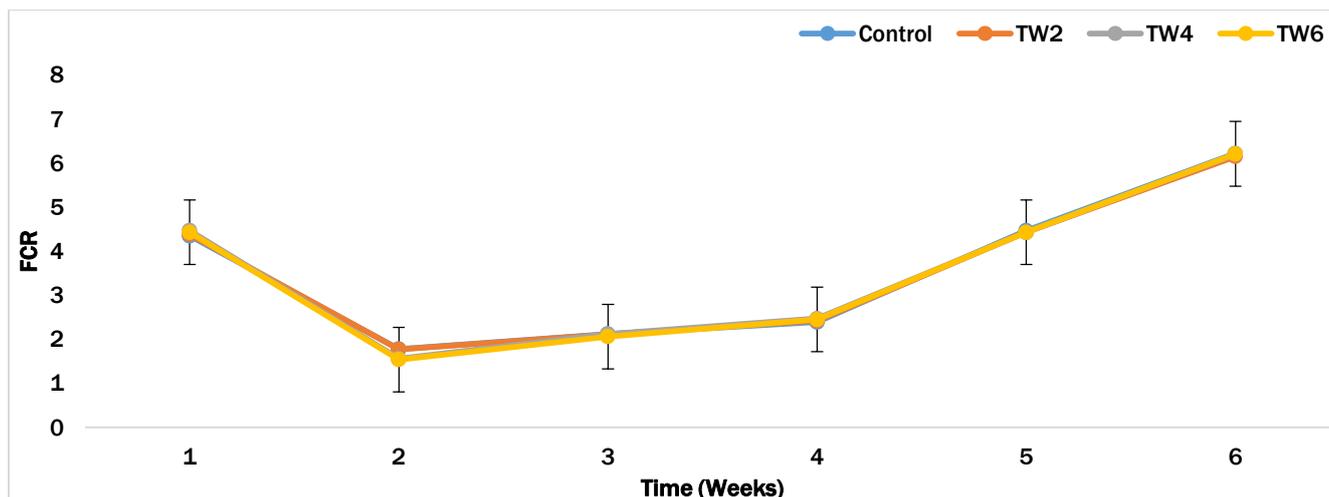


Figure 2 - Weekly FCR of Japanese quails fed graded levels of tomato waste as partial replacement of SM.

Carcass characteristics

Diets affected (SLW), (HCW), and (CDW) but did not affect (CL) and dressed weight percentage (Table 5). Quails fed on the TW0 diet had significantly higher SLW followed by those fed on the TW2 diet, and lastly, those fed on the TW4 and TW6 diets, which were statistically similar. Similarly, quails fed on the TW0 diet had significantly higher HCW and CDW than those fed on the TW2, TW4, and TW6 diets. However, quails fed on the TW4 diet had significantly similar HCW and CDW compared to those fed on the TW2 and TW6 diets. Regression analysis showed positive trends in SLW of Japanese quails with tomato waste inclusion levels (Table 6). Linear decreases were recorded in SLW, HCW, and CDW with increased tomato waste inclusion levels. No significant linear and quadratic trends for dressed percentage were detected.

Table 5 - Effects of feeding graded levels of tomato waste (g, unless otherwise stated) as partial replacement of soybean meal on carcass characteristics of six weeks old Japanese quails

Parameter ¹	Diets ²				SEM ³	P value	Significance ⁴	
	TW0	TW2	TW4	TW6			Linear	Quadratic
SLW	220.89 ^a	207.13 ^b	200.36 ^c	195.07 ^c	1.49	< 0.0001	*	*
HCW	165.60 ^a	156.44 ^b	151.55 ^{bc}	147.06 ^c	1.25	< 0.0001	*	NS
CDW	165.45 ^a	156.31 ^b	151.43 ^{bc}	146.95 ^c	1.24	< 0.0001	*	NS
CL	0.15	0.13	0.12	0.11	0.02	0.50	NS	NS
Dressed (%)	75.00	75.5	75.67	75.17	0.31	0.43	NS	NS

¹Parameter: SW= Slaughter weight; HCW = Hot carcass weight; CDW = Cold dressed weight; CL = Chilling loss. ²Diets: TW0 = Diet with no tomato waste; TW2 = Diet with 2% of soybean meal replaced with tomato waste; TW4 = Diet with 4% of soybean meal replaced with tomato waste; TW6 = Diet with 6% of soybean meal replaced with tomato waste. ³SEM= Standard error of the mean. ⁴Significance: NS = Not significant; * = Significant; ^{abc}Within a row, different superscripts denote significant differences (P < 0.05) between treatments.

Table 6 - Linear and quadratic trends of carcass characteristics of Japanese quails fed graded levels of tomato waste as partial replacement of soybean meal

Parameter	Equation	P value	R ²
SLW	Y = 220.5 (± 1.6) - 7.3(± 1.3) X + 0.5 (± 0.2) X ²	0.02	0.87
HCW	Y = 165.2 (± 1.4) - 4.7(± 1.1) X	< 0.0001	0.83
CDW	Y = 164.9 (± 1.4) - 4.7(± 1.1) X	< 0.0001	0.83

¹Parameter: LW= Live weight (g); HCW = Hot carcass weight (g); CDW = Cold dressed weight (g); CL = Chilling loss (g).

Diet significantly affected the quail back only but did not affect the drumstick, wings, thighs, and breast (Table 7). Quails fed on the TW0 diet had significantly heavier backs than those fed on the TW4 and TW6 diets. Quails fed on diet TW2 had significantly similar back weights to those fed on TW0 and TW4 diets. Quails fed on the TW4 diet had a lighter back but were statistically similar to quails fed on diets TW2 and TW6. The regression analysis revealed linear decreases for the drumstick [y = 12.9 (± 0.4) - 0.1(± 0.3) X; R² = 0.29; P = 0.01] as dietary levels of tomato waste increased. Neither linear nor quadratic effects were observed on wings, thighs, back, and breast.

Diet significantly affected the liver and heart weights only but did not affect the weights of gizzards, proventriculus, spleen, and fat (Table 8). Quails fed on TW0 diet had significantly heavier liver followed by those fed on TW2 diet and lastly Quails fed on TW4 and TW6 diets had statistically similar liver and hear weights. Quails fed on TW0 diet had heavier hearts than those fed TW4 and TW6 diets. However, quails fed on TW2 diet had significantly similar heart weights with those fed on TW0, TW4, and TW6 diets. The regression results showed that there were linear decreases for liver with incremental levels of tomato waste [$y = 1.6 (\pm 0.1) - 0.05 (\pm 0.3) X$; $R^2 = 0.31$; $P = 0.03$]. However, proventriculus increased linearly [$y = 0.3 (\pm 0.02) + 0.02 (\pm 0.02) X$; $R^2 = 0.22$; $P = 0.04$] with tomato waste incremental levels. No significant linear and quadratic trends were observed on gizzards, spleen, fat and heart.

Diet significantly affected the weight of the small intestines but did not affect the length of the small intestines, and length and weight of the large intestines (Table 9). Quails fed on the TW0 (4.55 %) diet had heavier small intestines than those fed on the TW4 and TW6 diets. However, the quails fed on TW2 diet had significantly similar small intestines weights with those on TW0, TW4, and TW6 diets. The regression analysis results showed a quadratic decrease on small intestines weight with tomato waste levels [$y = 3.2 (\pm 0.2) - 0.3 (\pm 0.1) X + 0.1 (\pm 0.02) X^2$; $R^2 = 0.28$; $P = 0.01$]. No significant linear and quadratic trends were observed on small and, large intestines length, and large intestines weights.

Table 7 - Effects of feeding graded levels of tomato waste (%HCW), as partial replacement of soybean meal on external organs of six weeks old Japanese quails

Parameter ¹	Diets ²					P value	Significance ⁴	
	TW0	TW2	TW4	TW6	SEM ³		Linear	Quadratic
Drumstick	11.06	11.65	11.19	10.78	0.29	0.23	NS	NS
Wings	8.07	8.06	8.06	8.07	0.01	0.72	NS	NS
Thighs	21.00	20.52	20.28	20.15	0.53	0.68	NS	NS
Back	18.65 ^a	18.12 ^{ab}	17.15 ^{bc}	17.25 ^c	0.31	0.01	*	NS
Breast	37.97	37.30	36.97	36.48	0.67	0.29	NS	NS

¹Diets: TW0 = Diet with no tomato waste; TW2 = Diet with 2% of soybean meal replaced with tomato waste; TW4 = Diet with 4% of soybean meal replaced with tomato waste; TW6 = Diet with 6% of soybean meal replaced with tomato waste. ²SEM= Standard error of the mean. ³Significance: NS = Not significant; * = Significant; ^{abc} Within a row, different superscripts denote significant differences ($P < 0.05$) between treatments.

Table 8 - Effects of feeding graded levels of tomato waste (%HCW), as partial replacement of soybean meal on internal organs of six weeks old Japanese quails

Parameter ¹	Diets ²					P value	Significance ⁴	
	TW0	TW2	TW4	TW6	SEM ³		Linear	Quadratic
Liver	3.62 ^a	3.39 ^b	3.19 ^c	3.12 ^c	0.04	< 0.0001	*	NS
Gizzards	2.03	2.08	2.05	2.04	0.02	0.13	NS	NS
Proventriculus	0.40	0.40	0.43	0.39	0.02	0.34	NS	NS
Spleen	0.18	0.08	0.08	0.08	0.05	0.47	NS	NS
Abdominal fat	0.07	0.19	0.08	0.08	0.06	0.42	NS	NS
Heart	0.83 ^a	0.80 ^{ab}	0.75 ^b	0.76 ^b	0.01	0.01	*	NS

¹Diets: TW0 = Diet with no tomato waste; TW2 = Diet with 2% of soybean meal replaced with tomato waste; TW4 = Diet with 4% of soybean meal replaced with tomato waste; TW6 = Diet with 6% of soybean meal replaced with tomato waste. ²SEM= Standard error of the mean. ³Significance: NS = Not significant; * = Significant; ^{abc} Within a row, different superscripts denote significant differences ($P < 0.05$) between treatments.

Table 9 - Effects of feeding graded levels of tomato waste (% , unless otherwise stated) as partial replacement of soybean meal on intestines of six weeks old Japanese quails

Parameter ¹	Diets ²					P value	Significance ⁴	
	TW0	TW2	TW4	TW6	SEM ³		Linear	Quadratic
Small intestines (cm)	60.00	60.00	60.00	59.17	0.92	0.89	NS	NS
Small intestines (%)	4.55 ^a	4.39 ^{ab}	3.88 ^b	3.85 ^b	0.18	0.03	NS	*
Large intestines (cm)	12.58	12.22	11.88	11.58	0.40	0.36	NS	NS
Large intestines (%)	0.77	0.77	0.75	0.75	0.05	0.94	NS	NS

¹Diets: TW0 = Diet with no tomato waste; TW2 = Diet with 2% of soybean meal replaced with tomato waste; TW4 = Diet with 4% of soybean meal replaced with tomato waste; TW6 = Diet with 6% of soybean meal replaced with tomato waste. ²SEM= Standard error of the mean. ³Significance: NS = Not significant; * = Significant; ^{a,b,c} Within a row, different superscripts denote significant differences ($P < 0.05$) between treatments.

DISCUSSION

Growth performance

Repeated measures analysis showed significant week x diet interaction effects on weekly FI, WG, FCR, PER, and GE, demonstrating that tomato waste inclusion influenced quail growth performance over time. The significantly lower FI in quails fed TW6 diets in weeks 1, 3, and 6 suggest that the higher inclusion levels reduced FI. Common organoleptic characteristics associated with tannin compounds are astringency and bitterness which reduce voluntary FI (Choi and Kim, 2020) which could be linked with FI reductions. Similarly, Tabeidian et al. (2011) reported that growth performance was lowered even at modest inclusion levels of tomato waste at 3% in the starter and 9% in the finisher phase of broiler chicks. In contrast, Shehata et al. (2018) and Muhammad et al. (2023) reported increased feed consumption when tomato pomace-containing diets were fed to laying Japanese quails at 2.5 and 5% inclusion levels. This discrepancy could be due to the different species (broilers vs. Japanese quails) used. It is noted that different poultry species act differently when exposed to the same diet (Mnisi and Mlambo, 2018). Quails fed on the TW0 diet had similar feed intake to TW4, TW2, and TW6 suggesting that tomato waste at 6% inclusion did not affect the voluntary FI of quails, suggesting that the diet (tomato waste) can be used up to 6% inclusion level. This is in line with the findings of Mohammed et al. (2021) who reported that supplementing tomato pomace at 6% did not affect FI of broiler chicks. Contrary to our expectations, in week 2, quails fed on TW0 and TW2 diets had significantly higher FI than those fed on other diets, suggesting that tomato waste can be used at 2% inclusion without affecting the palatability of the diets. Lower FI of quails fed TW4 and TW6 diets could be due to tannins that exert a bitter taste resulting in decreased feed consumption (Hassan et al., 2020). The physicochemical characteristics of the entire diet are greatly affected by a little alteration in an ingredient during diet formation.

In weeks 4 and 5, diet did not affect FI suggesting that tomato waste inclusion rates up to 6% can produce similar results as the control diet. This could be linked with the maturity of the GIT of Japanese quails which efficiently handled the fiber contents of the diets. These results agree with Gungor et al. (2024) who reported that the supplementation of dried tomato pomace to broiler diets up to 10% had no significant effect on FI. Similarly, Jouzi et al. (2015) found no significant difference between the diet groups supplemented with 5% dried tomato pomace and the control group of Japanese quails.

Quails fed on TW0 and TW2 diets gained more weight in weeks 1, 2, 4, and 6, indicating that they were able to use the nutrients in the diets implying that the ANFs such as tannins, pectins, and insoluble fiber were negligible. These ANFs temper with feed utilization (Szabo et al., 2019). The bioactive components found in tomato waste include lycopene and β -carotene, which have substantial antioxidant capacity (Szabo et al., 2019). Lycopene is a powerful antioxidant that can protect muscle cells from the damaging effects of reactive oxygen species and biomolecule destruction (Mezban et al., 2019). Our results agree with Sahin et al. (2008) who reported that a 5% inclusion level of tomato waste in broiler diets improved the body weight of broiler chickens.

At weeks 3 and 5 diet did not significantly affect the weight gain of quails, suggesting that tomato waste could be incorporated into the diet without affecting Japanese quails' feed utilization. This was due to statistically similar FI, nutrient intake, and feed utilisation. In line with our results, Nikolakakis et al. (2004) reported no significant differences in body weight and BWG on growing quails fed on diets containing tomato pulp at 5 and 10% levels. Contrary to this finding, Alagawany et al. (2021) noted a significant improvement in body weight and BWG when quails were fed on 6% sundried tomato pomace at 3 to 5 weeks of age compared to those fed on the control diet. The current result implies that sun-dried tomato waste can replace SM by 6% in the quail diet without detrimentally affecting weight gain.

There were no significant dietary effects on FCR at weeks 1, 3, 4, 5, and 6 implying that tomato waste can be used in quail diets as partial replacement of SM. In agreement with our results, Jouzi et al. (2015) obtained no significant effect on FCR in growing Japanese quails fed on tomato powder at 2, 4, 6, and 8% inclusion levels. Similar to the weight gain, at week 2, quails fed on TW0 and TW2 diets in this study had comparable FCR suggesting that the quails utilized the feed regardless of the fiber content in the diet. The quails fed on TW4 and TW6 had improved FCR probably due to the high fiber content that is responsible for the improvements in villus height and overall epithelial cell arrangement which increases nutrient absorption and hence better FCR. In addition, high fiber content increases the retention time of the digesta along the GIT, causing optimum digestion and absorption of nutrients for anabolic purposes.

Fiber is known to be involved in the modification of the intestinal length, villus height, crypt depth, as well as, the passage rate and size through different segments of the intestines (Rezaei et al., 2018; Tejada and Kim, 2021). However, as fiber increased the FCR began to increase from weeks 2 to 6 due to the presence of tannin and pectins that cause a reduction in nutrient absorption due to the formation of insoluble complexes with proteins leading to poorer FCR observed. It is known that monogastric animals do not produce the cellulolytic enzymes required to break down the fiber but utilise the microbial enzymes in their caeca, though less efficient in degrading fiber (Jha and Mishra, 2021). Additionally, fiber has been considered a diluent of the diet and is known to increase the passage rate of the gastrointestinal tract owing to the reduction of nutritional utilization (Mateos et al., 2012). Hosseini-Vashan et al. (2016)

found that supplementing 5% tomato pomace increased broiler FCR during heat stress. In contrast with the present results, [Lira et al. \(2010\)](#) reported that using more than 20% tomato pomace in the diet of broiler chickens from 1 to 28 days may decrease FCR. The current results indicated that tomato waste inclusions possibly interfered with the use of nutrients in the rations at higher inclusion levels of tomato waste. The current results suggest that the inclusion of tomato waste at 4 and 6% compromised FCR.

The comparison between the average body weight and the amount of protein in the diet is known as the protein efficiency ratio (PER) which is influenced by the productivity of the animals ([Ratriyanto et al., 2017](#)). The rise in PER values in the present study indicates that the birds were able to use the protein they consume more effectively. In this study, quails fed on TW0 and TW2 diets in week 2 had similar PER suggesting that tomato waste can be used at the inclusion of 2% without affecting the PER. There is very limited literature on the PER of poultry-fed tomato waste. There was a significant reduction in PER at weeks 3 and 4 in quails fed tomato-based diet, suggesting that the inclusion of tomato waste in diets above 2% hindered protein utilization. This implies that as the tomato waste increases, the tannins and pectins that bind protein increase resulting in protein utilisation being hindered. Also, the feed was slowly digested since the quails do not produce the cellulolytic enzymes required to quickly break down fiber. This implies that the inclusion of tomato waste as a partial replacement of SM compromised PER. No significant dietary effects were noted at 1, 5, and 6 weeks of age across diets, suggesting that protein utilization was similar to the control diet. This implies that tomato waste can be incorporated in the quail diets at 2% without compromising the PER.

Quails fed on TW0 and TW2 diets at 1 and 2 weeks of age had significantly higher GE compared to those fed TW4 and TW6 diets, indicating that tomato waste reduced the GE when included in the quail's diets at higher levels. This reduction in GE could be due to the presence of pectins, tannins and fiber which increase with increasing tomato waste inclusion levels. According to [Brenes et al. \(2016\)](#) and [Mnisi et al. \(2022\)](#), anti-nutrients inhibit protein utilisation and nutrient digestibility owing to depressed growth. Furthermore, it is speculated that the capacity of the gut is very limiting in young quails; hence their inability to produce enough fibrinolytic enzymes that help in the digestion of fiber. Our results are in line with [Cavalcante et al. \(2007\)](#) who reported the negative effect of tomato waste in the early stages of quail life due to great sensitivity of young chicks to ingestion of diets with high fiber content. Fiber is a naturally occurring plant component associated with physiological, structural, and functional changes in the GIT ([Deehan et al., 2022](#)). At weeks 3 and 4, quails fed on tomato-based diets had reduced GE suggesting that tomato waste impaired growth when included at higher levels i.e. above 2% inclusion level. Increasing the inclusion level of tomato waste in the diet led to an increase in tannin contents, which interfered with protein utilization, thus suppressing growth. Therefore, the inclusion of tomato waste as a partial replacement for SM compromised GE. In agreement with our results, [Tabeidian et al. \(2011\)](#) found that growth performance in broilers was lowered even at a modest inclusion rate of 3% in the starter phase and 9% in the finisher phase.

Carcass characteristics

Among the variables that are known to affect carcass features in birds include age, sex, diet, genetics, and conditions of slaughter ([Young et al., 2001](#)). In this study, the results showed that diet had an impact on HCW, CDW, and SLW, and TW6 diets showed notably greater HCW and CDW in Japanese quails. This was due to reduced FI in tomato-based diets. In this study, the inclusion of tomato waste in diets linearly decreased carcass yield, HCW, CDW, CL, and weight of drumsticks, thigh, wing, and breast. This observation may be related to the lower FI observed in quails fed on diet containing tomato waste, which led to inadequate nutrient uptake for muscle growth and ineffective weight gains. This suggests that feeding tomato waste to quail could have a negative impact on the viability and profitability of quail businesses at high inclusion levels. These results agree with [Jouzi et al. \(2015\)](#) and [Lira et al. \(2010\)](#) who reported a linear decrease in carcass yield, wings, breast, and drumstick when quails were fed on diets containing 8% of tomato waste. In contrast to our results, [Yitbarek \(2013\)](#) reported that broilers fed a diet containing 5% tomato pomace had a higher carcass yield than other treatments. However, broiler chicks fed a diet containing 15% dried tomato pomace showed a lack of significant difference on carcass characteristics suggesting that tomato pomace can be used at 15% without detrimental effect on carcass characteristics.

The gastrointestinal morphology of poultry is related to variations in dietary fiber content as an adaptation mechanism to make use of the high levels of fiber ([Jha et al., 2019](#)). The lower liver weights on tomato-based diets in the present study could have been triggered by a negligible concentration of secondary plant compounds such as pectins and tannins in tomato waste, which require detoxification by the liver when available in the diets. This lower liver weight is linked with less ANFs in the diet. A previous study of [Leke et al. \(2018\)](#) showed that the heart and gizzard weights were not significantly affected by dietary tomato waste at 0, 3, 6, 9, and 12% inclusion levels. In contrast with our results, [Mateos et al. \(2012\)](#) observed that birds fed fibrous diets had enlarged gastrointestinal organs. Nevertheless, despite the presence of crude fiber in the tomato waste-containing diets in this study, examination of the visceral organs (fat, spleen, proventriculus, and gizzards) revealed no significant effects. It is likely that dietary tomato waste levels as high as 6% were not enough to cause physio-anatomical changes in the fat, spleen, proventriculus, or gizzards.

- AOAC (2005). Official Methods of Analysis of the Association of Official's Analytical Chemists, 18th ed.; Association of Official Analytical Chemists: Arlington, VA, USA. method number, 920.39, 924.05, and 984.13. <https://www.cabidigitallibrary.org/doi/full/10.5555/19951414840>
- Aragaw S, Urge M, Zeryehun T and Lemma F (2021). Effect of dried tomato waste meal on growth performance and carcass characteristics of Cobb 500 broiler chicks. *Livestock Research for Rural Development*, 33(5): 65. <http://www.lrrd.org/lrrd33/5/3365fethu.html>
- Ayaşan T (2013). Effects of dietary inclusion of protexin (probiotic) on hatchability of Japanese quails. *Indian Journal of Animal Science*, 83(1): 78-81. <https://epubs.icar.org.in/index.php/IJAnS/article/download/26451/12103/58179>
- Bonos EM, Christaki EV and Florou-Paneri PC (2010). Performance and carcass characteristics of Japanese quail as affected by sex or mannan oligosaccharides and calcium propionate. *South African Journal of Animal Science*, 40(3): 173-184. <https://doi.org/10.4314/sajas.v40i3.2>
- Brenes A, Viveros A, Chamorro S and Arija I (2016). Use of polyphenol rich grape by-products in monogastric nutrition. A review. *Animal Feed Science and Technology*, 211: 1-17. <https://doi.org/10.1016/j.anifeedsci.2015.09.016>
- Cavalcante SB, Fuentes MFF, Freitas ER, Espindola G and Brag C (2007). Effect of including coconut bran in diets for chickens cut. *Revista Ciência Agronômica*, 38(3): 297-303. [Article link](#)
- Choi J and Kim WK (2020). Dietary application of tannins as a potential mitigation strategy for current challenges in poultry production: A Review. *Animals (Basel)*, 10(12): 2389. <https://doi.org/10.3390/ani10122389>
- Climate-Data.org (2022). Climate data for cities worldwide. <http://en.climate-data.org>
- Dalle Zotte A, and Cullere M (2024). Rabbit and quail: Little known but valuable meat sources. *Czech Journal of Animal Science*, 69(2):39-47. <https://www.agriculturejournals.cz/pdfs/cjs/2024/02/02.pdf>
- Deehan EC, Zhang Z, Riva A, Amert AM, Perez-Muñoz ME, Nguyen NK, et al. (2022). Elucidating the role of the gut microbiota in the physiological effects of dietary fiber. *Microbiome*, 10: 77. <https://doi.org/10.1186/s40168-022-01248-5>
- Dou Z, Dierenfeld ES, Wang X, Chen X, and Shurson GC (2024). A critical analysis of challenges and opportunities for upcycling food waste to animal feed to reduce climate and resource burdens. *Resources, Conservation and Recycling*, 203:107418. <https://doi.org/10.1016/j.resconrec.2024.107418>
- Google earth pro software (2022). Google earth pro-Inc. <https://earth.google.com/web/>
- Gungor E, Altop A and Erener G (2024). Effect of fermented tomato pomace on the growth performance, antioxidant capacity, and intestinal microflora in broiler chickens. *Animal Science Journal*, 95(1): e13885. <https://doi.org/10.1111/asj.13885>
- Hassan ZM, Manyelo TG, Salaleli L and Mabelebele M (2020). The effects of tannins in monogastric animals with special reference to alternative feed ingredients. *Molecules*, 25: 4680. <https://doi.org/10.3390/molecules25204680>
- Hemid AEA, Abd El-Gawad AH and El-Wardany I (2010). Alleviating effect of some environmental stress factors on productive performance in Japanese quail 2. Laying performance. *World Journal of Agricultural Sciences*, 6(5): 517-524. [http://www.idosi.org/wjas/wjas6\(5\)/8.pdf](http://www.idosi.org/wjas/wjas6(5)/8.pdf)
- Hosseini-Vashan SJ, Golian A and Yaghobfar A (2016). Growth, immune, antioxidant, and bone responses of heat stress-exposed broilers fed diets supplemented with tomato pomace. *International Journal of Biometeorology*, 60(8): 1183-92. <https://doi.org/10.1007/s00484-015-1112-9>
- Illgner P and Nel E (2000). The geography of edible insects in sub-Saharan Africa: A study of the mophane caterpillar. *Geography Journal*, 166: 336-51. <https://doi.org/10.1111/j.1475-4959.2000.tb00035.x>
- Jha R and Mishra P (2021). Dietary fiber in poultry nutrition and their effects on nutrient utilization, performance, gut health, and on the environment: A review. *Journal of Animal Science and Biotechnology*, 12: 51. <https://doi.org/10.1186/s40104-021-00576-0>
- Jha R, Fohse JM, Tiwari UP, Li L and Willing BP (2019). Dietary fiber and intestinal health of monogastric animals. *Frontiers of Veterinary Science*, 6: 48. <https://doi.org/10.3389/fvets.2019.00048>
- Jouzi H, Vali N and Pourreza J (2015). The effects of tomato pulp powder supplementation on performance and some blood parameters in Japanese quail (*Coturnix Japonica*). *Journal of Agricultural and Biological Science*, 10(3): 103-107. [Google Scholar](#)
- Leke JR, Mandey JS, Ratulangi F and Najoan M (2018). Effect of tomato (*Solanum lycopersicum* L.) protein on carcass and meat quality of kampung chicken. *Journal of the Indonesian Tropical Animal Agriculture*, 43(1): 35-42. <https://doi.org/10.14710/jitaa.43.1.35-42>
- Lira RC, Rabello C-V, Ludke MDCMM, Ferreira PV, Lana GRQ and Lana SRV (2010). Productive performance of broiler chickens fed tomato waste. *Ras Brasil Zootechnology*, 39(5): 1074–1081. <https://doi.org/10.1590/S1516-35982010000500018>
- Lu S, Chen S, Li H, Paengkoum S, Taethaisong N, Meethip W, Surakhunthod J, Sinpru B, et al. (2022). Sustainable valorisation of tomato pomace (*Lycopersicon esculentum*) in Animal Nutrition: A Review. *Animals (Basel)*, 12(23): 3294. <https://doi.org/10.3390/ani12233294>
- Manyeula F, Mlambo V, Marume U and Sebola NA (2019). Nutrient digestibility, haemo-biochemical parameters and growth performance of an indigenous chicken strain-fed canola meal-containing diets. *Tropical Animal Health and Production*, 51: 2343-2350. <https://doi.org/10.1007/s11250-019-01949-4>
- Mateos GG, Jimenez-Moreno E, Serrano MP and Lazaro RP (2012). Poultry response to high levels of dietary fiber sources varying in physical and chemical characteristics. *Journal of Applied Poultry Research*, 21: 156-174. <https://doi.org/10.3382/japr.2011-00477>
- Mezban A, Kavan BP, Kiani A and Masouri B (2019). Effect of dietary lycopene supplementation on growth performance, blood parameters and antioxidant enzymes status in broiler chickens. *Livestock Research for Rural Development*. 31(12): 12. <https://www.lrrd.org/lrrd31/1/bahma31012.html>
- Mnisi CM and Mlambo V (2018). Canola meal as an alternative dietary protein source in quail (*Coturnix coturnix*) diets - A review. *Acta Agriculturae Scandinavica, Section A - Animal Science*, 68(4): 207-218. <https://doi.org/10.1080/09064702.2019.1679873>
- Mnisi CM, Mhlongo G and Manyeula F (2022). Fruit pomaces as functional ingredients in poultry nutrition: A review. *Frontiers in Animal Science*, 3: 40. <https://doi.org/10.3389/fanim.2022.883988>
- Mnisi CM, Mlambo V, Kumanda C and Crafford A (2021). Effect of graded levels of red grape pomace (*Vitis vinifera* L.) powder on physiological and meat quality responses of Japanese quail. *Acta Agriculturae Scandinavica. Animal Science*, 70(2): 100-106. <https://doi.org/10.1080/09064702.2021.1923796>
- Mohammed LS, Sallam EA, Edris SN, Khalifa OA, Soliman MM and Shehata SF (2021). Growth performance, economic efficiency, meat quality, and gene expression in two broiler breeds fed different levels of tomato pomace. *Veterinary Research Communications*, 45: 381-397. <https://doi.org/10.1007/s11259-021-09819-x>

- Muhammad MD, Chand N, Naz S, Alhidary AA, Shah AA, Khan RU and et Al. (2023). Adaptation to heat stress in broilers using dried tomato pomace and zinc: Effects on growth performance, oxidative stress, intestinal features and humoral immunity. *Pakistan Journal of Zoology*, 1-8. <https://dx.doi.org/10.17582/journal.pjz/20220308070343>
- Narinc D, Kamaran E and Aksoy T (2014). Effects of slaughter age and mass selection on slaughter and carcass characteristics in 2 lines of Japanese quail. *Poultry Science*, 93: 762-769. <http://dx.doi.org/10.3382/ps.2013-03506>
- National Research Council (1994). Nutrient requirements of poultry. 9th rev. ed. National Academy of Sciences, Washington, D.C. [https://books.google.co.bw/books?hl=en&lr=&id=bbV1FUqRcMOC&oi=fnd&pg=PT13&dq=National+Research+Council+\(1994\).&ots=llhs2FhuQA&sig=E44-d87lDtlM5GGF1hdgZt5ar7A&redir_esc=y#v=onepage&q=National%20Research%20Council%20\(1994\).&f=false](https://books.google.co.bw/books?hl=en&lr=&id=bbV1FUqRcMOC&oi=fnd&pg=PT13&dq=National+Research+Council+(1994).&ots=llhs2FhuQA&sig=E44-d87lDtlM5GGF1hdgZt5ar7A&redir_esc=y#v=onepage&q=National%20Research%20Council%20(1994).&f=false)
- Nikolakakis I, Banakis D, Florou-Paneri P, Dotas V, Giannenas I and Botsoglou N (2004). Effect of dried tomato pulp on performance and carcass characteristics of growing quails. *Archiv fur Geflugelkunde*, 68(1): 34-38. Available at: <https://www.european-poultry-science.com/effect-of-dried-tomato-pulp-on-performance-and-carcass-characteristics-of-growing-quails.QUIEPTQ5NjE2NjYmTUJEPTE2MTAxCZQQUdFX1RQTD1QcmIudHBzZXZpZXcuaHRUk1FVEFfUk9CT1Q9TOZG.html>
- Okello AO, Otieno DJ, Nzuma JM, Kidoido MM and Tanga CM (2023). Smallholder farmers' willingness to pay for commercial insect-based chicken feed in Kenya. *International Food and Agribusiness Management Review*, 26(1):67-87. <https://doi.org/10.22434/IFAMR2022.0047>
- Olawumi SO (2015). Carcass characteristics of Coturnix quail as affected by sex and housing system. *International Journal of Agriculture, Forestry and Fisheries*, 3(3): 76-79. <http://www.openscienceonline.com/journal/archive2?journalId=706&paperId=1652>
- Ophir AG, Persaud KN and Galef BG J (2005). Avoidance of relatively aggressive male Japanese quail (*Coturnix coturnix japonica*) by sexually experienced conspecific females. *Journal of Comparative Psychology*, 119(1): 3-7. <https://psycnet.apa.org/doi/10.1037/0735-7036.119.1.3>
- Rahman ANMA, Hoque MN, Talukder AK and Das ZC (2016). A survey of Japanese quail (*Coturnix coturnix japonica*) farming in selected areas of Bangladesh. *Veterinary World*, 9(9): 940-947. <https://doi.org/10.14202/vetworld.2016.940-947>
- Ratriyanto A, Indreswari R and Nuhriawangsa AMP (2017). Effects of dietary protein level and betaine supplementation on nutrient digestibility and performance of Japanese quails. *Revista Brasileira de Ciéncia Avícola*, 19: 445-454. <https://doi.org/10.1590/1806-9061-2016-0442>
- Rezaei M, Torshizi MAK, Wall H and Ivarsson E (2018). Body growth, intestinal morphology and microflora of quail on diets supplemented with micronised wheat fiber. *British Poultry Science*, 59: 422-429. <https://doi.org/10.1080/00071668.2018.1460461>
- Sahin N, Orhan C, Tuzcu M, Sahin K and Kucuk O (2008). The effects of tomato powder supplementation on performance and lipid peroxidation in quail. *Poultry Science*, 87: 276-283. <https://doi.org/10.3382/ps.2007-00207>
- Sezer M, Berberoglu E and Ulutas Z (2006). Genetic association between sexual maturity and weekly live-weights in laying type Japanese quail. *South African Journal of Animal Science*, 36(2): 142-148. <https://doi.org/10.4314/sajas.v36i2.3997>
- Shehata SF, Kamel ER, Abo-Salem MES and Atallah ST (2018). Effect of some dietary supplementation on economic efficiency of growing Japanese Quails. *Benha Veterinary Medicine Journal*, 34: 219-231. <https://dx.doi.org/10.21608/bvmj.2018.54243>
- Siamabele B (2021). The significance of soybean production in the face of changing climates in Africa. *Cogent Food & Agriculture*, 7: 1933745. <https://doi.org/10.1080/23311932.2021.1933745>
- Statistical Analysis Systems (2012). SAS 9.4. Qualification tools user's guide. SAS Institute Inc: Cary N.C. https://www.sas.com/en_us/home.html
- Szabo K, Dulf FV, Diaconeasa Z and Vodnar DC (2019). Antimicrobial and antioxidant properties of tomato processing byproducts and their correlation with the biochemical composition. *LWT-Food Science and Technology*, 116: 108558. <https://doi.org/10.1016/j.lwt.2019.108558>
- Tabeidian SA, Toghiani M, Toghiani M and Shahidpour A (2011). Effect of incremental levels of dried tomato pomace with and without dietary enzyme supplementation on growth performance, carcass traits and ileal protein digestibility of broiler chicks. *Journal of Animal and Veterinary Advance*, 10: 443-448. <https://www.cabidigitallibrary.org/doi/full/10.5555/20113076649>
- Tamburawa MS, Zango MH, Khaleel AG and Makinde OJ (2018). Response of finisher broilers fed toasted cotton seed cake meal-based diets. *Wayamba Journal of Animal Science*, 10: 1641-1647. [Google Scholar](https://doi.org/10.1080/23311932.2018.1533745)
- Tejeda OJ and Kim WK (2021). Role of dietary fiber in poultry nutrition. *Animals*, 11: 461. <https://doi.org/10.3390/ani11020461>
- Yitbarek MB (2013). Effect of feeding different levels of dried tomato pomace on the performance of Rhode Island Red grower chicks in Wolaita Zone, Southern Ethiopia. *Asian Journal of Poultry Science*, 7(1): 27-33. <https://doi.org/10.3923/ajpsaj.2013.27.33>
- Young LL, Northcutt JK, Buhr RJ, Lyon CE and Ware GO (2001). Effects of age, sex, and duration of post-mortem aging on percentage yield of parts from broiler chicken carcasses. *Poultry Science*, 80: 376-379. <https://doi.org/10.1093/ps/80.3.376>

Publisher's note: Scienceline Publication Ltd. remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access: This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If the material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <https://creativecommons.org/licenses/by/4.0/>.

Instructions for Authors

[OJAFR EndNote Style](#)  | [Word Template](#)  | [Declaration form](#)  | [Authorship Agreement Form](#) 

Manuscripts as Original Research Paper, Review, Short Communication and Case Reports are invited for peer-review publishing in the *Online Journal of Animal and Feed Research (ISSN 2228-7701)*.

Papers can be in any relevant fields of Animal Sciences (Animal Nutrition, Physiology, Reproduction, Genetics and Breeding, Behavior, Health, Husbandry and its economy, Animal products and Veterinary medicines of domestic animals) and relative topics. The journal does encourage papers with emphasis on the nutritive value and utilization of feeds that is depended to methods of Improvement, Assessment, Conserving and Processing feeds, Agronomic and climatic factors, Metabolic, Production, Reproduction and Health responses to dietary inputs (e.g., Feeds, Feed Additives, Specific Feed Components, Mycotoxins). Also, Mathematical models relating directly to animal-feed interactions, Analytical and experimental methods for Feed Evaluation as well as Animal Production studies with a focus on Animal Nutrition that do have link to a feed (Food Science and Technology) are acceptable relative topics for OJAFR. [...view full aims and scope](#)

Submission

The manuscripts should be submitted using our [online](#) submission forms ([Scienceline Online Submission Form](#) ; [OJAFR Online Submission Form](#) ). For facile submission, please embed all figures and tables at the end of the manuscript to become one single file for submission. Once submission is complete, the system will generate a manuscript ID and password sent to the author's contact email. If you have any difficulty in submitting the manuscript, kindly send via emails: editors@ojafr.com ; editorojafr@gmail.com. All manuscripts must be checked (by an English native speaker) and submitted in English for evaluation in a totally confidential and impartial way.

Supplementary information:

Author guidelines are specific for each journal. Our MS Word template can assist you by modifying your page layout, text formatting, headings, title page, image placement, and citations/references such that they agree with the guidelines of the journal. If you believe your article is fully edited per journal style, please use our [Word template](#) before submission. Supplementary materials may include figures, tables, methods, videos, and other materials. They are available online linked to the original published article. Supplementary tables and figures should be labeled with a "S", e.g. "Table S1" and "Figure S1". The maximum file size for supplementary materials is 10MB each. Please keep the files as small as possible to avoid the frustrations experienced by readers with downloading large files.

Submission to the Journal is on the understanding that:

1. The article has not been previously published in any other form and is not under consideration for publication elsewhere;
2. All authors have approved the submission and have obtained permission for publishing work.
3. Researchers have proper regard for conservation and animal welfare considerations. Attention is drawn to the '[Guidelines for the Treatment of Animals in Research and Teaching](#)'. Any possible adverse consequences of the work for populations or individual organisms must be weighed against the possible gains in knowledge and its practical applications. If the approval of an ethics committee is required, please provide the name of the committee and the approval number obtained.

Ethics declarations

If experimental research includes animal subjects (involving live vertebrates and/or higher invertebrates), the authors will need to include one of the following appropriate ethics declarations in the Methods section of manuscript.

1. A statement that identifies the institutional and/or licensing committee that approved the experiments, including any relevant details (e.g. the board/committee names that gave the approval).
2. The authors confirm that all experiments were performed in accordance with relevant named guidelines and regulations.
3. A statement confirms that the authors complied with the ARRIVE guidelines and or the Interdisciplinary Principles and Guidelines for the Use of Animals in Research, Testing, and Education by the New York Academy of Sciences, Ad Hoc Animal Research Committee.

If the manuscript contains photos or parts of photos of patients, informed consent from each patient should be obtained. Patient's identities and privacy should be carefully protected in the manuscript.

Presentation of the article

Main Format

First page of the manuscripts must be properly identified by the title and the name(s) of the author(s). It should be typed in Times New Roman (font sizes: 12pt in capitalization for the title and the main text, double spaced, in A4 format with 2cm margins. All pages and lines of the main text should be numbered consecutively throughout the manuscript. The manuscript must be saved in a .doc or .docx formats. Abbreviations in the article title are not allowed except the well-known ones.

Manuscripts should be arranged in the following order:

- a. TITLE (brief, attractive and targeted)
- b. Name(s) and Affiliation(s) of author(s) (including postcode) and corresponding Email
- c. ABSTRACT
- d. Key words (separate by semicolons; or comma,)
- e. Abbreviations (used in the manuscript)
- f. INTRODUCTION
- g. MATERIALS AND METHODS
- h. RESULTS
- i. DISCUSSION
- j. CONCLUSION

The sections "RESULTS AND DISCUSSION" can be presented jointly.

The sections "DISCUSSION AND CONCLUSION" can be presented jointly.

k. DECLARATIONS

l. REFERENCES

m. Tables

n. Figures

Article Sections Format

Title should be a brief phrase describing the contents of the paper. Title Page should include full names and affiliations of the author(s), the name of the corresponding author along with phone and email information. Present address(es) of the author(s) should appear as a footnote.

Abstract should be informative and completely self-explanatory, briefly present the topic, state the scope of the experiments, indicate significant data, and point out major findings and conclusions. The abstract should be 150 to 300 words in length. Complete sentences, active verbs, and the third person should be used, and the abstract should be written in the past tense. Standard nomenclature should be used and abbreviations should be avoided. No literature should be cited.

Following the abstract, about 3 to 7 **key words** should be listed.

Introduction should provide a clear statement of the problem, the relevant literature on the subject, and the proposed approach or solution. It should be understandable to colleagues from a broad range of scientific disciplines.

Materials and Methods should be complete enough to allow experiments to be reproduced. However, only truly new procedures should be described in detail; previously published procedures should be cited, and important modifications of published procedures should be mentioned briefly. Capitalize trade names and include the manufacturer's name and address. Subheadings should be used. Methods in general use need not be described in detail.

Results should be presented with clarity and precision. The results should be written in the past tense when describing findings in the author(s)'s experiments. Previously published findings should be written in the present tense. Results should be explained, but largely without referring to the literature. Discussion, speculation and detailed interpretation of data should not be included in the results but should be put into the discussion section.

Discussion should interpret the findings in view of the results obtained in this and in past studies on this topic. State the conclusions in a few sentences at the end of the paper. The Results and Discussion sections can include subheadings, and when appropriate, both sections can be combined.

Conclusion should be brief and tight, providing a few specific tasks to accomplish: 1-Re-assert/Reinforce the Thesis; 2-Review the Main Points; 3-Close Effectively. The Conclusion section should not be similar to the Abstract content.

Declarations including Ethics, Consent to publish, Competing interests, Authors' contributions, and Availability of data and materials are necessary.

Acknowledgments of persons, grants, funds, etc should be brief.

Tables should be kept to a minimum and be designed to be as simple as possible. Tables are to be typed double-spaced throughout, including headings and footnotes. Each table should be on a separate page, numbered consecutively in Arabic numerals and supplied with a heading and a legend. Tables should be self-explanatory without reference to the text. The details of the methods used in the experiments should preferably be described in the legend instead of in the text. The same data should not be presented in both table and graph forms or repeated in the text.

The Figure legends should be typed in numerical order on a separate sheet. Graphics should be prepared using applications capable of generating high resolution GIF, TIFF, JPEG or PowerPoint before pasting in the Microsoft Word manuscript file. Use Arabic numerals to designate figures and upper case letters for their parts (Figure 1). Begin each legend with a title and include sufficient description so that the figure is understandable without reading the text of the manuscript. Information given in legends should not be repeated in the text.

DECLARATIONS

Please ensure that the sections: Ethics (and consent to participate), Consent to publish, Competing interests, Authors' contributions, and Availability of data and materials are included at the end of your manuscript in a Declarations section.

Consent to Publish

Please include a 'Consent for publication' section in your manuscript. If your manuscript contains any individual person's data in any form (including individual details, images or videos), consent to publish must be obtained from that person, or in the case of children, their parent or legal guardian. All presentations of case reports must have consent to publish. You can use your institutional consent form or our consent form if you prefer. You should not send the form to us on submission, but we may request to see a copy at any stage (including after publication). If your manuscript does not contain any individual person's data, please state "Not applicable" in this section.

Authors' Contributions

For manuscripts with more than one author, OJAFR requires an Authors' Contributions section to be placed after the Competing Interests section. An 'author' is generally considered to be someone who has made substantive intellectual contributions to a published study. To qualify as an author one should 1) have made substantial contributions to conception and design, or acquisition of data, or analysis and interpretation of data; 2) have been involved in drafting the manuscript or revising it critically for important intellectual content; and 3) have given final approval of the version to be published. Each author should have participated sufficiently in the work to take public responsibility for appropriate portions of the content. Acquisition of funding, collection of data, or general supervision of the research group, alone, does not justify authorship. We suggest the following format (please use initials to refer to each author's contribution): AB carried out the molecular genetic studies, participated in the sequence alignment and drafted the manuscript. JY carried out the immunoassays. MT participated in the sequence alignment. ES participated in the design of the study and performed the statistical analysis. FG conceived of the study, and participated in its design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript. For authors that equally participated in a study please write '**All/Both authors contributed equally to this work.**' Contributors who do not meet the criteria for authorship should be listed in an acknowledgements section.

Competing Interests

Competing interests that might interfere with the objective presentation of the research findings contained in the manuscript should be declared in a paragraph heading "Competing interests" (after Acknowledgement section and before References). Examples of competing interests are ownership of stock in a company, commercial grants, board membership, etc. If there is no competing interest, please use the statement "The authors declare that they have no competing interests." *Online Journal of Animal and Feed Research* adheres to the definition of authorship set up by the International Committee of Medical Journal Editors (ICMJE). According to the ICMJE authorship criteria should be based on 1) substantial contributions to

conception and design of, or acquisition of data or analysis and interpretation of data, 2) drafting the article or revising it critically for important intellectual content and 3) final approval of the version to be published. Authors should meet conditions 1, 2 and 3. It is a requirement that all authors have been accredited as appropriate upon submission of the manuscript. Contributors who do not qualify as authors should be mentioned under Acknowledgements.

Change in authorship

We do not allow any change in authorship after provisional acceptance. We cannot allow any addition, deletion or change in the sequence of author names. We have this policy to prevent fraud.

Acknowledgements

We strongly encourage you to include an Acknowledgements section between the Authors' contributions section and Reference list. Please acknowledge anyone who contributed towards the study by making substantial contributions to conception, design, acquisition of data, or analysis and interpretation of data, or who was involved in drafting the manuscript or revising it critically for important intellectual content, but who does not meet the criteria for authorship. Please also include their source(s) of funding. Please also acknowledge anyone who contributed materials essential for the study. Authors should obtain permission to acknowledge from all those mentioned in the Acknowledgements. Please list the source(s) of funding for the study, for each author, and for the manuscript preparation in the acknowledgements section. Authors must describe the role of the funding body, if any, in study design; in the collection, analysis, and interpretation of data; in the writing of the manuscript; and in the decision to submit the manuscript for publication.

Data deposition

Nucleic acid sequences, protein sequences, and atomic coordinates should be deposited in an appropriate database in time for the accession number to be included in the published article. In computational studies where the sequence information is unacceptable for inclusion in databases because of lack of experimental validation, the sequences must be published as an additional file with the article.

REFERENCES

OJAFR initially accepts the manuscripts in PDF, Word or TeX/LaTeX formats; Word files are preferred, especially those prepared using [EndNote®](#). However, our team will reformat the articles of non-EndNote users via EndNote in Galley proof stage, if accepted.

An OJAFR reference style for [EndNote](#) may be found [here](#).

How to install additional styles? Please [click here](#)

How to turn on "Jumping" from a citation to the bibliography? Please [click here](#)

1. All references to publications made in the text should be presented in a list with their full bibliographical description.
2. In the text, a reference identified by means of an author's name should be followed by the date of the reference in parentheses. When there are more than two authors, only the first author's surname should be mentioned, followed by 'et al'. In the event that an author cited has had two or more works published during the same year, the reference, both in the text and in the reference list, should be identified by a lowercase letter like 'a' and 'b' after the date to distinguish the works.
3. References in the text should be arranged chronologically (e.g. Kelebeni, 1983; Usman and Smith, 1992 and Agindotan et al., 2003). 'et al.' should not be italic. The list of references should be arranged alphabetically on author's surnames, and chronologically per author. If an author's name in the list is also mentioned with co-authors, the following order should be used: Publications of the single author, arranged according to publication dates - publications of the same author with one co-author - publications of the author with more than one co-author. Publications by the same author(s) in the same year should be listed as 1992a, 1992b, etc.
4. Names of authors and titles of journals published in non-latin alphabets should be transliterated in English.
5. A sample of standard reference is "1st Author surname A, 2nd Author surname B and 3rd Author surname C (2013). Article title should be regular, in sentence case form, and 9 pt. Online Journal of Animal and Feed Research, Volume No. (Issue No.): 00-00." (Journal titles should be full and not italic.)
6. If available please add DOI numbers or the link of articles at the end of each reference.

Examples (at the text)

Abayomi (2000), Agindotan et al. (2003), (Kelebeni, 1983), (Usman and Smith, 1992), (Chege, 1998; Chukwura, 1987a,b; Tijani, 1993,1995), (Kumasi et al., 2001).

Examples (at references section)

a) For journal

Graulet B (2014). Ruminant milk: A source of vitamins in human nutrition. *Animal Frontiers*, 4(2):24-30. [Link](#), [DOI](#)

Miller BA and Lu CD (2019). Current status of global dairy goat production: An overview. *Asian-Australasian Journal of Animal Sciences*, 32(8): 1219. [Link](#), [DOI](#)

Xu P, Zhang Z, Peng P, Yang J, Li X, Yuan T, et al. (2022). Study on vacuum drying kinetics and processing of the *Lonicera japonica* Thunb. aqueous extracts. *LWT - Food Science and Technology*. 167: 1-9. [Link](#), [DOI](#)

b) For symposia reports and abstracts

Cruz EM, Almatar S, Aludul EK and Al-Yaqout A (2000). Preliminary Studies on the Performance and Feeding Behaviour of Silver Pomfret (*Pampus argentens euphrasens*) Fingerlings fed with Commercial Feed and Reared in Fibreglass Tanks. *Asian Fisheries Society Manila, Philippine*, 13: 191-199. [Link](#), [DOI](#)

c) For edited symposia, special issues, etc., published in a journal

Korevaar H (1992). The nitrogen balance on intensive Dutch dairy farms: a review. In: A. A. Jongebreur et al. (Editors), *Effects of Cattle and Pig Production Systems on the Environment: Livestock Production Science*, 31: 17-27. [Link](#), [DOI](#)

d) For books

AOAC (1990). Association of Official Analytical Chemists. *Official Methods of Analysis*, 15th Edition. Washington D.C. pp. 69-88. [Link](#), [DOI](#)

Pelczar JR, Harley JP, Klein DA (1993). *Microbiology: Concepts and Applications*. McGraw-Hill Inc., New York, pp. 591-603. [Link](#), [DOI](#)

e) Books, containing sections written by different authors

Kunev M (1979). Pig Fattening. In: A. Alexiev (Editor), Farm Animal Feeding. Vol. III. Feeding of Different Animal Species, Zemizdat, Sofia, p. 233-243 (Bg). [Link](#), [DOI](#)

In referring to a personal communication the two words are followed by the year, e.g. (Brown, J. M., personal communication, 1982). In this case initials are given in the text. Where available, URLs for the references should be provided.

Formulae, numbers and symbols

1. Typewritten formulae are preferred. Subscripts and superscripts are important. Check disparities between zero (0) and the letter O (0 vs. O), and between one (1) and the letter I (1 vs. I).
2. Describe all symbols immediately after the equation in which they are first used.
3. For simple fractions, use the solidus (/), e.g. 10 /38.
4. Equations should be presented into parentheses on the right-hand side, in tandem.
5. Levels of statistical significance which can be used without further explanations are *P <0.05, **P <0.01, and ***P <0.001.
6. In the English articles, a decimal point should be used instead of a decimal comma.
7. Use Symbol fonts for "±"; "≤" and "≥" (avoid underline).
8. In chemical formulae, valence of ions should be given, e.g. Ca²⁺ and CO₃²⁻, not as Ca⁺⁺ or CO₃.
9. Numbers up to 10 should be written in the text by words. Numbers above 1000 are recommended to be given as 10 powered x.
10. Greek letters should be explained in the margins with their names as follows: Αα - alpha, Ββ - beta, Γγ - gamma, Δδ - delta, Εε - epsilon, Ζζ - zeta, Ηη - eta, Θθ - theta, Ιι - iota, Κκ - kappa, Λλ - lambda, Μμ - mu, Νν - nu, Ξξ - xi, Οο - omicron, Ππ - pi, Ρρ - rho, Σσ - sigma, Ττ - tau, Υυ - ipsilon, Φφ - phi, Χχ - chi, Ψψ - psi, Ωω - omega. Please avoid using math equations in Word whenever possible, as they have to be replaced by images in xml full text.

Abbreviations

Abbreviations should be presented in one paragraph, in the format: "term: definition". Please separate the items by ";".

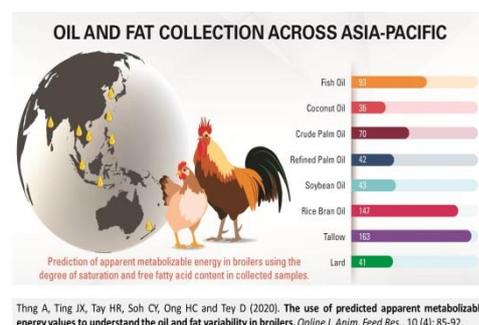
E.g. ANN: artificial neural network; CFS: closed form solution;

Graphical Abstract

Authors of accepted articles should provide a graphical abstract (a beautifully designed feature figure) to represent the paper aiming to catch the attention and interest of readers. Graphical abstract will be published online in the table of content. The graphical abstract should be colored, and kept within an area of 12 cm (width) × 6 cm (height) or with similar format. Image should have a minimum resolution of 300 dpi and line art 1200dpi.

Note: Height of the image should be no more than the width. Please avoid putting too much information into the graphical abstract as it occupies only a small space. Authors can provide the graphical abstract in the format of PDF, Word, PowerPoint, jpg, or png, after a manuscript is accepted for publication.

If you have decided to provide a Professional Graphical Abstract, please click [here](#).



Review/Decisions/Processing

Firstly, all manuscripts will be checked by one of the plagiarism finding tools ([iThenticate](#) and or [Turnitin](#)). A double-blind reviewing model is used by OJAFR for non-plagiarized papers. The manuscript is edited by the English language editor and checked by at least 2 reviewers at least 2 reviewers who are not part of the journal's editorial staff and mostly suggested by section editors. Manuscripts that are judged to be of insufficient quality or unlikely to be competitive enough for publication are returned to the authors at the initial stage.

We always try to avoid delays in the reviewing process, but it relies on the time and cooperation of the referees that works without any remuneration, hence, it may take 2 weeks to 2 months. One unfavorable review means that the paper will not be published and possible decisions are: accept as is, minor revision, major revision, or reject. The corresponding authors should submit back their revisions within 14 days in the case of minor revision, or 30 days in the case of major revision.

To submit a revision please click [here](#), fill out the form, and mark Revised, mention the article code (for example OJAFR-1108), attach the revision (MS word) and continue submission. Manuscripts with significant results are typically reviewed and published at the highest priority. After review and editing the article, a final formatted proof is sent to the corresponding author once again to apply all suggested corrections during the article process. The editor who received the final revisions from the corresponding authors shall not be held responsible for any mistakes shown in the final publication. Manuscripts with significant results are typically reviewed and published at the highest priority.

Language editing

No paper will be rejected for poor language. However, if you would like assistance with writing your manuscript, you can consider asking colleagues for their input and/or use a professional editing service such as those provided by our affiliates [American Journal Experts \(USA\)](#) and or [London Proofreaders \(UK\)](#). In addition, we may offer a Scienceline service (English editing, additional scientific editing, and translation) in a modest fee, for those articles that are in the revision stage, upon request. For more information please visit [here](#). The use of a language editing service has no bearing on editorial decisions and is not a requirement for publication.

Plagiarism: There is an instant policy towards plagiarism (including self-plagiarism) in our journals. Manuscripts (main text not including references list and title page) are screened for plagiarism by [iThenticate](#) and or [Turnitin](#) with default sensitivity before or during publication, and if found they will be rejected at any stage of processing.

Declaration: After the manuscript is accepted for publication, a [declaration form](#) will be sent to the corresponding author who is responsible for coauthors' agreements to publication of submitted work in OJAFR after any amendments arising from the peer review. All the authors should also approve any change in authorship (i.e., adding, removing or reordering existing authors) after initial submission. Authors should determine the order of authorship among themselves. In addition, any alterations must be

clarified to the Editor/Editor-in-chief via the [Authorship Agreement Form](#). For more information please read [Authorship and Authors' Responsibilities](#).

Date of issue

All accepted articles are published bimonthly around 25th of January, March, May, July, September and November, each year in full text on the Internet.

Publication charges

The publication costs are covered through article processing charges (APCs) and No submission fee, or any other processing fees are required for the publication of the accepted article. There is a modest APC of 180 Euro(€) editor fee for the processing of each primary accepted paper (1000-4000 words) to encourage high-quality submissions. APCs are only charged for articles that pass the pre-publication checks and are ready to be published. A surcharge will be placed on any article that is over 4000 words in length to cover the additional processing costs. We encourage the authors to submit manuscripts with no more than 4000 words (not including Abstract, Methods, References and figure legends). Payment can be made by credit card, bank transfer, money order or check. Instruction for payment is sent during the publication process as soon as the manuscript is accepted. Meanwhile, this journal encourages the academic institutions in low-income countries to publish high quality scientific results, free of charge.

WORD COUNT	PRICE*
1000-4000 words (medium article)	€180
over 4000 words (long article)	€280

* The prices are valid until 30th December 2024.

The Waiver policy

The submission fee will be waived for invited authors, authors of hot papers, and corresponding authors who are editorial board members of the *Online Journal of Animal and Feed Research*. The Journal will consider requests to waive the fee for cases of financial hardship (for high quality manuscripts and upon acceptance for publication). Requests for waiver of the submission fee must be submitted via individual cover letter by the corresponding author and cosigned by an appropriate institutional official to verify that no institutional or grant funds are available for the payment of the fee. Letters including the manuscript title and manuscript ID number should be sent to editors@ojafr.com. It is expected that waiver requests will be processed and authors will be notified within two business day.

The OA policy

Online Journal of Animal and Feed Research is an Open Access journal which means that all content is freely available without charge to the user or his/her institution. Users are allowed to read, download, copy, distribute, print, search, or link to the full texts of the articles, or use them for any other lawful purpose, without asking prior permission from the publisher or the author. This is in accordance with the [BOAI definition of Open Access](#).

Scienceline Language Editing Services

We suggest that authors whose first language is not English have their manuscripts checked by a native English speaker before submission. This is optional, but will help to ensure that any submissions that reach peer review can be judged exclusively on academic merit. We offer a Scienceline service, and suggest that authors contact as appropriate. Please note that use of language editing services is voluntary, and at the author's own expense. Use of these services does not guarantee that the manuscript will be accepted for publication, nor does it restrict the author to submitting to Scienceline journals. You can send the article/s to the following Emails: administrator@science-line.com ; info@science-line.com

For more information about editing services please visit [here](#).

Submission Preparation Checklist

Authors are required to check off their submission's compliance with all of the following items, and submissions may be returned to authors that do not adhere to the following guidelines:

- The submission has not been previously published, nor is it before another journal for consideration (or an explanation has been provided in -Comments to the Editor).
- The submission file is in Microsoft Word, RTF, or PDF document file format.
- Where available, URLs for the references have been provided.
- The text is double-spaced; uses a 12-point font; and all illustrations, figures, and tables are placed within the text at the appropriate points, rather than at the end.
- The text adheres to the stylistic and bibliographic requirements outlined in the Author Guidelines.



[ABOUT US](#) | [CONTACT US](#) | [PRIVACY POLICY](#)

Scienceline Publication, Ltd.

Ömer Nasuhi Bilmen Road, Dönmez Apart., G Block, No: 1/6, Yakutiye, Erzurum/25100, TURKEY

Phone: +90 538 770 8824 (TURKEY)

Homepage: www.science-line.com

Emails: administrator@science-line.com ; saeid.azar@atauni.edu.tr

SCIENCELINE PUBLISHING CORPORATION

Scienceline Publication Ltd. is a limited liability non-profit non-stock corporation incorporated in Turkey (Company No. 0757086921600001). Scienceline journals that concurrently belong to many societies, universities and research institutes, publishes internationally peer-reviewed open access articles and believe in sharing of new scientific knowledge and vital research in the fields of life and natural sciences, animal sciences, engineering, art, linguistic, management, social and economic sciences all over the world. Scienceline journals include:

Online Journal of Animal and Feed Research



ISSN 2228-7701; Bi-monthly
View Journal | Editorial Board
Email: editors@ojafir.ir
Submit Online >>

Journal of Civil Engineering and Urbanism



ISSN 2252-0430; Bi-monthly
View Journal | Editorial Board
Email: ojceu@ojceu.ir
Submit Online >>

Journal of Life Sciences and Biomedicine



ISSN: 2251-9939; Bi-monthly
View Journal | Editorial Board
Email: editors@jlsb.science-line.com
Submit Online >>

Asian Journal of Medical and Pharmaceutical Researches



ISSN: 2322-4789; Quarterly
View Journal | Editorial Board
Email: editor@ajmpr.science-line.com
Submit Online >>

Journal of World's Poultry Research



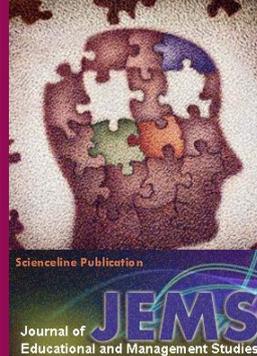
ISSN: 2322-455X; Quarterly
View Journal | Editorial Board
Email: editor@jwpr.science-line.com
Submit Online >>

World's Veterinary Journal



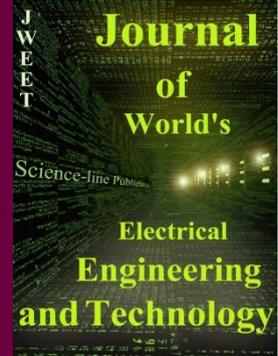
ISSN: 2322-4568; Quarterly
View Journal | Editorial Board
Email: editor@wj.science-line.com
Submit Online >>

Journal of Educational and Management Studies



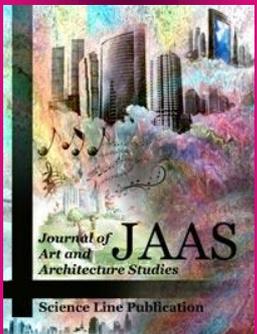
ISSN: 2322-4770; Quarterly
View Journal | Editorial Board
Email: info@jems.science-line.com
Submit Online >>

Journal of World's Electrical Engineering and Technology



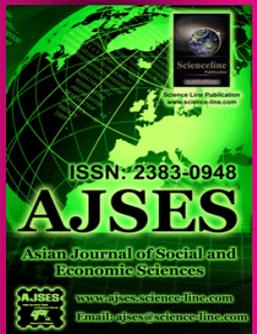
ISSN: 2322-5114; Irregular
View Journal | Editorial Board
Email: editor@jwee.science-line.com
Submit Online >>

Journal of Art and Architecture Studies



ISSN: 2383-1553; Irregular
View Journal | Editorial Board
Email: jaas@science-line.com
Submit Online >>

Asian Journal of Social and Economic Sciences



ISSN: 2383-0948; Quarterly
View Journal | Editorial Board
Email: ajses@science-line.com
Submit Online >>

Journal of Applied Business and Finance Researches



ISSN: 2382-9907; Quarterly
View Journal | Editorial Board
Email: jabfr@science-line.com
Submit Online >>

Scientific Journal of Mechanical and Industrial Engineering



ISSN: 2383-0980; Quarterly
View Journal | Editorial Board
Email: sjmie@science-line.com
Submit Online >>

ABOUT
LEADERSHIP
AIMS AND SCOPE
PUBLISHING ETHICS
POLICIES
TERMS AND CONDITIONS
CONTACT US

Scienceline is a non-profit organisation inspired by research funders and led by scholars. Our mission is to help researchers accelerate discovery and innovation by operating a platform for research communication that encourages and recognises the most responsible behaviours in science.

Scienceline Publications, Ltd is a limited liability non-profit non-stock corporation registered in the State of Erzurum, Turkey, with company number 0757086921600001, and branch number 18677/25379 at the address: Scienceline Publications, Ltd., Ömer Nasuhi Bilmen Road, Dönmez Apart., G1/6, Yakutiye, Erzurum 25100, Turkey